Review of: "Non-Darwinian Molecular Biology"

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Invited Commentary on "Non-Darwinian Molecular Biology," Alexander F. Palazzo* and Nevraj S. Kejiou, Front. Genet., 16 February 2022 | https://doi.org/10.3389/fgene.2022.831068 peerreview@reach.geios.com

When I first saw the title of this review article, I was interested to write a commentary because I thought the review would highlight many of the exciting new topics in evolutionary genomics and molecular biology. When I read the article, however, I was disappointed to find that Drs. Palazzo and Kejiou denigrated important contemporary research and preferred an out-of-date view of evolution producing "messy" genomes as well as embracing the long discredited notion of "junk DNA.

Molecular Biology and genomics have made it possible to study evolutionary processes with unprecedented precision and revealed the major roles played by important molecular agents, such as transposable elements and (protein) noncoding DNA and RNA sequences. The authors have chosen to ignore the growing body of evidence revealing how these unexpected players in genome evolution have contributed to the origins of adaptive innovations and new taxa, especially in the most complex eukaryotes. Their concluding summary makes this retrograde point of view quite clear:

"Most molecular biologists use an antiquated model of how evolution shapes biological processes leading them to an unrealistic hyper-adaptationalist view. A prime example of this is the interpretation of the ENCODE project results. Ultimately, this ultra-Darwinian mindset perpetuates the notion that the genome, and life itself, is like a Swiss watch—ornate, and complicated, with every part hand crafted for a specific purpose. This view is completely compatible with the idea that genome is pure information. However, this view is based on ignorance of developments in molecular evolution. It also ignores principles of biochemistry, that predict suboptimal reactions and widespread promiscuity. A more modern view of the eukaryotic cell, shaped by drift-dominated evolution, is a messy junk-filled entity, full of Rube-Goldberg contraptions that were hobbled together by non-adaptive forces. With this new vantage point, certain aspects of eukaryotic biology become clarified, including the evolution of complexity."

Palazzo and Kejiou's concept of non-Darwinian evolution is to downplay Darwin's

focus on the importance of adaptive novelty as reflecting "an unrealistic hyper-adaptationalist view," and they choose research on noncoding RNA functionalities stimulated by the ENCODE (Encyclopedia of DNA Elements) project as the chief example of what they characterize as a mistaken "hyper-adaptationalist" focus, disregarding the fact that noncoding RNA research is currently one of the hottest topics in molecular cell biology and genetics (see below on PubMed results). Instead, they prefer to focus on a notion of genomic and cellular "sloppiness" predicted by neutral evolution theories and molecular buffering processes.

Molecular Biology, and more particularly the field of genomics, are indeed Non-

Darwinian for a very good reason. But it is not the anti-adaptationist argument of this review. Non-Darwinian evolutionary theory arises because molecular genetics and genomics have provided us with new ways of thinking about genome content, genome functioning, and evolutionary genome change. Protein-coding DNA constitutes less than 2% of our own genomes (de Koning et al., 2011), and an early comparative genome analysis found that noncoding DNA content in a genome is a more accurate predictor of organismal complexity than protein-coding content (Liu et al., 2013).

Readers should consider how thinking about the basic biology and genetic principles underpinning evolution studies have changed in recent years when deciding what parts of the scientific literature to follow:

1. What Palazzo and Kejiou label as "messiness" is considered by disciplines such as Evo-Devo to reflect robust redundancy and complexity that ensure the exceptional reliability of inherited characteristics under varying conditions (Keane et al., 2014; Kim et al., 2014; Oliveira et al., 2014; Payne & Wagner, 2014; Plata & Vitkup, 2014; Ruz et al., 2014; Zheng & Triesch, 2014; Cui et al., 2015; Jung et al., 2015; Araujo & Liotta, 2018; diCenzo et al., 2018; Osterwalder et al., 2018).

2. Instead of discrete phenotypic traits defined by independently evolving genetic units, we now recognize that cellular and organismal properties result from integrated adaptive systems that depend on expression of coordinated genomic networks (e.g. animal body plan development) (Li et al., 2020; Nishihara, 2020; Panni et al., 2020; Chu et al., 2021; Dandage et al., 2021; Duran et al., 2021; Etxebeste, 2021; Hagolani et al., 2021; Mottes et al., 2021; Sharma et al., 2021).

3. Genome network evolution has been found to involve the ability of transposable

elements to spread appropriate expression signals to multiple different genetic loci and so integrate them into coordinated systems (Jacques et al., 2013; Johnson, 2017; Trizzino et al., 2017; Morata et al., 2018; Rishishwar et al., 2018; Sundaram & Wang, 2018; Trizzino et al., 2018; Fawcett & Innan, 2019; Baud et al., 2020; Moschetti et al., 2020; Nishihara, 2020; Qiu & K hler, 2020; Voronova et al., 2020; Almojil et al., 2021; Nicolau et al., 2021; Senft & Macfarlan, 2021; Zhang et al., 2021) (https://shapiro.bsd.uchicago.edu/Distributed_genome_network_innovation_attributed _to_mobile_DNA_elements.html).

4. Many aspects of cell and organismal function have been documented by genetic analysis to require the presence of noncoding RNAs, which act as enzymes, sensors, and polyvalent aggregation centers for assembling multimolecular complexes in both the nucleus and cytoplasm. (Johnson & Guigo, 2014; Wang et al., 2016; Andergassen & Rinn, 2021; Borkiewicz et al., 2021; Oo et al., 2021; Statello et al., 2021) (Quinodoz & Guttman, 2014; Chujo et al., 2016; Chishima et al., 2018; Mattick, 2018; Salviano-Silva et al., 2018; Daneshvar et al., 2020). The functional diversity of noncoding RNAs is beginning to rival that of proteins, and these versatile molecules carry out some very high level and taxonomically restricted regulatory functions (https://shapiro.bsd.uchicago.edu/Regulatory_Functions_Reported_for_Long_Noncoding_IncRNA_molecules.html).

The fact is that so-called "junk DNA" elements (transposons, retrotransposons, repeats, noncoding sequences) are the genome components that change most during evolutionary transitions, and all have been well-documented to help generate major adaptive novelties. The importance of noncoding, transposable and repetitive DNA elements and of their RNA transcripts as research subjects is reflected in their publication numbers. Here are the numbers I found with different PubMed search entries:

noncoding RNA - 257,261 results long noncoding RNA - 38,262 results transposable element - 29,836 results repetitive DNA - 32,783 results noncoding DNA - 22,441 results chromosome structure - 116,574 results genome network - 84,415 results neutral evolution - 16,971 results While these numbers provide only a rough estimate of the research being done on a topic, it is clear that many molecular biologists and molecular evolutionists are producing discoveries and publishing their findings at a significant clip. To me, having studied and practiced molecular genetics, it would seem to be a poor choice to follow the advice of Palazzo and Kejiou and ignore all of this exciting and revelatory research.

REFERENCES

Almojil D, Bourgeois Y, Falis M, Hariyani I, Wilcox J & Boissinot S. (2021). The Structural, Functional and Evolutionary Impact of Transposable Elements in Eukaryotes. Genes (Basel) 12.

Andergassen D & Rinn JL. (2021). From genotype to phenotype: genetics of mammalian long non-coding RNAs in vivo. Nat Rev Genet.

Araujo RP & Liotta LA. (2018). The topological requirements for robust perfect adaptation in networks of any size. Nat Commun 9, 1757.

Baud A, Wan M, Nouaud D, Francillonne N, Anxolab h re D & Quesneville H. (2020).

Traces of transposable elements in genome dark matter coopted by flowering gene regulation networks BioRxiv.

Borkiewicz L, Kalafut J, Dudziak K, Przybyszewska-Podstawka A & Telejko I. (2021). Decoding LncRNAs. Cancers (Basel) 13.

Chishima T, Iwakiri J & Hamada M. (2018). Identification of Transposable Elements Contributing to Tissue-Specific Expression of Long Non-Coding RNAs. Genes (Basel) 9.

Chu T, Mouillet JF, Cao Z, Barak O, Ouyang Y & Sadovsky Y. (2021). RNA Network Interactions During Differentiation of Human Trophoblasts. Front Cell Dev Biol 9, 677981.

Chujo T, Yamazaki T & Hirose T. (2016). Architectural RNAs (arcRNAs): A class of long noncoding RNAs that function as the scaffold of nuclear bodies. Biochim Biophys Acta 1859, 139-146.

Cui H, Tsuda K & Parker JE. (2015). Effector-triggered immunity: from pathogen perception to robust defense. Annu Rev Plant Biol 66, 487-511.

Dandage R, Berger CM, Gagnon-Arsenault I, Moon KM, Stacey RG, Foster LJ & Landry

CR. (2021). Frequent Assembly of Chimeric Complexes in the Protein Interaction

Network of an Interspecies Yeast Hybrid. Mol Biol Evol 38, 1384-1401.

Daneshvar K, Ardehali MB, Klein IA, Hsieh FK, Kratkiewicz AJ, Mahpour A, Cancelliere

SOL, Zhou C, Cook BM, Li W, Pondick JV, Gupta SK, Moran SP, Young RA,

Kingston RE & Mullen AC. (2020). IncRNA DIGIT and BRD3 protein form phaseseparated

condensates to regulate endoderm differentiation. Nat Cell Biol 22, 1211-

1222.

de Koning AP, Gu W, Castoe TA, Batzer MA & Pollock DD. (2011). Repetitive elements may comprise over two-thirds of the human genome. PLoS Genet 7, e1002384. diCenzo GC, Benedict AB, Fondi M, Walker GC, Finan TM, Mengoni A & Griffitts JS. (2018). Robustness encoded across essential and accessory replicons of the ecologically versatile bacterium Sinorhizobium meliloti. PLoS Genet 14, e1007357. Duran BOS, Garcia de la Serrana D, Zanella BTT, Perez ES, Mareco EA, Santos VB, Carvalho RF & Dal-Pai-Silva M. (2021). An insight on the impact of teleost whole genome duplication on the regulation of the molecular networks controlling skeletal muscle growth. PLoS One 16, e0255006. Etxebeste O. (2021). Transcription Factors in the Fungus Aspergillus nidulans: Markers of

Genetic Innovation, Network Rewiring and Conflict between Genomics and Transcriptomics. J Fungi (Basel) 7.

Fawcett JA & Innan H. (2019). The Role of Gene Conversion between Transposable
Elements in Rewiring Regulatory Networks. Genome Biol Evol 11, 1723-1729.
Hagolani PF, Zimm R, Vroomans R & Salazar-Ciudad I. (2021). On the evolution and
development of morphological complexity: A view from gene regulatory networks.
PLoS Comput Biol 17, e1008570.

Jacques PE, Jeyakani J & Bourque G. (2013). The majority of primate-specific regulatory sequences are derived from transposable elements. PLoS Genet 9, e1003504. Johnson AD. (2017). The rewiring of transcription circuits in evolution. Curr Opin Genet Dev 47, 121-127.

Johnson R & Guigo R. (2014). The RIDL hypothesis: transposable elements as functional domains of long noncoding RNAs. RNA 20, 959-976.

Jung SA, Chapman CA & Ng WL. (2015). Quadruple quorum-sensing inputs control Vibrio cholerae virulence and maintain system robustness. PLoS Pathog 11, e1004837.

Keane OM, Toft C, Carretero-Paulet L, Jones GW & Fares MA. (2014). Preservation of genetic and regulatory robustness in ancient gene duplicates of Saccharomyces cerevisiae. Genome Res 24, 1830-1841.

Kim J, Vandamme D, Kim JR, Munoz AG, Kolch W & Cho KH. (2014). Robustness and evolvability of the human signaling network. PLoS Comput Biol 10, e1003763.

Li T, Chen B, Yang P, Wang D, Du B & Kang L. (2020). Long Non-coding RNA Derived from IncRNA-mRNA Co-expression Networks Modulates the Locust Phase Change.

Genomics Proteomics Bioinformatics 18, 664-678.

Liu G, Mattick JS & Taft RJ. (2013). A meta-analysis of the genomic and transcriptomic composition of complex life. Cell Cycle 12, 2061-2072.

Mattick JS. (2018). The State of Long Non-Coding RNA Biology. Noncoding RNA 4.

Morata J, Mar n F, Payet J & Casacuberta JM. (2018). Plant Lineage-Specific Amplification

of Transcription Factor Binding Motifs by Miniature Inverted-Repeat Transposable Elements (MITEs). Genome Biol Evol 10, 1210-1220. Moschetti R, Palazzo A, Lorusso P, Viggiano L & Marsano RM. (2020). "What You Need, Baby, I Got It": Transposable Elements as Suppliers of Cis-Operating Sequences in Drosophila. Biology (Basel) 9. Mottes F, Villa C, Osella M & Caselle M. (2021). The impact of whole genome duplications on the human gene regulatory networks. PLoS Comput Biol 17, e1009638. Nicolau M, Picault N & Moissiard G. (2021). The Evolutionary Volte-Face of Transposable Elements: From Harmful Jumping Genes to Major Drivers of Genetic Innovation. Cells 10. Nishihara H. (2020). Transposable elements as genetic accelerators of evolution: contribution to genome size, gene regulatory network rewiring and morphological innovation. Genes Genet Syst 94, 269-281. Oliveira MM, Shingleton AW & Mirth CK. (2014). Coordination of wing and whole-body development at developmental milestones ensures robustness against environmental and physiological perturbations. PLoS Genet 10, e1004408. Oo JA, Brandes RP & Leisegang MS. (2021). Long non-coding RNAs: novel regulators of cellular physiology and function. Pflugers Arch. Osterwalder M, Barozzi I, Tissieres V, Fukuda-Yuzawa Y, Mannion BJ, Afzal SY, Lee EA, Zhu Y, Plajzer-Frick I, Pickle CS, Kato M, Garvin TH, Pham QT, Harrington AN, Akiyama JA, Afzal V, Lopez-Rios J, Dickel DE, Visel A & Pennacchio LA. (2018). Enhancer redundancy provides phenotypic robustness in mammalian development. Nature 554, 239-243. Panni S, Lovering RC, Porras P & Orchard S. (2020). Non-coding RNA regulatory networks. Biochim Biophys Acta Gene Regul Mech 1863, 194417. Payne JL & Wagner A. (2014). The robustness and evolvability of transcription factor binding sites. Science 343, 875-877. Plata G & Vitkup D. (2014). Genetic robustness and functional evolution of gene duplicates. Nucleic Acids Res 42, 2405-2414. Qiu Y & K hler C. (2020). Mobility connects: transposable elements wire new transcriptional networks by transferring transcription factor binding motifs. Biochem Soc Trans 48, 1005-1017. Quinodoz S & Guttman M. (2014). Long noncoding RNAs: an emerging link between gene regulation and nuclear organization. Trends Cell Biol 24, 651-663. Rishishwar L, Wang L, Wang J, Yi SV, Lachance J & Jordan IK. (2018). Evidence for positive selection on recent human transposable element insertions. Gene 675, 69-79. Ruz GA, Goles E, Montalva M & Fogel GB. (2014). Dynamical and topological robustness

of the mammalian cell cycle network: a reverse engineering approach. Biosystems 115, 23-32. Salviano-Silva A, Lobo-Alves SC, Almeida RC, Malheiros D & Petzl-Erler ML. (2018). Besides Pathology: Long Non-Coding RNA in Cell and Tissue Homeostasis. Noncoding RNA 4. Senft AD & Macfarlan TS. (2021). Transposable elements shape the evolution of mammalian development. Nat Rev Genet 22, 691-711. Sharma R, Kumar S & Song M. (2021). Fundamental gene network rewiring at the second order within and across mammalian systems. Bioinformatics. Statello L, Guo CJ, Chen LL & Huarte M. (2021). Gene regulation by long non-coding RNAs and its biological functions. Nat Rev Mol Cell Biol 22, 96-118. Sundaram V & Wang T. (2018). Transposable Element Mediated Innovation in Gene Regulatory Landscapes of Cells: Re-Visiting the "Gene-Battery" Model. Bioessays 40. Trizzino M, Kapusta A & Brown CD. (2018). Transposable elements generate regulatory novelty in a tissue-specific fashion. BMC Genomics 19, 468. Trizzino M, Park Y, Holsbach-Beltrame M, Aracena K, Mika K, Caliskan M, Perry GH, Lynch VJ & Brown CD. (2017). Transposable elements are the primary source of novelty in primate gene regulation. Genome Res 27, 1623-1633. Voronova A, Rend n-Anaya M, Ingvarsson P, Kalendar R & Rungis D. (2020). Comparative Study of Pine Reference Genomes Reveals Transposable Element Interconnected Gene Networks. Genes (Basel) 11. Wang X, Ai G, Zhang C, Cui L, Wang J, Li H, Zhang J & Ye Z. (2016). Expression and diversification analysis reveals transposable elements play important roles in the origin of Lycopersicon-specific IncRNAs in tomato. New Phytol 209, 1442-1455. Zhang Y, Li Z, Zhang Y, Lin K, Peng Y, Ye L, Zhuang Y, Wang M, Xie Y, Guo J, Teng W, Tong Y, Zhang W, Xue Y, Lang Z & Zhang Y. (2021). Evolutionary rewiring of the wheat transcriptional regulatory network by lineage-specific transposable elements. Genome Res 31, 2276-2289. Zheng P & Triesch J. (2014). Robust development of synfire chains from multiple plasticity

mechanisms. Front Comput Neurosci 8, 66.