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# Research Article

# Reduced Blood to Brain Glucose Transport as the Cause for Hyperglycemia: A Model That Resolves Multiple Anomalies in Type 2 Diabetes

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Classically, type 2 diabetes is believed to be a result of insulin resistance and relative insulin deficiency. However, evidence has been accumulating against the insulin resistance models. The absence of lasting hyperglycemia in insulin receptor knockouts or insulin suppression, evidence for hyperinsulinemia preceding insulin resistance, the perplexing hyperinsulinemic normoglycemic state, reduced glucose transport to the brain preceding hyperglycemia, signs of vasculopathy preceding hyperglycemia, absent or poor correlation between fasting glucose and insulin, and a very strong positive correlation between indices of insulin resistance and β cell function in population data are some of the anomalous findings that classical glucose homeostasis models have not addressed so far. With increasing evidence for neuronal involvement in glucose regulation, we propose a refined model of glucose regulation that considers brain glucose and insulin levels as the ultimate target of homeostasis and combines central and peripheral mechanisms of regulation. A model considering a reduced rate of blood-to-brain transportation of glucose and insulin as the primary pathology explains most of the patterns, with or without insulin resistance or any other defect in glucose regulation mechanisms. Apart from resolving multiple anomalies, the model also accounts for the failure of glucose normalization in effectively reducing diabetic complications and mortality.

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# 1. Introduction

# 1.1. Flow and organization of the article

The hypotheses being considered and the outcome of the article deviate substantially from the prevalent mainstream beliefs in the field of type 2 diabetes mellitus (T2DM). Since the disruptive inference needs to be viewed in a broader perspective, we explain the flow of the article right at the beginning so that the reader does not lose focus amid the diversity of details considered. We first point out that the classical theory of T2DM, based on insulin resistance and inadequate compensation, has accumulated too many anomalies over the last few decades and therefore is under serious question. We need a new way of thinking and a specific model that can address all or most of the anomalies. After a brief historical account (section 1.2), we first list (section 1.3) the observed features in glucose homeostasis and T2DM that a model needs to explain and point out that the prevalent theory and models address only a minority of them. Then we summarize the predominant thinking based on peripheral mechanisms of glucose regulation (section 1.4), followed by the attempts so far to accommodate central regulation (section 1.5), along with the deficiencies in these ways of thinking. On this background, the objectives of the model are specified (section 1.6). Section 2 describes the assumptions (2.1) and the structure of the model (2.2). Results of the model are presented in three parts, namely the

steady-state solutions, simulations of glucose dynamics, and the population-level predictions. The discussion highlights the implications of the model for understanding T2DM, why a fundamental revision of the theory of T2DM is required, and what the implications are for clinical practice.

# 1.2. The burden of history

Presently, any metabolic disorder with consistently raised plasma glucose and insulin resistance, which cannot be categorized into either type 1 diabetes or gestational diabetes, is defined as type 2 diabetes mellitus (T2DM)<sup>[1]</sup>. It is the most common form of diabetes, accounting for over 90% of all cases. The classically perceived cause of hyperglycemia in T2DM is insulin resistance followed by failure of compensatory insulin response. Historically, the role of the brain in glucose homeostasis was revealed by experiments in which damage to certain parts of the brain was shown to impair homeostatic control<sup>[2]</sup>. However, at a later stage, pancreatic extracts were shown to lower glucose, and eventually, insulin was identified as the active principle [3]. It is necessary to note that the differentiation between type 1 and type 2 diabetes was not made at that time. The success of insulin was so spectacular that all other mechanisms of glucose regulation were almost forgotten [4]. The distinction between type 1 and type 2 diabetes was to become clear half a century later, and it was realized that unlike in type 1, insulin deficiency was not the primary cause of hyperglycemia in type 2 diabetes. However, by this time, the thinking in the field was so much insulin-centered that the inability of normal or increased levels of insulin to regulate plasma glucose was termed insulin resistance, without carefully evaluating alternative hypotheses [5].

The insulin resistance hypothesis is criticized for having a circular logic that makes it unfalsifiable [5]. Insulin resistance is said to be responsible for the inability of normal or raised levels of insulin to regulate glucose. However, insulin resistance itself is measured by the inability of insulin to regulate glucose. At the clinical level, there is no independent measure of insulin resistance that can be used to test the causal role of insulin resistance. Impairment of insulin signaling or fasting insulin levels in experiments using (i) tissue-specific insulin receptor knockouts [6][7] (ii) insulin-degrading enzyme knockouts [8][9] (iii) pharmaceutical insulin suppression by diazoxide and octreotide [10][11][12][13] (iv) insulin gene suppression by RNAi [14] (v) alteration in insulin gene dosage [15] failed to alter fasting glucose in the expected direction. In epidemiological data, fasting insulin and fasting glucose are poorly correlated, while post-meal insulin and glucose are strongly correlated, unlike what would be expected by any of the classical models. With converging multiple lines of evidence, Diwekar-Joshi and Watve [5] raised doubts on whether insulin resistance and failure of compensatory hyperinsulinemia is a necessary and sufficient explanation of fasting hyperglycemia.

Many mathematical models have been constructed based on the assumption of insulin resistance. A common set of assumptions is shared by most of the models that fasting glucose is the steady-state achieved by a balance between glucose uptake by tissues and glucose production by the liver. One of the foundational assumptions is that both of the above processes are regulated by insulin signaling in the steady-state [16]. Therefore, insulin signaling has been central to these models. Since recent research has exposed the inadequacies of this model [5], there is a need to hunt for better alternative models.

An alternative to the classical model, which assumes the brain glucose level as the target of homeostasis, has been suggested, and mathematical models incorporating this concept have been constructed [17][18]. These models have not been explored sufficiently towards a comparative evaluation with peripheral models with respect to their predictions and their physiological as well as clinical implications. Here, we refine the brain-centered model further by incorporating recent evidence, evaluate its performance in explaining the accumulated anomalies, and explore its physiological and clinical implications.

### 1.3. What a glucose homeostasis model needs to explain

Most models of glucose homeostasis have a limited goal of explaining altered fasting steadystate glucose levels and the alterations in the glucose curve in the pre-diabetic and diabetic states. However, now we have a large number of experimental results that are potentially anomalous. A model prediction matrix, in which each model is tested for matching its predictions with multiple known empirical patterns, is an appropriate and robust approach to compare alternative models. Such an approach has been used to test alternative hypotheses in a number of other contexts [19][20][21][22], and it can be the right approach to test alternative models of glucose homeostasis in T2DM. A model of choice would be one that can predict all or most of the known physiological patterns in type 2 diabetes so that no or minimal anomalies are left. We take a few steps towards a model prediction matrix, although a lot more exploration of many other models is required before constructing such a matrix.

The following is a list of known empirical patterns with which a glucose homeostasis model needs to be consistent.

- 1. Steady-state fasting glucose: Fasting glucose is assumed to be at a steady state with adequate evidence [23][16][24]. Achieving a steady state is one of the primary objectives of every homeostasis model. Many of the key concepts of classical models are also based on achieving a steady state in the fasting condition [16][24].
- Steady-state fasting insulin: Normally, this is not a problem for any model; however, many models have a problem explaining the altered steady-state insulin level in a prediabetic state, as explained below.
- 3. Normoglycemia with hyperinsulinemia in a prediabetic state: A prediabetic state is often accompanied by a high level of fasting insulin (FI) but normal fasting glucose (FG) in the plasma. This is not easy to achieve in a model. The classical qualitative explanation of this state is that it is a state of insulin resistance with a compensatory rise in FI such that FG remains normal. For this to happen, there is a need to estimate the level of insulin resistance and regulate insulin secretion accordingly. It is possible to hypothesize a mechanism of sensing insulin resistance and conveying this information to the pancreas. Such a mechanism can be glucose-dependent or glucose-independent. Currently, no glucose-independent mechanism of accurately assessing insulin resistance and conveying it to β cells is known. A common assumption is that insulin resistance reduces glucose uptake, thereby increasing FG. The rise in FG stimulates insulin production, and the rise in insulin levels normalizes glucose again. However, this mechanism fails to achieve a steady-state hyperinsulinemic normoglycemic condition $^{[5]}$ . Plasma insulin has a short half-life, and after glucose levels are normalized, it is unlikely to stay at a higher level unless there is a glucose-independent trigger for insulin secretion. Explaining a hyperinsulinemic normoglycemic steady state is a tricky challenge that any model needs
- 4. Hyperinsulinemia preceding insulin resistance and T2DM: Although hyperinsulinemia is believed to be a compensatory response to insulin resistance, a number of studies show that hyperinsulinemia precedes obesity and insulin resistance [25][14][26][27][28][29]. This raises two independent questions. One is how and why hyperinsulinemia precedes insulin resistance. The second, more perplexing question is that if hyperinsulinemia is not a compensatory response to insulin resistance, the failure of compensation explanation becomes redundant. This needs to be replaced by a more coherent explanation of hyperglycemia.
- 5. Explanations of features of the impaired glucose tolerance curve: There are three main features of an altered glucose tolerance curve that a model should explain, namely
  - 1. Increased height of peaks
  - 2. Delayed return to normal
  - Increase in the time difference between glucose and insulin peaks in the diabetic state.
- 6. Simultaneous presence of normal fasting glucose and impaired glucose tolerance (NFG-IGT) state: It is common to find that in a prediabetic state, the glucose tolerance curve is altered with a normal level of fasting glucose. It is tricky to explain this in a model.
- 7. Stress hyperglycemia: Why and how stress causes hyperglycemia only in some individuals [30] needs to be explained by a model with realistic and testable assumptions.
- 8. Hyperglycemia after intensive exercise: If in a fasting state *FG* is a resultant of the rate of liver glucose production and glucose disposal, exercise that increases glucose disposal should decrease *FG*. However, in some studies, plasma glucose remains unchanged or is increased after exercise [31][32]. A model needs to predict this result contextually.

- 9. Glucose dynamics in insulin receptor knockouts: An important challenge is to explain why insulin receptor knockouts specific to muscle, fat cells, or liver result in normal fasting glucose but altered post-glucose load curves. Similarly, insulin-degrading enzyme knockouts do not alter FG. Also, insulin suppression by agents such as octreotide or diazoxide fails to alter FG. A model should explain these observations. Diwekar-Joshi and Watve [5] further differentiated between the consequential steady-state (CSS) and targeted steady-state (TSS) models and demonstrated that the results of the insulin receptor knockout and insulin suppression experiments can be explained by a TSS but not by a CSS model. In effect, to be able to account for these results, a model needs to be a TSS model.
- 10. Reduced glucose transport to the brain in obese or prediabetic individuals: It has been known for a long time that in type 2 diabetes, the rate of glucose transport from blood to brain is slowed down substantially [33]. In rodent models, the GLUT-1 expression in brain capillaries is shown to be reduced [34][16][35]. This has been viewed as a response to hyperglycemia. However, Hwang et al. [36] show that subnormal transport is evident in obese and presumably prediabetic states, even before hyperglycemia appears. A model needs to account for this pattern as a causal or consequential phenomenon.
- 11. The T2DM-specific islet amyloid deposition: Islet amyloid deposition is frequently associated with type 2 diabetes but not observed in type 1. The causes of islet amyloid deposition should be compatible with the glucose homeostasis model [37][38][39].
- 12. Beta cell deterioration pattern: The deterioration of  $\beta$  cells appears to follow a peculiar pattern in T2DM, in which a substantial proportion of  $\beta$  cells survive lifelong [40][41][42][43] [18]. If dysfunction and damage to the  $\beta$  cell population are assumed to be essential prerequisites of hyperglycemia in a model, the models need to account for the peculiar population dynamics of  $\beta$  cells.
- 13. Increased liver glucose production and ketogenesis with SGLT2 inhibitors: Over the last decade, SGLT2 inhibitors, which allow greater clearance of glucose through urine, have offered a novel means of combating hyperglycemia. Lowering of plasma glucose by SGLT2 inhibitors is accompanied by increased liver glucose production as well as increased ketogenesis [44][45][46][47], the causes of which should be made clear by the model.
- 14. The Somogyi phenomenon: i.e., hyperglycemia following the infusion of insulin, is seen in at least some cases of T2DM but not in healthy individuals. This can happen without being overtly hypoglycemic [48][49].
- 15. Hyperglycemia in brain injury and bacterial meningitis: In cases of brain injury, hyperglycemia is common<sup>[50]</sup>. In bacterial meningitis, hyperglycemia is often observed accompanied by a low cerebro-spinal fluid (CSF) glucose level<sup>[51][52]</sup>. A model should be able to account for the apparent contradiction.
- 16. Good correlation between post-meal glucose and insulin but poor correlation between FG and FI  $^{[5]}$  in population data.
- 17. HOMA- $\beta$  and HOMA-IR correlation in population data: Although *FG* and *FI* are poorly correlated in the prediabetic state, HOMA- $\beta$  and HOMA-IR have a strong positive correlation in population data<sup>[5]</sup>.
- 18. Failure of glucose normalization to reduce the frequency of complications and mortality: Unlike T1DM, in T2DM, tight regulation of plasma glucose has failed to show a reduction in mortality consistently across multiple large-scale clinical trials [53][54][55][56][57][58][59] [60][61]. If hyperglycemia is the primary pathology of type 2 diabetes, preventing or correcting it should have reduced the frequency of complications and mortality considerably.
- 19. Reversal of hyperglycemia by FGF 21 in all models of rodent diabetes: Independent of the cause of hyperglycemia, a single injection of FGF 21 was able to achieve long-term normalization of plasma glucose [62].

While most models intend to explain 1, 2, and 5 of the above, they either fail to explain or do not address the others. A satisfactory model should predict all the phenomena noted above. If an additional explanation that does not contradict the model accounts for the phenomenon, the model can be said to be compatible. Compatibility with empirical findings cannot be taken as proof or validation of the model, but if a model directly contradicts one or more of the empirical

findings, it becomes serious grounds for rejecting the model. Later, when we describe model results, we will use the numbers in the above list in square brackets to indicate that the model accounted for this pattern.

## 1.4. <u>Peripheral models of glucose homoeostasis</u>

There is a long history of development of mathematical models of glucose homeostasis and the origin and progression of type 1 and type 2 diabetes (reviewed by [63][64]). The focus of the field is so much on peripheral mechanisms that the review by Mari et al. [64] does not even cite the models incorporating the role of the brain. Ajmera et al. [63] briefly mention the Gouhua et al. [17] model but do not elaborate on its potential implications. A class of models attempts to capture glucose homeostasis at the systems level, whereas other models look at individual components such as insulin-dependent glucose uptake, liver glucose production, β cell dysfunction, glucose-stimulated insulin secretion, and incretin effects in greater detail. Nevertheless, these models together have not accounted for the majority of the patterns listed above. The central assumption of these models is more or less invariant and revolves around insulin resistance and the failure of the insulin response to adequately compensate for it [64]. Diwekar-Joshi and Watve<sup>[5]</sup> claimed that any model with this set of assumptions is not compatible with empirical patterns 3, 4, 6, 9, 16, and 17 of the above. Whether any variation of these models can do so has not been adequately explored. Furthermore, all the peripheral regulation models are CSS models in that the fasting steady-state is a consequence of the rate of liver glucose production and a concentration-dependent rate of glucose uptake. In these models, a change in either or both of the rates inevitably alters the steady-state. This contrasts with the TSS models, in which alteration in these rates alters the time required to reach a steady-state but does not alter the steady-state glucose level<sup>[5]</sup>. An attempt to develop a TSS model with only peripheral mechanisms has not been made to the best of our knowledge.

We do not intend to review glucose homeostasis models here. However, we observe from published reviews of glucose homeostasis models [63][64] that most models do not address the apparently anomalous empirical findings, particularly 3, 4, 6, 9, 10–13, and 15–18 among the ones listed above. A detailed exploration into whether some variations of these models can address the apparently anomalous patterns is beyond the scope of this paper, but we remain open to this possibility.

# 1.5. Brain-centered models of glucose homeostasis

It is quite well known that a number of neuronal mechanisms in the brain are involved in energy homeostasis. Nevertheless, for some reason, they did not occupy a central stage in the mainstream thinking about T2DM and glucose regulation models until recently. A number of recent publications highlight the role of the brain in different ways [65][66][67][18][68][69][4][70][71] [72][73]. Although there is substantial evidence for the role of the central nervous system in glucose regulation, a sound theory addressing the question of why a dual control system evolved, how the two components interact, what goes wrong during T2DM, and why, is yet to develop. The prevalent thinking about the role of the brain in glucose regulation has not deviated from the baseline assumptions that peripheral glucose is the target of homeostasis, insulin resistance is the primary pathology, and some defect in the central and/or peripheral regulation system is required for glucose dysregulation. If the peripheral glucose level is the target of homeostasis, then a central regulation system is bound to be highly inefficient because of the inevitable delay in crossing the blood-brain barrier [74]. But central mechanisms still exist. Therefore, it is necessary and possible to rethink these assumptions to see whether that helps resolve the mounting anomalies in the field.

Peters (2004), Gaohua and Kimura<sup>[17]</sup>, and Watve<sup>[18]</sup> proposed that the primary target of glucose homeostasis is to regulate glucose levels in the brain. Plasma glucose levels are only a means to achieve the required supply of glucose to the brain. Since the transport of glucose to the brain is more restricted, when an adequate supply of glucose to the brain is ensured, it is likely that the supply to other organs is already ensured. This is a fundamental deviation from the assumption that plasma glucose levels are under homeostatic control.

In the Gaohua and Kimura<sup>[17]</sup> as well as in the Watve<sup>[18]</sup> model, the rate of glucose transport across the blood-brain barrier (BBB) is assumed to be an adaptation to hyperglycemia. The finding that glucose transport is reduced to an intermediate level in obese and prediabetic individuals<sup>[75][36]</sup> suggests that the deficiency in transport precedes hyperglycemia rather than following it. Therefore, it is more likely to be causal than consequential. Glucose deficiency in the brain is known to induce liver glucose production and suppress insulin release through autonomic control<sup>[76]</sup>. The sympathetic tone has also been shown to be higher in T2DM<sup>[77]</sup>. Therefore, glucose deficiency in the brain owing to altered vascular function is likely to be primary, which results in altered plasma glucose dynamics mediated by autonomic inputs. It is possible that there is no defect in the glucose sensing and regulation mechanisms in T2DM. Only defective glucose transportation may be responsible for the glucose dysregulation. Our model incorporates and examines this possibility to address the question of whether it helps resolve the anomalies.

Impaired vascular function is known to be central to diabetic complications. The classical thinking has been that hyperglycemia causes the types of vasculopathies typical of T2DM. However, it has not been ruled out that vasculopathies are not primary. There is considerable evidence that microvascular alterations precede T2DM and are good predictors of it [78][79][80] [81]. A sedentary lifestyle and deficiency of many specific types of physical activities and behaviors alter the expression of growth factors and endocrine mechanisms involved in angiogenesis (Watve<sup>[18]</sup>). It has been demonstrated repeatedly that many growth factors and angiogenic factors are responsive to behaviors, physical activity, and exercise [82][83][84][85][86] [87][88]. Deficiency of these behaviors is likely to lead to a primary deficiency of angiogenic mechanisms [18]. The possibility that altered vascular function in the brain is the primary reason why glucose transport from blood to brain is reduced needs to be considered. This creates chronic glucose deficiency in the brain, to which the brain reacts by influencing multiple mechanisms of glucose regulation. The predominant mechanism is autonomic signaling. It is well known that sympathetic and parasympathetic tones are altered in T2DM<sup>[77]</sup>. The possibility that altered autonomic signaling is causal rather than consequential to diabetes looks promising, based on studies showing that changes in autonomic function precede T2DM[89].

Brain glucose uptake is insulin-independent  $\frac{[90][91]}{[91]}$ . Nevertheless, insulin has many other functions in the brain, and insulin signaling is known to be important in memory, cognition, decision-making, behavior, and also in regulating energy reserves  $\frac{[92][93][94]}{[93][94]}$ . The brain is rich in insulin receptors, and activation of certain cognitive functions in the brain is a likely cause of increased brain glucose utilization by insulin stimulation  $\frac{[95][96]}{[95]}$ . Although there are indications of insulin synthesis in the brain itself, there are many uncertainties in its implications  $\frac{[97]}{[97]}$ . Also, there is no human data on brain-derived insulin. Therefore, it is difficult to incorporate this factor in the model currently. The assumption that the brain has a fundamental requirement for pancreatic insulin, independent of glucose metabolism, is fair, and therefore mechanisms to ensure the required insulin levels in the brain are as critical as ensuring a minimum brainglucose level. This is also ensured by autonomic mechanisms. Autonomic inputs are known to regulate the  $\beta$  cell population as well as insulin release from  $\beta$  cells  $\frac{[98]}{}$ . Therefore, brain control over insulin production independent of peripheral glucose is also an essential part of the theory.

Ensuring a minimum supply of glucose as well as insulin to the brain is crucial during fasting when the plasma levels are low. The post-meal levels of glucose and insulin in the plasma are much higher, and at this time, the brain need not actively regulate the plasma levels of both. This can be taken care of by peripheral mechanisms.

# 1.6. Objectives of the model

Our attempt is to construct a model whose predicted outcomes match with all or most of the observed phenomena listed in section 1.3, qualitatively. We intend to construct a model in which different hypotheses for hyperglycemia can be used to make differential testable predictions. This approach can allow us to evaluate comparatively which causal factors,

individually or in combination, can give us the set of predictions that match with the patterns listed above.

Out of the parameters required for the model, only some have empirical estimates available (Table 1). In the absence of realistic estimates for all parameters, we do not intend to make a model that makes quantitative predictions. When a sufficiently large number of parameters can be optimized, it is not difficult to fit the data quantitatively. Therefore, we prefer qualitative predictions over quantitative ones. We test whether the model is able to predict the pattern observed empirically under some set of parameters. The ability to predict a given pattern is not proof of the validity of the model, but the inability to predict an observed phenomenon at any set of parameters within a realistic range is a strong reason to call the model either inadequate or wrong. If an additional consideration compatible with the model is able to explain a pattern not explained by the main model, the model can be called inadequate but not falsified. However, if the model outcome directly contradicts a consistent and reproducible empirical finding, it can be considered falsifying evidence. We emphasize the need to evaluate our model in comparison with classical models along these lines.

### 2. Methods

# 2.1. Assumptions of the model

A unique assumption of our model, based on the analysis by Diwekar-Joshi and Watve $^{[5]}$ , is that the mechanism of regulation of glucose and insulin is different in the steady-state and postmeal state. In the steady-state, the central mechanisms are more important, whereas in the post-meal state, mainly the peripheral mechanisms are at work. Insulin-induced glucose uptake and insulin-dependent inhibition of liver glucose production happen only above a threshold glucose and insulin level, respectively. The presence of such thresholds and their context-dependent variability has been known for a long time $^{[99][100][101]}$ . Below the threshold, other mechanisms regulate the levels $^{[101]}$ . We assume in the model that the thresholds are under neuro-endocrine control and fine-tuned by the brain. The thresholds found in isolated cell cultures are slightly lower than the normal fasting blood sugar, and it is difficult to estimate thresholds in vivo $^{[99][100]}$  (Henquin et al., 2015). In the presence of autonomic signals, we assume the thresholds to be above the steady-state target levels. In conditions under which the fasting levels need to increase, we assume the thresholds to increase proportionately. This may happen with peripheral or central mechanisms. These assumptions about the threshold make the model a TSS model.

The steady-state glucose is decided by a balance between the basal level of liver glucose production, which is regulated by neuronal mechanisms, and insulin-independent glucose uptake by tissues. The steady-state insulin level is decided by the basal rate of glucose-independent insulin production, which is regulated by neuronal mechanisms against the insulin degradation rate. Glucose is transported from plasma to brain by facilitated diffusion proportionate to the capillary surface area in the brain and GLUT-1 expression. Steady-state brain glucose is determined by glucose transport to the brain and utilization by brain tissue. The brain needs to maintain a target level of glucose, and the brain ensures it by directly regulating liver glucose production through neuronal mechanisms.

Insulin has cognitive and other functions in the brain  $\frac{[102][92][103][93][104]}{[93][104]}$ , for which the brain requires a target level of insulin, which it ensures by autonomic regulation of  $\beta$  cell number and basal insulin secretion independent of peripheral glucose. To ensure the brain target level for insulin, a minimum plasma insulin level needs to be maintained. Maintenance of this level is assumed to be independent of glucose-stimulated insulin secretion.

The rate of transport of glucose and insulin from blood to brain is not constant. Both are affected by capillary density and blood flow. But they also depend upon specific transport mechanisms. Glucose is transported across the blood-brain barrier mainly through the specific transporter GLUT-1. The expression of GLUT-1 in the brain and other tissues is variable and dependent on multiple dietary, endocrine, and growth factor-related mechanisms [105][106][107] [108]. Glucose transport from blood to brain is diminished in obesity and prediabetes [36], resulting in hypometabolism [109]. Recent multiple independent studies reproducibly report glucose level-dependent hypometabolism in several regions of the brain in various stages of

diabetes, including prediabetes, along with detectable cognitive decline [110][111][112][113][114]. Insulin transport to the brain is also reduced in obesity [115]. The ratio of CSF to plasma insulin is inversely proportionate to obesity and insulin resistance [116]. Moreover, inducing endothelial dysfunction and reducing glucose transport experimentally by endothelial–specific deletion of HIF  $1-\alpha$  resulted in hyperglycemia [117]. Antipsychotic drugs are known to impair angiogenesis [118][119][120] and thereby induce hyperglycemia [121][122] (Henderson, 2012). These findings validate our assumptions, based on which we incorporate the possibility of diminished glucose and insulin transport to the brain being the primary pathology of T2DM, and hyperglycemia only a consequence.

### The model

Since we assume that the steady-state and post-meal mechanisms of glucose regulation are not identical, we model and analyze the two states in two phases of the model. The variables and parameters used in the model and the range of parameters used are in Table 1.

serial no	Variables and parameters	meaning	units	Value/range used in simulations			
A. Input parameters							
1	$K_1$	Insulin independent glucose uptake maximum	mg/dL/min	20-50			
2	K <sub>1m</sub>	Half saturation constant for insulin independent glucose uptake	mg/dl	50			
3	K <sub>2</sub>	Maximum insulin induced suppression of PG, including glucose disposal and suppression of L.	per µU/mL	0.001 to 0.05			
4	K <sub>2m</sub>	Half saturation constant for insulin induced suppression of PG	μU/mL	15			
5	K <sub>5</sub>	Maximum glucose dependent insulin secretion	μU/mL/min	2-20			
6	K <sub>5m</sub>	Half saturation constant for glucose induced insulin secretion	mg/dL	50			
7	K <sub>8</sub>	Maximum rate of glucose transport to brain	Mg/dL/min	3 - 8			
8	K <sub>8m</sub>	Half saturation constant for glucose transport to brain	mg/dL	50-100			
9	K <sub>9</sub>	Maximum rate of Insulin transport to brain	μU/mL/min	0.6 - 1			
10	K <sub>9m</sub>	Half saturation constant for insulin transport to brain	μU/mL	0.3-1			
11	K <sub>10</sub>	Brain glucose utilization rate	unitless	0.08 to 0.12			
12	$d_1$	Insulin degradation rate	unitless	0.1 to 0.2			
13	BGt	Target BG	mg/dL	25			
14	BIt	Target BI	μU/mL	1-5			
15	$I_{S}$	Insulin sensitivity as compared to normal which is assumed 1.	unitless	0-1			
16	L <sub>max</sub>	Maximum capacity of liver glucose production	Mg/dL/min	30			
17	K <sub>4max</sub>	Maximum capacity of peripheral glucose independent insulin secretion	μU/mL/min	10			
		B. Derived variables					
18	L	Rate of liver glucose production	mg/dL/min	Eq. 6 or iterative			
19	$K_4$	Rate of peripheral glucose independent insulin secretion	μU/mL/min	Eq 7 and 8 or iterative			
		C. Outcome variables					
20	PG	Plasma glucose	Mg/dL	dynamic			
21	PI	Plasma insulin	μU/mL	dynamic			
22	PG	Steady-state plasma glucose	mg/dL	Eq. 5			
23	PI	Steady-state plasma insulin	μU/mL	Eq. 7			
24	BG	Brain glucose	mg/dL	dynamic			
25	ВІ	Brain insulin concentration	μU/mL	dynamic			
26	PGT	glucose threshold above which GSIS begins	mg/dL	5 to 10 mg/dL above PG			

serial no	Variables and parameters	meaning	units	Value/range used in simulations
27	PIT	Insulin threshold above which insulin induced glucose disposal begins	μU/mL	2-5 μU/mL above PI

Table 1. all variables and parameters considered in the model.

### 2.2. The Model

### A. Modeling steady-state glucose

As per the assumption, below a threshold *PIT*, insulin-induced glucose uptake and insulin-induced inhibition of liver glucose production are negligible. Therefore, we write,

### 1. Plasma Glucose:

$$\frac{dPG}{dt} = \begin{cases}
L - \frac{K_1 * PG}{K_{1m} + PG}, +Gt & (\underline{PI} < PIT) \\
L - \frac{K_1 * PG}{K_{1m} + PG} + Gt - \left(I_S * \frac{K_2 * PI}{K_{2m} + PI}\right) * PG, & (PI > PIT)
\end{cases}$$
(Eq. 1)

### 2. Plasma Insulin:

$$\frac{dPI}{dt} = \begin{cases} K_4 - (d1 * PI), & (\underline{PG} < PGT) \\ K_4 - d1 * PI + \frac{K_5 * PG}{K_{5m} + PG} * (PG - PGT1), & (\underline{PG} > PGT) \end{cases}$$
(Eq. 2)

### 3. Brain glucose:

$$\frac{dBG}{dt} = \frac{K_8*(PG-BG)}{K_{8m}+(PG-BG)} - K_{10}*BG$$
 (Eq. 3)

Since the brain forms only about 3 % of total body mass, we assume the instantaneous plasma glucose diffused to the brain is a negligible fraction of total plasma glucose. Therefore, that term is not included in the plasma glucose dynamics. Similarly, insulin diffusion to the brain is not represented in the plasma insulin dynamics. Nevertheless, the rate of plasma to brain diffusion is crucial in determining the brain glucose and insulin dynamics.

### 4. Brain insulin:

$$\frac{dBI}{dt} = \frac{K_9 * (PI - BI)}{K_{9m} + (PI - BI)} - d_1 * BI$$
 (Eq. 4)

At steady-state (SS), all differential terms become zero, and steady-state levels can be calculated using equilibrium solutions.

As per the assumption of the model, the brain needs a certain level of glucose, and that is the target of homeostasis BGt. In order to ensure BGt at given  $K_8$ ,  $K_{10}$ , and  $K_{8m}$ , the SS plasma glucose PG needs to be

$$\underline{PG} = \frac{BG_t * K_{10} * K_{8m}}{K_8 - (BGt * K_{10})} + BGt$$
 (Eq. 5)

We assume that the brain ensures this plasma glucose level by regulating liver glucose production *L*. To maintain the desired plasma glucose level at steady-state, the liver glucose production should be:

$$L = \frac{K_1 * \underline{PG}}{K_{1m} + \underline{PG}}$$
 (Eq. 6)

The brain can either ensure this by sending a calculated signal to the liver through the autonomic system. Alternatively, this can be ensured by the brain by increasing sympathetic inputs when BG < BGt and parasympathetic when it is above target [123][124][125]. The required L

can be achieved iteratively by  $L_{(t+1)} = L_{(t)} + K_s$ . (BGt - BG) where  $K_s$  is the magnitude of the iterative step.

Similarly, to ensure required brain insulin levels, plasma insulin should be

$$\underline{PI} = \frac{d_1 * BIt * K_{9m}}{K_9 - (d_1 * BIt)} + BIt$$
 (Eq. 7)

To ensure this much plasma insulin level,  $K_{\Delta}$  should be

$$K_4 = d_1 \cdot \underline{PI} \tag{Eq. 8}$$

Similar to glucose, the brain can ensure the target level of insulin in the brain by neuronally modulating  $K_4$  in a calculated or iterative manner. Specific autonomic inputs are known to regulate  $\beta$  cell number as well as insulin release from  $\beta$  cells  $\frac{[102][76][125]}{[102][76][125]}$ .

We also assume that both L and  $K_4$  have a fixed upper limit as  $L_{max}$  and  $K_{4max}$  when the maximum capacity of the liver and pancreas is reached. L and  $K_4$  cannot exceed this limit even if neuronal inputs become more intense. This happens when  $K_8$  declines below a threshold such that,

$$K_8 < rac{K_1 - L_{ ext{max}} \left( BGt * K_{10*} K_{8m} 
ight)}{L_{ ext{max}} * K_{1m} - BGt \left( K_1 - L_{ ext{max}} 
ight)} - \left( BGt * K_{10} 
ight)$$
 (Eq. 9)

There is a potential conflict here. Sympathetic tone is known to increase  $L^{[126][127]}$  whereas parasympathetic signal is required for the proliferation of  $\beta$  cells. Simultaneously, sympathetic signal is known to inhibit insulin release from  $\beta$  cells  $\frac{[128][129]}{[129]}$ . If the brain is deficient in glucose as well as insulin, both sympathetic and parasympathetic tones will be simultaneously higher  $\frac{[39]}{[39]}$ . This is incorporated into the model by updating  $K_4$  iteratively according to both BG and BI levels simultaneously,

$$K_{4(t+1)} = K_{4(t)} + K_p(BIt - BI) - K_s(BGt - BG)$$
 (Eq. 10)

where  $K_p$  and  $K_s$  are the iteration step lengths.

By this consideration, if sympathetic stimulation of liver glucose production is adequate to restore the required BG, BGt-BG will tend to zero, and there will be no interference in insulin release by  $\beta$  cells. However, in case the rise in liver glucose production is inadequate to ensure the required BG,  $K_4$  will be affected, which will also influence  $\beta$  cell function.

# B. <u>Simulating glucose tolerance curve</u>

While for the steady state, equilibrium solutions can be algebraically derived, the post-glucose load curve and its properties can only be obtained by simulations. Simulations were run using the same set of equations and giving a positive ephemeral *Gt* to simulate food intake.

In all the simulations used, insulin resistance can be simulated by altering Is,  $\beta$  cell dysfunction by reducing  $K_4$  and/or  $K_5$ , and vascular defect slowing down the transport of glucose and insulin to the brain and other organs by altering  $K_8$ ,  $K_9$ , and  $K_1$ . Mental stress is assumed to increase  $K_{10}$ . By differentially altering these parameters, the model can separately and collectively examine the effects of insulin resistance,  $\beta$  cell dysfunction, reduced rates of diffusion across the BBB, and stress on the glucose tolerance curve.

<u>T2DM- causal analysis</u>: We ask the question which minimal set of changes can give rise to a stable increase in fasting glucose, changes in the glucose tolerance curve characteristic of type 2 diabetes, along with other patterns listed above. The putative causal factors are examined individually and in combination. The factors include insulin resistance (decreased  $I_s$ ), subnormal β cell response (reduced  $K_4$  and/or  $K_5$ ), reduced insulin-independent glucose uptake (decrease in  $K_1$ ), reduced blood-to-brain transmission of glucose ( $K_8$ ) and insulin ( $K_9$ ), and stress-related increase in brain glucose consumption ( $K_{10}$ ). Subnormal or defective vasculature is expected to decrease  $K_1$ ,  $K_8$ , and  $K_9$  proportionately; however, the decrease in the three parameters may not be proportionate if the vasculature in different parts of the body is differentially affected. Also, glucose transporters and their expressions can alter in a tissue-

specific manner. Therefore, we allow proportionate as well as differential decreases in the three parameters.

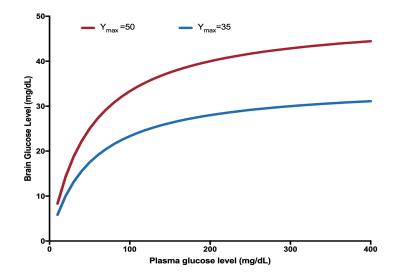
# 3. Results

### A. Steady-state solutions

We observed that the steady-state solutions and the autonomic iteration simulations match well in the end result except when the limits of L and  $K_4$  are reached. However, it took a long time to reach the desired level by the iterative approach, particularly when the desired level was substantially different from the starting level. In real life, major changes in vascular function or glucose transporter levels reflected in  $K_8$ ,  $K_9$ ,  $K_1$  happen gradually. Therefore, the desired level can be attained by autonomic fine-tuning over time. Since in the iterative simulations, a long time was required to reach SS, we used the steady-state solutions for quicker results.

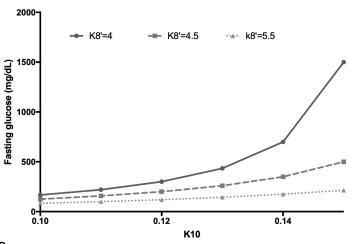
It can be seen that the steady-state levels of PG in the model are a stable steady-state since substituting PG < PG leads to a positive value and PG > PG leads to a negative value of dPG/dt. The same applies to plasma insulin, brain glucose, and brain insulin. This ensures a stable steady-state of glucose-insulin in the plasma as well as in the brain during fasting [1,2]. From equations 5, 6, and 8, we see that FG does not change by altering Is or FI. This is compatible with the empirical findings that insulin receptor knockouts and insulin suppression experiments fail to increase fasting plasma glucose [9]. This result is unique to our model and is due to segregating fasting and post-meal glucose regulation mechanisms by using thresholds. This is the only explanation offered so far for patterns [9], [16], and [17]. Prediction [10] directly follows from reducing  $K_8$  and  $K_9$ .

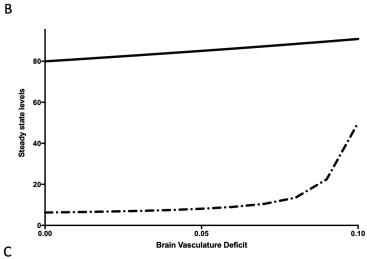
The steady-state solution of our model shows that FG can increase as a result of an increase in  $K_{10}$  or a decrease in  $K_8$ . That is, if the glucose consumption in the brain increases and/or the rate of glucose transport from blood to brain decreases. Both effects are interdependent, and the shape of the glucose response is decided by the interaction between  $K_8$  and  $K_{10}$ . Increased  $K_{10}$  increases FG marginally and almost linearly when  $K_8$  is large. When  $K_8$  decreases, even a marginal rise in  $K_{10}$  can induce a disproportionately greater rise in FG (fig 1b). This means that stress-induced hyperglycemia is unlikely to be seen in healthy individuals, while it is more likely in individuals with reduced vascular transport [7].



Α

# Effect of brain glucose transport and utilization on fasting glucose levels





**Figure 1.** The same input different output phenomenon A. A conceptual diagram with arbitrary units: A saturation relationship in glucose and insulin

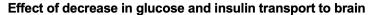
transport described by a Michaelis Menten type of equation has important consequences that can account for many phenomena observed in prediabetic and diabetic stages. For example, here, to achieve an increase in brain glucose by 6 mg/dL with  $Y_{max} = 50$ , plasma glucose needs to increase only by 6 mg/dL, but at  $Y_{max} = 35$ , for the same target increase in brain glucose, 25 mg/dL of increment in plasma glucose is needed. This non-linearity explains many phenomena, including prediabetic hyperinsulinemia and stress-induced diabetes in our model of glucose homeostasis.

B. A simulation using the steady-state model: It can be seen that as  $K_8$  decreases, for the same change in brain requirement  $K_{10}$ , a non-linear escalated increase in plasma glucose is required.

C. A simulation result assuming a correlated decrease in  $K_8$  and  $K_9$ . It is possible that while plasma glucose (solid line) shows a marginal increase, plasma insulin (dotted line) increases substantially, since the parameters of glucose and insulin transport curves are different. This is a potential explanation for the hyperinsulinemic normoglycemic state.

The origin of the hyperinsulinemic normoglycemic condition can be explained by the difference in the parameters of the saturation curves of transport dynamics of glucose and insulin. If the insulin diffusion is assumed to be nearer to saturation and glucose diffusion is sufficiently away from saturation, at the same level of vascular function deficiency, insulin will increase disproportionately more than glucose (Figure 1). This is a possible cause of the hyperinsulinemic normoglycemic state [3], and the reason why hyperinsulinemia precedes hyperglycemia [4], which requires neither insulin resistance nor a compensatory hyperinsulinemic response. When this happens, both HOMA-IR and HOMA  $\beta$  increase, although there is no change in true insulin resistance and compensatory insulin response. This gives a false impression of insulin resistance as clinically defined, although the actual insulin resistance at the cell level may not have changed.

Further, as the transport saturation constants  $K_{8m}$  and  $K_{9m}$  continue to decrease proportionately, FG increases monotonically, but FI shows a non-monotonic curve in which FI increases with a moderate decrease in transport rates but decreases after a threshold decrease in transport rates. This non-monotonicity is predicted by the iterative autonomic inputs model. When the required L approaches or exceeds  $L_{max}$ , the steady-state BG remains smaller than BGL. The resultant increase in the sympathetic signal inhibits insulin release from  $\beta$  cells. This leads to reduced insulin secretion, and insulin levels drop substantially. Thus, without bringing in additional factors, the model explains the early phase rise in fasting insulin as well as the later phase decline in the course of T2DM [4] (Figure 2).



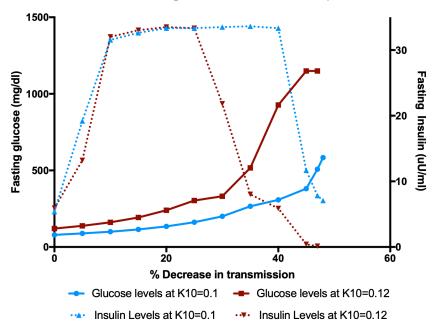


Figure 2. Effect of correlated decrease in  $K_8$  and  $K_9$  on fasting plasma glucose and insulin. Note that for a small to moderate decrease, FI increases rapidly with marginal change in FG. However, after a threshold decrease in the transport rates, FI declines sharply whereas FG increases rapidly. This pattern matches the course of clinical T2DM without involving insulin resistance, compensatory hyperinsulinemia, and  $\beta$  cell exhaustion. Simulation-specific other parameters were  $K_{\rm Im}$  =50,  $K_1$ =30,  $K_{\rm 2m}$ =15,  $K_2$ =0.01,  $K_{\rm 5m}$ =0.01,  $K_5$ =0.5,  $d_1$ =0.15,  $I_s$ =1, BGt=25, BIt=5,  $K_8$ m=80,  $K_8$ =5,  $K_9$ m=0.3,  $K_9$ =0.8,  $K_{\rm 10}$ =0.1 (Blue line) and 0.12 (Red line).

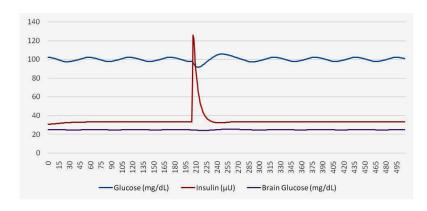
This decline in the insulin response does not require β cell dysfunction as a causal mechanism, but there are more complex possible effects on β cells. Simultaneous activation of sympathetic and parasympathetic inputs to  $\beta$  cells is implicated in  $\beta$  cell amyloidogenesis [39]. Parasympathetic stimulation is known to increase β cell number. However, sympathetic signaling suppresses insulin release, resulting in increased retention time of insulin along with amylin, which beyond a threshold retardation may result in spontaneous polymerization of amyloid protein, leading to poisoning of  $\beta$  cells and amyloidogenesis [39]. This is an alternative causal interpretation of  $\beta$  cell dysfunction and decline in its population. This mechanism has a built-in negative feedback loop, giving rise to a steady-state  $\beta$  cell population. This result is compatible with the finding in which the  $\beta$  cell population remains subnormal lifelong [11,12]. This is in contrast with the dynamics expected by the classical thinking in which  $\beta$  cells are destroyed by gluco-lipotoxicity or oxidative stress. This mechanism has a built-in positive feedback cycle. As a part of the  $\beta$  cell population is destroyed, the insulin secretion is decreased, which would increase glucose, further accelerating gluco-lipotoxicity and oxidative stress, and thereby  $\beta$  cell loss. Such a positive feedback vicious cycle can only stop with complete destruction of the  $\beta$  cell population. This prediction of the classical model does not match the finding of sustained presence of a subnormal  $\beta$  cell population in T2DM.

By the classical model, prolonged sustained exercise in the fasting state should result in lower plasma glucose. However, an experimental increase in glucose uptake resulting from physical activity during fasting does not necessarily result in a decrease in FG. This is mainly because as FG starts decreasing, FG decreases consequently. FG being lower than FG induces a sympathetic-mediated increase in FG, and the change in FG by physical activity is more or less compensated. This may happen through the agency of glucagon as well, since a strong link between autonomic signals and glucagon secretion is known. Therefore, at times, plasma

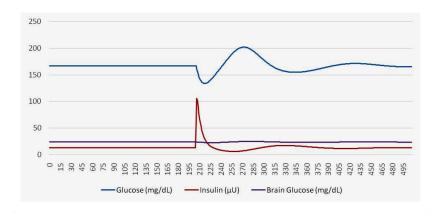
glucose may actually increase after intensive exercise. Further, if exercise involves heightened activation of cerebellar mechanisms of coordination,  $K_{10}$  may also increase, resulting in higher rather than lower FG in response to exercise [8].

A high dose of exogenous insulin results in increased plasma glucose after a short time lag, which is known as the Somogyi phenomenon. The phenomenon is conditional and not seen in every diabetic. It is said to involve the counter-regulatory response when facing central hypoglycorrhachia. However, often this response is seen without peripheral hypoglycemia. This is best explained by a reduced  $K_8$  that gives rise to a decreased BG/FG ratio. With a smaller  $K_8$ , at apparently normal or increased plasma glucose, brain glucose can still be lower than the target, which gives a sympathetic signal to increase L. Simulations using the autonomic iteration model show this phenomenon quite well at low  $K_8$ . At the same level of insulin infusion, at lower  $K_8$ , there is a more intense hyperglycemic response (Figure 3) [14]. A model without accommodating changes in  $K_8$  does not predict the Somogyi phenomenon without an obvious hypoglycemic state.

# At transportation rate 90%



# At transportation rate 70%



**Figure 3.** Effect of high-dose insulin on peripheral glucose under two rates of glucose and insulin transport across the blood-brain barrier (Correlated decrease in  $K_8$  and  $K_9$ ). A paradoxical rise in blood sugar following insulin administration is more prominent when the transport rate is substantially lower than normal. Other parameters ( $K_{\rm Im}$  =50,  $K_1$ =30,  $K_2$ m=15,  $K_2$ =0.08,  $K_5$ m=0.01,  $K_5$ =15,  $K_1$ =0.15,  $K_2$ =1, BGt=25, BIt=4.5,  $K_8$ m=80,  $K_8$ =5,  $K_9$ m=0.3,  $K_9$ =0.8,  $K_{10}$ =0.1)

The response is restricted to a narrow set of conditions. For an intense hyperglycemic response to insulin administration, it is necessary that insulin sensitivity is good, *BG* is close to *BGt* 

before insulin transfusion, and  $K_8$  is subnormal.

Potentially, one can visualize two distinct possible classes of reasons why blood sugar increases in response to brain injury. In the first, there is some impairment in the glucose regulation mechanism. In the second, the requirement for glucose is increased during the wound healing process, and hyperglycemia is a mechanism of the body to meet the demand. It is known that infections such as meningitis, stroke, or hemorrhage lead to transient hyperglycemia. Such hyperglycemia is often accompanied by lower BG levels [52][130][15]. In such cases, hypoglycemic treatments can be counterproductive and result in a delayed or derailed repair process. This is a likely reason why, in patients under intensive care, strict regulation of plasma glucose increased mortality instead of the expected decrease [60].

When hyperglycemia is a process of meeting increased glucose demand or compensating for subnormal transport, removal of plasma glucose by any means is expected to increase L. This is observed to happen when SGLT2 inhibitors decrease the urinary threshold and drive out plasma glucose. If increasing L is not sufficient to restore the required glucose level, there is a shift to ketogenesis since the brain can use keto acids as an alternative source of energy. Therefore, increased L with or without increased ketogenesis is expected by our model following treatment with SGLT2 inhibitors [13].

### B. Glucose tolerance curve

Unlike the steady-state predictions, the patterns of the post-meal glucose curve by our model are not qualitatively different from classical models. The area under the curve, height of the peak, time required to return to steady-state, and the time difference between glucose and insulin peaks are increased by decreased  $I_5$ ,  $K_5$ , or  $K_8$  individually or in combination. A decrease in  $I_5$  or  $K_5$  does not increase FG but changes the shape of the glucose tolerance curve. A decreased  $K_8$  may alter both simultaneously. The altered curve shows the three typical features, namely a taller peak, delayed return to steady-state, and a longer gap between the glucose peak and the insulin peak [5]. This result is not unique to our model, and classical models also show the three features. The classical model results in simultaneous and proportionate alterations in both fasting and post-meal glucose and therefore fails to explain the NFG-IGT state. In our model, decreasing  $I_5$  or  $K_7$  without a change in  $K_8$  results in an NFG-IGT state [6].

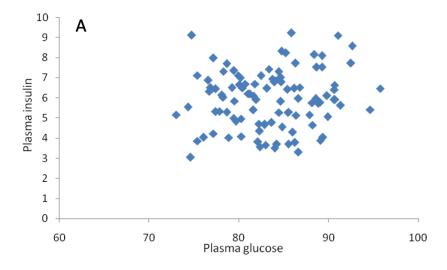
# C. Population simulations

For both the steady-state and post-glucose load dynamics, we give population distributions to  $K_1$ ,  $K_5$ ,  $K_8$ ,  $K_9$ ,  $I_S$  and also incorporate normally distributed error in glucose and insulin measurements in fasting and post-meal sampling. We also incorporate correlated changes in  $K_1$ ,  $K_8$ , and  $K_9$ , which are expected as a result of hypo-vascularization in the brain. These simulations are run to observe whether we get the anomalous correlations in the fasting versus post-meal state and in the HOMA indices as observed empirically [131][5].

By classical models, the regression correlation parameters of the glucose-insulin relationship are not different in the fasting state versus post-glucose load, although the range of variables is different, as shown previously by Diwekar-Joshi and Watve [5]. Also, if we assume HOMA-IR to faithfully reflect insulin resistance and HOMA  $\beta$  to faithfully represent  $\beta$  cell response, then there is no reason why the two indices should be correlated.

The assumption behind our model that there are different mechanisms at work under fasting versus post-glucose load conditions is necessary to explain the large difference in the regression correlation parameters in fasting versus post-meal levels. If the same set of mechanisms in the fasting and post-meal conditions are operational, whatever the model used, it is imperative that fasting correlation regression parameters are stronger or comparable to post-meal parameters. Simulations with our model are able to give a poor FG-FI correlation along with a strong post-meal glucose-insulin correlation under multiple conditions [16] (Figure 4). When the variance in  $K_8$  and  $K_9$  is small but that in  $K_1$ ,  $K_5$ , Gt, and  $I_5$  individually or in combination is large, the fasting correlations are weak and post-meal correlations are strong; the post-meal glucose-insulin regression slope is substantially greater than the FG-FI slope. Also, whenever the FG-FI correlation is weak, the indices HOMA-IR and HOMA  $\beta$  are strongly correlated, similar to the epidemiological data [17]. The difference between fasting and post-meal regression correlation patterns is not predicted by the classical models and is unique to

our model, which assumes a different set of regulatory mechanisms in the fasting and postmeal states.



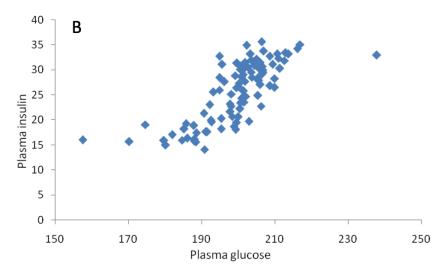
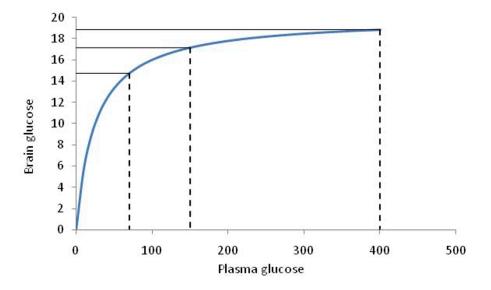


Figure 4. Plasma glucose and insulin correlation in population simulation data in the fasting steady-state (A) and post-meal (B) condition. The empirical finding that there is a strong positive correlation in the post-meal data but poor correlation in the fasting steady-state was possible over a large parameter space in our model. Depicted here are the simulation results (post-meal  $R^2$ =0.57, steady-state  $R^2$ =0.014) in which only  $K_1$  was given a wider population distribution. (mean (S.D.))  $K_1$ =35(7),  $K_5$ =10(0.00001),  $K_8$ =5(0.000005), and  $K_9$ =0.8(0.000008). Other parameters ( $K_{Im}$ =50,  $K_{2m}$ =15,  $K_2$ =0.05,  $d_1$ =0.15,  $I_s$ =1, BGt=25, BI=4.5,  $K_{8m}$ =80,  $K_{10}$ =0.1).

### D. Effect of glucose normalization on arresting diabetic complications and mortality

By classical thinking, chronic hyperglycemia is responsible for diabetic complications, and preventing hyperglycemia should arrest complications. In contrast, the thought behind our model is that vascular problems are primary, which alter the rate of glucose-insulin transport to the brain, and hyperglycemia is an offshoot symptom that may not be causal to diabetic complications. The diabetic complications can arise directly from the vasculopathy. Therefore, controlling glucose may not have any effect on diabetic complications [18]. On the other hand, forcefully reducing plasma sugar without addressing vascular problems can create more severe glucose deficiency in the brain and other organs, thereby turning counterproductive. Because

of the saturating dynamics of transport, a curvilinear relationship is expected between plasma glucose and brain glucose in such a way that a moderate reduction in hyperglycemia will change brain glucose availability marginally, whereas tight glucose regulation can have a disproportionately larger effect (Figure 5). Therefore, tight glucose regulation may increase mortality and other adverse outcomes. This prediction is compatible with some of the tight versus moderate control clinical trials, including ACCORD, NICE sugar trial, and UGDP [18].

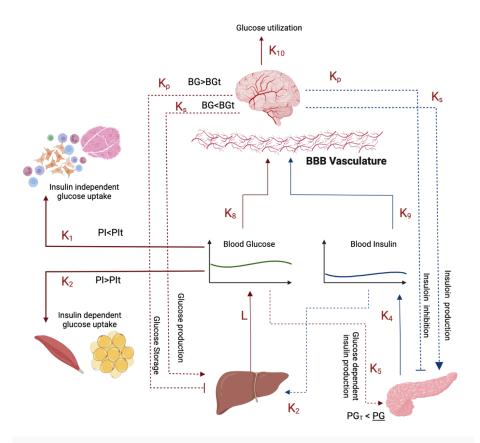


**Figure 5.** Effect of glucose lowering on brain glucose availability. For example, a reduction in FG from 400 mg/dl to 150 mg/dl corresponds to a decrease in BG by 1.68 mg/dl, but a further reduction from 150 to 70 mg/dl reduces BG by 2.41 mg/dl. Therefore, moderate sugar control may not affect brain glucose supply drastically, but tight sugar control is expected to affect it more seriously.

In a nutshell, our model explains all of the 19 patterns a model needs to explain. No other model has attempted this diverse task. And the classical models have clearly failed to be compatible with many of them.

# 4. Discussion

The main inference from our model, stated most conservatively, is that a brain-centered model can potentially explain most of the anomalies faced by peripheral models and therefore needs greater attention (Figure 6). If supported well, by exploring its testable predictions suggested below and possibly more, it has the potential to bring in a radical change in the fundamental view as well as clinical practice of T2DM.



**Figure 6.** Effect of changes in vasculature in the blood-brain barrier. The model predicts that glucose and insulin transportation to the brain has a major role in the development of diabetic symptoms compared to other peripheral changes. The blood-brain barrier vasculature hence may hold the key to understanding the shift from a normal to a diabetic condition. (created using BioRender)

### <u>Testability of assumptions and additional predictions:</u>

The assumption that the thresholds *PGT* and *PIT* can be modified by autonomous signaling needs to be tested empirically. Although currently the thresholds are known to be flexible, information about the conditions and mechanisms of change is poorly known. Our assumption that the parameters of the saturation equation for glucose and insulin transport to the brain are different and that under normal conditions insulin transport is closer to saturation than glucose transport can be tested with carefully worked-out kinetics of the two transport mechanisms.

The assumption of our model that reduced transport of glucose and insulin to the brain is the primary pathology of T2DM, leading secondarily to hyperglycemia, makes more predictions that can be tested experimentally or epidemiologically. Experimentally, specifically blocking GLUT1 receptors in the brain should lead to hyperglycemia. Conversely, infusion of glucose directly into the brain should reduce peripheral hyperglycemia in the short run. This is already suggested by some experiments  $^{[132][66]}$ . It is also demonstrated that inducing primary endothelial dysfunction and reduced glucose transport to the brain by endothelial deletion of the hypoxia-inducible factor HIF-1 $\alpha$  results in hyperglycemia  $^{[117]}$ . More careful research in this direction to reveal the cause-effect relationship between vascular defects, brain glucose levels, and plasma glucose levels will be enlightening. Epidemiologically, hypoglycemia is shown to be associated with dementia and other neuronal problems  $^{[133][134][135][136]}$ , and tight glycemic control led to higher mortality as compared to moderate control in many of the clinical trials  $^{[56]}$   $^{[58][60]}$ , which demands investigations into the causal pathways. The question of whether tight glycemic control leads to subtle neuronal changes in the long run, as expected by our model,

needs careful investigation. The difference between fasting and post-meal regression correlation parameters between glucose and insulin is an important epidemiological line of evidence we have used. Chawla et al. (2017) and Diwekar-Joshi and Watve<sup>[5]</sup> showed this pattern across four different data sets. How generalized the pattern is needs to be tested in multiple population studies.

On the modeling front, it is necessary to undertake a comparative evaluation of the different models with respect to the battery of predictions that we listed here. Perhaps a few more predictions may be added. However, at present, many of the models and their possible modifications are not explored sufficiently to see whether they can explain the currently unexplained patterns under certain sets of conditions. A model prediction matrix would be an appropriate approach for such a comparative evaluation, but we may have to wait until all alternative models are explored sufficiently and elaborately on which of the empirical patterns they predict, which ones they are compatible with, and which ones they contradict. We have shown here that the brain-centered model predicts or is compatible with all the patterns listed in the introduction and does not contradict any.

# **Possible causes of T2DM:**

The classically believed causal factors, namely insulin resistance and  $\beta$  cell dysfunction, are not compatible with many of the empirical findings as shown by Diwekar-Joshi and Watve<sup>[5]</sup>. In our model, changes in insulin resistance and  $\beta$  cell dysfunction were neither necessary nor sufficient to account for all the patterns. Nevertheless, they were helpful in accounting for the altered glucose tolerance curve, although other factors could also account for it independent of insulin resistance. By incorporating insulin resistance in our model and assuming it to work only in the post-feeding state, patterns 1, 2, 5, 6, 9, 16, and 17 could be explained, but not others. In short, our model does not rule out insulin resistance as a phenomenon, but implies that it may not be central to T2DM. Primary vasculature defects reducing glucose and insulin transport in a mutually correlated or uncorrelated manner could explain all patterns and therefore make the most parsimonious causal hypothesis. Particularly, assuming that altered vascular function affects  $K_1$  early, followed by  $K_8$  and  $K_9$ , is sufficient to explain all the patterns without alteration in  $I_5$  or any other factor. The apparent  $\beta$  cell dysfunction is an inevitable effect of a higher degree of vascular dysfunction and altered autonomic signals and therefore may not be needed as an independent causal factor.

Being open to alternative possibilities is an important virtue of science, and it is particularly important with the limited clinical success of prevalent thinking along with mounting anomalous findings. Prevention of T2DM on a global scale has largely failed, and treatment has had limited and inconsistent success in arresting mortality and morbidities associated with T2DM[137][138][139][57]. Therefore, exploring alternative possible interpretations is a need of the time.

Triggered by multiple anomalies in the theory of glucose homeostasis and the origins of diabetic hyperglycemia, we have articulated here an alternative paradigm that potentially resolves in a logically and mathematically consistent manner all of the anomalous findings. Being mathematically and logically sound and compatible with evidence is not sufficient proof of a theory but certainly reflects on its potential to develop into a new alternative paradigm. For a large field such as T2DM, multiple efforts would be needed to evaluate competing paradigms. We have suggested a few more testable predictions that can help in this task. There can be more possible ways of testing them, which should come to light and be used to reach robust conclusions that have the potential to change the clinical course of prevention as well as treatment of an important global health problem.

If our proposed causal inference that vascular dysfunction is primary to T2DM is supported by more careful investigations, it is necessary and might even be sufficient to lead to all the observed symptoms and patterns. It explains the failure of glucose-normalizing treatment in arresting diabetic complications. Simultaneously, it suggests alternative lines of treatment that should focus on normalizing vascular and neuronal function rather than focusing on glucose normalization. The deficiencies of stimuli normally required for growth factors and other angiogenic and neuroprotective factors can be forecasted as the best bet for the new approach. However, rigorous efforts are needed to strengthen the evidence base for selecting the right one among the alternative paradigms.

Clinically, glucose normalization as a treatment, which has already failed empirically, fails theoretically as well with the success of our model. Therefore, this target of treatment needs to be given up completely. No drug has been developed so far to effectively normalize vasculature. Since a large number of signals govern the angiogenesis process, a single-molecule approach typical of pharmacology research is unlikely to work. Exercise and fitness interventions have been largely successful in arresting T2DM, its complications, and mortality independent of weight loss and sugar control [140][141][142][143]. This stands in contrast with the failure of glucose normalization treatment in reducing complications and mortality [144], but more focused work is needed. We still do not understand the mechanisms by which the weight- and sugar-independent effects of exercise work. They are likely to work through growth factor activation [82][145][146]. There is also inadequate understanding of how different types of exercise exert differential effects on physiology [147]. Refining this line of treatment appears to have greater promise for the future.

# **Ethics Statement**

This study was based on mathematical modeling and analysis of previously published data. No new experiments involving human participants or animals were performed by the authors; therefore, institutional ethical approval was not required.

# **Informed Consent Statement**

This study did not involve direct participation of human subjects; informed consent was not applicable, as the study utilized mathematical modeling and publicly available or previously published data.

# **Data Availability Statement**

The mathematical model equations are described in the Methods section. Further inquiries regarding simulation code or specific parameter sets used can be directed to the corresponding author.

# **Author Contributions**

Conceptualization: A.O. and M.W.; Methodology: A.O. and M.W.; Software: A.O.; Formal Analysis: A.O.; Investigation: A.O.; Writing — Original Draft Preparation: M.W.; Writing — Review & Editing: A.O. and M.W.

# References

- △DeFronzo RA, Ferrannini E, Groop L, Henry RR, Herman WH, Holst JJ, Hu FB, Kahn CR, Raz I, Sh ulman GI, Simonson DC, Testa MA, Weiss R (2015). "Type 2 diabetes mellitus." Nature Reviews Di sease Primers. 1(1):15019. doi:10.1038/nrdp.2015.19.
- △Bernard C (1859). Lessons on the phenomena of life. General physiology course of the Museum o f Natural History. Paris: Librairie Delagrave.
- ∆Banting FG, Best CH, Collip JB, Campbell WR, Fletcher AA (1922). "Pancreatic Extracts in the Tre atment of Diabetes Mellitus." Canadian Medical Association Journal. 12(3):141–146. PMID <u>203140</u> <u>60</u>.
- 4. a. Lundqvist MH, Almby K, Abrahamsson N, Eriksson JW (2019). "Is the Brain a Key Player in Glu cose Regulation and Development of Type 2 Diabetes?" Frontiers in Physiology. **10**:457. doi:10.338 9/fphys.2019.00457.
- 5. <u>a. b. c. d. e. f. g. h. i. j. k. l. m. n. o. P. Diwekar-Joshi M, Watve M (2020)</u>. "Driver versus navigator causat ion in biology: The case of insulin and fasting glucose." PeerJ. 8:e10396. doi:10.7717/peerj.10396.
- 6. ∆Blüher M, Michael MD, Peroni OD, Ueki K, Carter N, Kahn BB, Kahn CR (2002). "Adipose tissue se lective insulin receptor knockout protects against obesity and obesity-related glucose intoleranc e." Developmental Cell. 3(1):25–38. doi:10.1016/S1534-5807(02)00199-5.
- 7. ∆Kadowaki T (2000). "Insights into insulin resistance and type 2 diabetes from knockout mouse models." Journal of Clinical Investigation. 106(4):459–465. <a href="http://www.ncbi.nlm.nih.gov/pmc/arti">http://www.ncbi.nlm.nih.gov/pmc/arti</a>

### cles/PMC380257/.

- Costes S, Butler PC (2014). "Insulin-Degrading Enzyme Inhibition, a Novel Therapy for Type 2 D iabetes?" Cell Metabolism. 20(2):201–203. doi:10.1016/j.cmet.2014.07.016.
- Maianti JP, McFedries A, Foda ZH, Kleiner RE, Du XQ, Leissring MA, Tang WJ, Charron MJ, Seelig er MA, Saghatelian A, Liu DR (2014). "Anti-diabetic activity of insulin-degrading enzyme inhibit ors mediated by multiple hormones." Nature. 511(7507):94–98. doi:10.1038/nature13297.
- 10. △Giustina A, Girelli A, Buffoli MG, Cimino A, Legati F, Valentini U, Giustina G (1991). "Low-dose oc treotide is able to cause a maximal inhibition of the glycemic responses to a mixed meal in obese type 2 diabetic patients treated with insulin." Diabetes Research and Clinical Practice. 14(1):47–5 4. doi:10.1016/0168-8227(91)90052-F.
- 11. △Lamberts SWJ, Hofland LJ (2019). "ANNIVERSARY REVIEW: Octreotide, 40 years later." Europea n Journal of Endocrinology. **181**(5):R173–R183. doi:<u>10.1530/EJE-19-0074</u>.
- 12. <sup>Δ</sup>Leahy JL, Bumbalo LM, Chen C (1994). "Diazoxide causes recovery of beta-cell glucose responsi veness in 90% pancreatectomized diabetic rats." Diabetes. **43**(2):173–179. doi:10.2337/diabetes.43. 2.173.
- 13. Amatsuda M, Kawasaki F, Mikami Y, Takeuchi Y, Saito M, Eto M, Kaku K (2002). "Rescue of betacell exhaustion by diazoxide after the development of diabetes mellitus in rats with streptozotoci n-induced diabetes." European Journal of Pharmacology. 453(1):141–148. doi:10.1016/S0014-2999 (02)02389-0.
- 14. <sup>a, b</sup>Mehran AE, Templeman NM, Brigidi GS, Lim GE, Chu KY, Hu X, Botezelli JD, Asadi A, Hoffman BG, Kieffer TJ, Bamji SX, Clee SM, Johnson JD (2012). "Hyperinsulinemia drives diet-induced obesi ty independently of brain insulin production." Cell Metabolism. 16(6):723–737. doi:10.1016/j.cmet.2012.10.019.
- 15. △Templeman NM, Clee SM, Johnson JD (2015). "Suppression of hyperinsulinaemia in growing fe male mice provides long-term protection against obesity." Diabetologia. **58**(10):2392–2402. doi:10.1007/s00125-015-3676-7.
- 16. <sup>a, b, c, d</sup>Matthews DR, Hosker JP, Rudenski AS, Naylor BA, Treacher DF, Turner RC (1985). "Homeos tasis model assessment: insulin resistance and β-cell function from fasting plasma glucose and i nsulin concentrations in man." Diabetologia. 28(7):412–419. doi:10.1007/BF00280883.
- 17. a, b, c, d Gaohua L, Kimura H (2009). "A mathematical model of brain glucose homeostasis." Theor etical Biology and Medical Modelling. 6(1):26. doi:10.1186/1742-4682-6-26.
- 18. <u>a, b, c, d, e, f,</u> gWatve M (2013). Doves, Diplomats, and Diabetes. New York: Springer New York. doi:<u>1</u> 0.1007/978-1-4614-4409-1.
- 19. <sup>△</sup>Shinde S, Ranade P, Watve M (2021). "Evaluating alternative hypotheses to explain the downwa rd trend in the indices of the COVID-19 pandemic death rate." PeerJ. 9:e11150. doi:10.7717/peerj.1115 <u>0</u>.
- 20. <sup>△</sup>Thakar JD, Kunte K, Chauhan AK, Watve AV, Watve MG (2003). "Nectarless flowers: ecological c orrelates and evolutionary stability." Oecologia. **136**(4):565–570. doi:10.1007/s00442-003-1304-6.
- 21. ^Vibishan B, Watve MG (2020). "Context-dependent selection as the keystone in the somatic evol ution of cancer." Scientific Reports. 10(1):4223. doi:10.1038/s41598-020-61046-7.
- 22. \(^AWatve M\), Diwekar-Joshi M (2016). "What to expect from an evolutionary hypothesis for a huma n disease: The case of type 2 diabetes." HOMO. 67(5):349–368. doi:10.1016/j.jchb.2016.07.001.
- 23. ^Lerner RL, Porte D Jr (1972). "Acute and steady-state insulin responses to glucose in nonobese di abetic subjects." Journal of Clinical Investigation. 51(7):1624–1631. doi:10.1172/JCI106963.
- 24. <sup>a. <u>b</u></sup>Turner RC, Holman RR, Matthews D, Hockaday TDR, Peto J (1979). "Insulin deficiency and ins ulin resistance interaction in diabetes: Estimation of their relative contribution by feedback anal ysis from basal plasma insulin and glucose concentrations." Metabolism. 28(11):1086–1096. doi:10.1016/0026-0495(79)90146-X.
- 25. Corkey BE, Shirihai O (2012). "Metabolic master regulators: sharing information among multiple systems." Trends in Endocrinology & Metabolism. 23(12):594–601. doi:10.1016/j.tem.2012.07.006.
- 26. △Pories WJ, Dohm GL (2020). "Letter to the Editor Re: Sera of Obese Type 2 Diabetic Patients Und ergoing Metabolic Surgery Instead of Conventional Treatment Exert Beneficial Effects on Beta C ell Survival and Function: Results of a Randomized Clinical Study." Obesity Surgery. 30(9):3603–3604. doi:10.1007/s11695-020-04665-3.

- 27. AShanik MH, Xu Y, Skrha J, Dankner R, Zick Y, Roth J (2008). "Insulin Resistance and Hyperinsulin emia: Is hyperinsulinemia the cart or the horse?" Diabetes Care. 31(Suppl 2):S262–S268. doi:10.23 37/dc08-s264.
- 28. <sup>△</sup>Weyer C, Hanson RL, Tataranni PA, Bogardus C, Pratley RE (2000). "A high fasting plasma insul in concentration predicts type 2 diabetes independent of insulin resistance: evidence for a pathog enic role of relative hyperinsulinemia." Diabetes. 49(12):2094–2101. doi:10.2337/diabetes.49.12.20 94.
- 29. <sup>A</sup>Wiebe N, Ye F, Crumley ET, Bello A, Stenvinkel P, Tonelli M (2021). "Temporal Associations Amo ng Body Mass Index, Fasting Insulin, and Systemic Inflammation." JAMA Network Open. 4(3):e21 1263. doi:10.1001/jamanetworkopen.2021.1263.
- Dungan KM, Braithwaite SS, Preiser JC (2009). "Stress hyperglycaemia." The Lancet. 373(9677): 1798–1807. doi:10.1016/S0140-6736(09)60553-5.
- 31. \(^{\text{Coggan AR (1991)}}\). "Plasma Glucose Metabolism During Exercise in Humans." Sports Medicine. 1 1(2):102–124. doi:\(\frac{10.2165/00007256-199111020-00003}\).
- 32. △Marliss EB, Vranic M (2002). "Intense Exercise Has Unique Effects on Both Insulin Release and It s Roles in Glucoregulation: Implications for Diabetes." Diabetes. 51(Suppl 1):S271–S283. doi:10.233 7/diabetes.51.2007.S271.
- 33. △Gjedde A, Crone C (1981). "Blood-Brain Glucose Transfer: Repression in Chronic Hyperglycemia." Science. 214 (4519):456–457. doi:10.1126/science.7027439.
- 34. ^Cornford EM, Hyman S, Cornford ME, Clare-Salzler M (1995). "Down-regulation of blood-brain glucose transport in the hyperglycemic nonobese diabetic mouse." Neurochemical Research. 20 (7):869–873. doi:10.1007/BF00969700.
- 35. ^Pardridge WM, Triguero D, Farrell CR (1990). "Downregulation of Blood-Brain Barrier Glucose T ransporter in Experimental Diabetes." Diabetes. 39(9):1040–1044. doi:10.2337/diab.39.9.1040.
- 36. <sup>a, b, c</sup>Hwang JJ, Jiang L, Hamza M, Sanchez Rangel E, Dai F, Belfort-DeAguiar R, Parikh L, Koo BB, Rothman DL, Mason G, Sherwin RS (2017). "Blunted rise in brain glucose levels during hyperglyc emia in adults with obesity and T2DM." JCI Insight. 2(20). doi:10.1172/jci.insight.95913.
- 37. △Höppener JWM, Oosterwijk C, Nieuwenhuis MG, Posthuma G, Thijssen JHH, Vroom TM, Ahrén B, Lips CJM (1999). "Extensive islet amyloid formation is induced by development of Type II diabete s mellitus and contributes to its progression: pathogenesis of diabetes in a mouse model." Diabet ologia. 42(4):427–434. doi:10.1007/s001250051175.
- 38. Porte D Jr, Kahn SE (1989). "Hyperproinsulinemia and Amyloid in NIDDM: Clues to Etiology of Is let β-Cell Dysfunction?" Diabetes. **38**(11):1333–1336. doi:10.2337/diab.38.11.1333.
- 39. a, b, c, dWatve M, Bodas A, Diwekar M (2014). "Altered autonomic inputs as a cause of pancreatic β-cell amyloid." Medical Hypotheses. **82**(1):49–53. doi:10.1016/j.mehy.2013.11.002.
- 40.  $\triangle$ Bacha F, Gungor N, Lee S, Arslanian SA (2013). "Progressive deterioration of  $\beta$ -cell function in ob ese youth with type 2 diabetes." Pediatric Diabetes. **14**(2):106–111. doi:10.1111/j.1399-5448.2012.00 915.x.
- 41.  $\triangle$ Butler AE, Janson J, Bonner-Weir S, Ritzel R, Rizza RA, Butler PC (2003). " $\beta$ -Cell Deficit and Incre ased  $\beta$ -Cell Apoptosis in Humans With Type 2 Diabetes." Diabetes. 52(1):102–110. doi:10.2337/diabetes.52.1.102.
- 42. ^Clark A, Jones LC, de Koning E, Hansen BC, Matthews DR (2001). "Decreased insulin secretion in type 2 diabetes: a problem of cellular mass or function?" Diabetes. 50(Suppl 1):S169-S171. doi:10.2 337/diabetes.50.2007.S169.
- 43. △Porte D Jr, Kahn SE (2001). "Beta-cell dysfunction and failure in type 2 diabetes: potential mech anisms." Diabetes. **50**(Suppl 1):S160–S163. doi:<u>10.2337/diabetes.50.2007.S160</u>.
- 44. <sup>△</sup>Limenta M, Ho CSC, Poh JWW, Goh SY, Toh DSL (2019). "Adverse Drug Reaction Profile of SGLT2 Inhibitor-Associated Diabetic Ketosis/Ketoacidosis in Singapore and their Precipitating Factors." Clinical Drug Investigation. 39(7):683–690. doi:10.1007/s40261-019-00794-5.
- 45. △Mistry S, Eschler DC (2021). "Euglycemic Diabetic Ketoacidosis Caused by SGLT2 Inhibitors and a Ketogenic Diet: A Case Series and Review of Literature." AACE Clinical Case Reports. 7(1):17–19. doi:10.1016/j.aace.2020.11.009.
- 46. <sup>△</sup>Op den Kamp YJM, de Ligt M, Dautzenberg B, Kornips E, Esterline R, Hesselink MKC, Hoeks J, Schrauwen-Hinderling VB, Havekes B, Oscarsson J, Phielix E, Schrauwen P (2021). "Effects of the SG LT2 Inhibitor Dapagliflozin on Energy Metabolism in Patients With Type 2 Diabetes: A Randomi zed, Double-Blind Crossover Trial." Diabetes Care. 44(6):1334–1343. doi:10.2337/dc20-2887.

- 47. △Pfützner A, Klonoff D, Heinemann L, Ejskjaer N, Pickup J (2017). "Euglycemic ketosis in patients with type 2 diabetes on SGLT2-inhibitor therapy—an emerging problem and solutions offered by diabetes technology." Endocrine. 56(1):212–216. doi:10.1007/s12020-017-1264-y.
- 48. ^Campbell IW (1976). "The somogyi phenomenon. A short review." Acta Diabetologica Latina. 13 (1–2):68–73. doi:10.1007/BF02591583.
- 49. <sup>△</sup>Reyhanoglu G, Rehman A (2021). Somogyi Phenomenon. StatPearls Publishing.
- 50. △Shi J, Dong B, Mao Y, Guan W, Cao J, Zhu R, Wang S (2016). "Review: Traumatic brain injury and hyperglycemia, a potentially modifiable risk factor." Oncotarget. 7(43):71052–71061. doi:10.18632/oncotarget.11958.
- 51. <sup>Δ</sup>Krishnan S, Chang AC, Stoltz BM, Prasadarao NV (2016). "Escherichia coli K1 Modulates Peroxis ome Proliferator—Activated Receptor γ and Glucose Transporter 1 at the Blood-Brain Barrier in N eonatal Meningitis." Journal of Infectious Diseases. 214(7):1092–1104. doi:10.1093/infdis/jiw306.
- 52. <sup>a.</sup> <sup>b</sup>Schut ES, Westendorp WF, de Gans J, Kruyt ND, Spanjaard L, Reitsma JB, van de Beek D (2009). "Hyperglycemia in bacterial meningitis: a prospective cohort study." BMC Infectious Diseases. 9 (1):57. doi:10.1186/1471-2334-9-57.
- 53. △Diabetes Prevention Program Research Group (2015). "Long-term effects of lifestyle intervention nor metformin on diabetes development and microvascular complications over 15-year follow-up: the Diabetes Prevention Program Outcomes Study." The Lancet Diabetes & Endocrinology. 3 (11):866–875. doi:10.1016/S2213-8587(15)00291-0.
- 54. Ferrannini E, DeFronzo RA (2015). "Impact of glucose-lowering drugs on cardiovascular disease in type 2 diabetes." European Heart Journal. **36**(34):2288–2296. doi:10.1093/eurheartj/ehv239.
- 55. ∆King P, Peacock I, Donnelly R (1999). "The UK prospective diabetes study (UKPDS): clinical and t herapeutic implications for type 2 diabetes." British Journal of Clinical Pharmacology. 48(5):643 –648. doi:10.1046/j.1365-2125.1999.00092.x.
- 56. a. <u>b</u>Klein R (2010). "Intensive treatment of hyperglycaemia: ACCORD." The Lancet. **376**(9739):391–392. doi:10.1016/S0140-6736(10)61028-8.
- 57. <sup>a, b</sup>Lee CG, Heckman-Stoddard B, Dabelea D, Gadde KM, Ehrmann D, Ford L, et al (2021). "Effect o f Metformin and Lifestyle Interventions on Mortality in the Diabetes Prevention Program and Di abetes Prevention Program Outcomes Study." Diabetes Care. 44(12):2775–2782. doi:10.2337/dc21-1046.
- 58. <sup>a, b</sup>Schwartz TB, Meinert CL (2004). "The UGDP Controversy: thirty-four years of contentious am biguity laid to rest." Perspectives in Biology and Medicine. **47**(4):564–574. doi:10.1353/pbm.2004. 0071.
- 59. <sup>△</sup>Turner R, Matthews D, Neil A, McElroy H (1998). "Tight blood pressure control and risk of macro vascular and microvascular complications in type 2 diabetes: UKPDS 38." BMJ. 317:703–713. doi:1 0.1136/bmj.317.7160.703.
- 60. <sup>a, b, c</sup>The NICE-SUGAR Study Investigators (2009). "Intensive versus Conventional Glucose Contr ol in Critically Ill Patients." New England Journal of Medicine. **360**(13):1283–1297. doi:10.1056/NEJ Moa0810625.
- 61. ^UK Prospective Diabetes Study (UKPDS) Group (1998). "Intensive blood-glucose control with sul phonylureas or insulin compared with conventional treatment and risk of complications in patie nts with type 2 diabetes (UKPDS 33)." Lancet. 352(9131):837–853.
- 62. <sup>△</sup>Laeger T, Baumeier C, Wilhelmi I, Würfel J, Kamitz A, Schürmann A (2017). "FGF21 improves glu cose homeostasis in an obese diabetes-prone mouse model independent of body fat changes." Di abetologia. 60(11):2274–2284. doi:10.1007/s00125-017-4389-x.
- 63. <sup>a, b, c</sup>Ajmera I, Swat M, Laibe C, Le Novère N, Chelliah V (2013). "The impact of mathematical mo deling on the understanding of diabetes and related complications." CPT: Pharmacometrics & Sy stems Pharmacology. 2(7):1-14. doi:10.1038/psp.2013.30.
- 64. a. b. c. dMari A, Tura A, Grespan E, Bizzotto R (2020). "Mathematical Modeling for the Physiologic al and Clinical Investigation of Glucose Homeostasis and Diabetes." Frontiers in Physiology. 11:15 48. https://www.frontiersin.org/article/10.3389/fphys.2020.575789.
- 65. <sup>△</sup>Lam TKT, Gutierrez-Juarez R, Pocai A, Rossetti L (2005). "Regulation of Blood Glucose by Hypot halamic Pyruvate Metabolism." Science. **309**(5736):943–947. doi:10.1126/science.1112085.
- 66. <sup>a, b</sup>Osundiji MA, Lam DD, Shaw J, Yueh CY, Markkula SP, Hurst P, Colliva C, Roda A, Heisler LK, Ev ans ML (2012). "Brain Glucose Sensors Play a Significant Role in the Regulation of Pancreatic Glu cose-Stimulated Insulin Secretion." Diabetes. 61(2):321–328. doi:10.2337/db11-1050.

- 67. <sup>Δ</sup>Perrin C, Knauf C, Burcelin R (2004). "Intracerebroventricular Infusion of Glucose, Insulin, and t he Adenosine Monophosphate-Activated Kinase Activator, 5-Aminoimidazole-4-Carboxamide-1-β-d-Ribofuranoside, Controls Muscle Glycogen Synthesis." Endocrinology. **145**(9):4025–4033. do i:10.1210/en.2004-0270.
- 68. △Deem JD, Muta K, Scarlett JM, Morton GJ, Schwartz MW (2017). "How Should We Think About th e Role of the Brain in Glucose Homeostasis and Diabetes?" Diabetes. 66(7):1758–1765. doi:10.2337/ dbi16-0067.
- 69. <sup>△</sup>Güemes A, Georgiou P (2018). "Review of the role of the nervous system in glucose homoeostasi s and future perspectives towards the management of diabetes." Bioelectronic Medicine. **4**:9. doi: 10.1186/s42234-018-0009-4.
- 70. <sup>△</sup>Brown JM, Scarlett JM, Schwartz MW (2019). "Rethinking the role of the brain in glucose homeo stasis and diabetes pathogenesis." J Clin Invest. **129**(8):3035-3037. doi:10.1172/JCI130904. PMID 31 329157.
- 71. \(^{Alonge KM}\), D'Alessio DA, Schwartz MW (2021). "Brain control of blood glucose levels: implicatio ns for the pathogenesis of type 2 diabetes." Diabetologia. 64(1):5-14. doi:\(\frac{10.1007/s00125-020-052}{93-3}\). PMID \(\frac{33043401}{2}\).
- 72. <sup>△</sup>MacDonald AJ, Yang YHC, Cruz AM, Beall C, Ellacott KLJ (2021). "Brain-Body Control of Glucose Homeostasis—Insights From Model Organisms." Frontiers in Endocrinology. 12:662769. doi:10.33 89/fendo.2021.662769.
- 73. ^Choi JH, Kim MS (2022). "Homeostatic Regulation of Glucose Metabolism by the Central Nervou s System." Endocrinology and Metabolism. 37(1):9–25. doi:10.3803/EnM.2021.1364.
- 74. ^Bentsen MA, Mirzadeh Z, Schwartz MW (2019). "Revisiting How the Brain Senses Glucose—And Why." Cell Metabolism. 29(1):11–17. doi:10.1016/j.cmet.2018.11.001.
- 75. △Büsing KA, Schönberg SO, Brade J, Wasser K (2013). "Impact of blood glucose, diabetes, insulin, a nd obesity on standardized uptake values in tumors and healthy organs on 18F-FDG PET/CT." N uclear Medicine and Biology. 40(2):206–213. doi:10.1016/j.nucmedbio.2012.10.014.
- 76. a. bBoland BB, Rhodes CJ, Grimsby JS (2017). "The dynamic plasticity of insulin production in  $\beta$ -c ells." Molecular Metabolism. 6(9):958–973. doi:10.1016/j.molmet.2017.04.010.
- 77. <sup>a, b</sup>Thackeray JT, Beanlands RS, Dasilva JN (2012). "Altered sympathetic nervous system signalin g in the diabetic heart: emerging targets for molecular imaging." American Journal of Nuclear M edicine and Molecular Imaging. 2(3):314–334.
- 78. Amuris DMJ, Houben AJHM, Schram MT, Stehouwer CDA (2012). "Microvascular Dysfunction Is A ssociated With a Higher Incidence of Type 2 Diabetes Mellitus." Arteriosclerosis, Thrombosis, and Vascular Biology. 32(12):3082–3094. doi:10.1161/ATVBAHA.112.300291.
- 79. <sup>^</sup>Nguyen TT, Wang JJ, Wong TY (2007). "Retinal Vascular Changes in Pre-Diabetes and Prehyper tension." Diabetes Care. 30(10):2708–2715. doi:10.2337/dc07-0732.
- 80. <sup>△</sup>Stehouwer CDA (2018). "Microvascular Dysfunction and Hyperglycemia: A Vicious Cycle With W idespread Consequences." Diabetes. 67(9):1729–1741. doi:10.2337/dbi17-0044.
- 81. △Zaleska-Żmijewska A, Piątkiewicz P, Śmigielska B, Sokołowska-Oracz A, Wawrzyniak ZM, Rom aniuk D, et al (2017). "Retinal Photoreceptors and Microvascular Changes in Prediabetes Measur ed with Adaptive Optics (rtx1™): A Case-Control Study." Journal of Diabetes Research. 2017:41742 92. doi:10.1155/2017/4174292.
- 82. <sup>a, b</sup>Aloe L, Bracci-Laudiero L, Alleva E, Lambiase A, Micera A, Tirassa P (1994). "Emotional stress i nduced by parachute jumping enhances blood nerve growth factor levels and the distribution of nerve growth factor receptors in lymphocytes." Proceedings of the National Academy of Sciences. 91(22):10440–10444. doi:10.1073/pnas.91.22.10440.
- 83. <sup>Δ</sup>Cao L, Liu X, Lin EJD, Wang C, Choi EY, Riban V, Lin B, During MJ (2010). "Environmental and Ge netic Activation of a Brain-Adipocyte BDNF/Leptin Axis Causes Cancer Remission and Inhibitio n." Cell. 142(1):52–64. doi:10.1016/j.cell.2010.05.029.
- 84. ^Chodari L, Mohammadi M, Ghorbanzadeh V, Dariushnejad H, Mohaddes G (2016). "Testosteron e and Voluntary Exercise Promote Angiogenesis in Hearts of Rats with Diabetes by Enhancing Ex pression of VEGF-A and SDF-1a." Canadian Journal of Diabetes. 40(5):436–441. doi:10.1016/j.jcjd.2016.03.004.
- 85. Lakshmanan J (1986). "Aggressive behavior in adult male mice elevates serum nerve growth factor levels." American Journal of Physiology-Endocrinology and Metabolism. **250**(4):E386–E392. doi:10.1152/ajpendo.1986.250.4.E386.

- 86. <sup>△</sup>Nexø E, Olsen PS, Poulsen K (1984). "Exocrine and endocrine secretion of renin and epidermal g rowth factor from the mouse submandibular glands." Regulatory Peptides. 8(4):327–334. doi:10.1 016/0167-0115(84)90043-0.
- 87. <sup>△</sup>Nexø E, Hollenberg MD, Bing J (1981). "Aggressive behaviour in mice provokes a marked increas e in both plasma epidermal growth factor and renin." Acta Physiologica Scandinavica. 111(3):367 –371. doi:10.1111/j.1748-1716.1981.tb06749.x.
- 88. <sup>△</sup>Tirassa P, Triaca V, Amendola T, Fiore M, Aloe L (2003). "EGF and NGF injected into the brain of old mice enhance BDNF and ChAT in proliferating subventricular zone." Journal of Neuroscience Research. 72(5):557–564. doi:10.1002/jnr.10614.
- 89. ^Carnethon MR, Jacobs DR Jr, Sidney S, Liu K (2003). "Influence of Autonomic Nervous System Dy sfunction on the Development of Type 2 Diabetes: The CARDIA study." Diabetes Care. 26(11):3035 –3041. doi:10.2337/diacare.26.11.3035.
- 90. <sup>△</sup>Gray SM, Meijer RI, Barrett EJ (2014). "Insulin Regulates Brain Function, but How Does It Get Th ere?" Diabetes. **63**(12):3992–3997. doi:<u>10.2337/db14-0340</u>.
- 91. ∆Hasselbalch SG, Knudsen GM, Videbaek C, Pinborg LH, Schmidt JF, Holm S, Paulson OB (1999).

  "No effect of insulin on glucose blood-brain barrier transport and cerebral metabolism in human s." Diabetes. 48(10):1915–1921. doi:10.2337/diabetes.48.10.1915.
- 92. <sup>a, b</sup>Kern W, Peters A, Fruehwald-Schultes B, Deininger E, Born J, Fehm HL (2001). "Improving Influence of Insulin on Cognitive Functions in Humans." Neuroendocrinology. **74**(4):270–280. doi:10.1159/000054694.
- 93. <sup>a, b</sup>Shemesh E, Rudich A, Harman-Boehm I, Cukierman-Yaffe T (2012). "Effect of Intranasal Insuli n on Cognitive Function: A Systematic Review." The Journal of Clinical Endocrinology & Metabol ism. 97(2):366–376. doi:10.1210/jc.2011-1802.
- 94. <sup>^</sup>Strachan MWJ (2003). "Insulin and cognitive function." The Lancet. **362**(9392):1253. doi:10.1016/ <u>S0140-6736(03)14615-6</u>.
- 95. <sup>^</sup>Bingham EM, Hopkins D, Smith D, Pernet A, Hallett W, Reed L, Marsden PK, Amiel SA (2002). "T he Role of Insulin in Human Brain Glucose Metabolism." Diabetes. 51(12):3384–3390. doi:10.2337/diabetes.51.12.3384.
- 96. ARebelos E, Rinne JO, Nuutila P, Ekblad LL (2021). "Brain Glucose Metabolism in Health, Obesity, and Cognitive Decline—Does Insulin Have Anything to Do with It? A Narrative Review." Journal of Clinical Medicine. 10(7):1532. doi:10.3390/jcm10071532.
- 97. △Dakic T, Jevdjovic T, Lakic I, Ruzicic A, Jasnic N, Djurasevic S, Djordjevic J, Vujovic P (2023). "The Expression of Insulin in the Central Nervous System: What Have We Learned So Far?" Internatio nal Journal of Molecular Sciences. 24(7):6586. doi:10.3390/ijms24076586.
- 98. <sup>^</sup>Thorens B (2015). "GLUT2, glucose sensing and glucose homeostasis." Diabetologia. **58**(2):221–2 32. doi:10.1007/s00125-014-3451-1.
- 99. <sup>a, b</sup>Chen NG, Tassava TM, Romsos DR (1993). "Threshold for Glucose-Stimulated Insulin Secretion in Pancreatic Islets of Genetically Obese (ob/ob) Mice is Abnormally Low." The Journal of Nutritio n. 123(9):1567–1574. doi:10.1093/jn/123.9.1567.
- 100. <sup>a, b</sup>Henquin JC, Dufrane D, Nenquin M (2006). "Nutrient Control of Insulin Secretion in Isolated N ormal Human Islets." Diabetes. 55(12):3470–3477. doi:10.2337/db06-0868.
- 101. <sup>a, b</sup>Sorensen JT (1985). A physiologic model of glucose metabolism in man and its use to design a nd assess improved insulin therapies for diabetes. Thesis. Cambridge (MA): Massachusetts Instit ute of Technology.
- 102. <sup>a</sup>. <sup>b</sup>Begg DP, Woods SC (2013). "Interactions between the central nervous system and pancreatic is let secretions: a historical perspective." Advances in Physiology Education. **37**(1):53–60. doi:10.115/2/advan.00167.2012.
- 103. △Rohner-Jeanrenaud F, Jeanrenaud B (1983). "The central nervous system-endocrine pancreas ax is." Annales d'endocrinologie. 44(4):217–227.
- 104. <sup>△</sup>Strachan MWJ (2005). "Insulin and cognitive function in humans: experimental data and thera peutic considerations." Biochemical Society Transactions. 33(Pt 5):1037–1040. doi:10.1042/BST20 051037.
- 105. △Boado RJ, Black KL, Pardridge WM (1994). "Gene expression of GLUT3 and GLUT1 glucose trans porters in human brain tumors." Molecular Brain Research. 27(1):51–57. doi:10.1016/0169-328X(94)90183-X.

- 106. <sup>^</sup>Ge X, Chen C, Hui X, Wang Y, Lam KSL, Xu A (2011). "Fibroblast Growth Factor 21 Induces Glucos e Transporter-1 Expression through Activation of the Serum Response Factor/Ets-Like Protein-1 i n Adipocytes." Journal of Biological Chemistry. 286(40):34533−34541. doi:10.1074/jbc.M111.24859
  1.
- 107. <sup>^</sup>Liu M, Cao H, Hou Y, Sun G, Li D, Wang W (2018). "Liver Plays a Major Role in FGF-21 Mediated G lucose Homeostasis." Cellular Physiology and Biochemistry. **45**(4):1423–1433. doi:10.1159/000487568.
- 108. △Schüler R, Seebeck N, Osterhoff MA, Witte V, Flöel A, Busjahn A, Jais A, Brüning JC, Frahnow T, K abisch S, Pivovarova O, Hornemann S, Kruse M, Pfeiffer AFH (2018). "VEGF and GLUT1 are highly heritable, inversely correlated and affected by dietary fat intake: Consequences for cognitive function in humans." Molecular Metabolism. 11:129–136. doi:10.1016/j.molmet.2018.02.004.
- 109. ≜Baker LD, Cross DJ, Minoshima S, Belongia D, Watson GS, Craft S (2011). "Insulin resistance and Alzheimer-like reductions in regional cerebral glucose metabolism for cognitively normal adults with prediabetes or early type 2 diabetes." Archives of neurology. 68(1):51–57. doi:10.1001/archne urol.2010.225.
- 110. <sup>△</sup>Chau ACM, Cheung EYW, Chan KH, Chow WS, Shea YF, Chiu PKC, Mak HKF (2020). "Impaired c erebral blood flow in type 2 diabetes mellitus A comparative study with subjective cognitive de cline, vascular dementia and Alzheimer's disease subjects." NeuroImage: Clinical. 27:102302. doi: 10.1016/j.nicl.2020.102302.
- 111. ^Sundermann EE, Thomas KR, Bangen KJ, Weigand AJ, Eppig JS, Edmonds EC, et al. (2021). "Pred iabetes Is Associated With Brain Hypometabolism and Cognitive Decline in a Sex-Dependent Ma nner: A Longitudinal Study of Nondemented Older Adults." Frontiers in neurology. 12:551975. doi: 10.3389/fneur.2021.551975.
- 112. <sup>△</sup>Képes Z, Aranyi C, Forgács A, Nagy F, Kukuts K, Hascsi Z, Esze R, Somodi S, Káplár M, Varga J, E mri M, Garai I (2021). "Glucose-level dependent brain hypometabolism in type 2 diabetes mellitu s and obesity." European Journal of Hybrid Imaging. 5(1):3. doi:10.1186/s41824-021-00097-z.
- 113. ^Blázquez E, Hurtado-Carneiro V, LeBaut-Ayuso Y, Velázquez E, García-García L, Gómez-Oliver F, Ruiz-Albusac JM, Ávila J, Pozo MÁ (2022). "Significance of Brain Glucose Hypometabolism, Alte red Insulin Signal Transduction, and Insulin Resistance in Several Neurological Diseases." Frontie rs in Endocrinology. 13:873301. doi:10.3389/fendo.2022.873301.
- 114. △Park J, Choi S, Kim R (2023). "Association between prediabetes and cognitive function in Parkins on's disease." Brain and behavior. 13(1):e2838. doi:10.1002/brb3.2838.
- 115. △Begg DP (2015). "Insulin Transport into the Brain and Cerebrospinal Fluid." Vitamins & Hormon es. 98:229–248. doi:10.1016/bs.vh.2014.12.007.
- 116. △Gray SM, Barrett EJ (2018). "Insulin transport into the brain." American Journal of Physiology-C ell Physiology. 315(2):C125—C136. doi:10.1152/ajpcell.00240.2017.
- 117. <sup>a, b</sup>Huang Y, Lei L, Liu D, Jovin I, Russell R, Johnson RS, Di Lorenzo A, Giordano FJ (2012). "Normal glucose uptake in the brain and heart requires an endothelial cell-specific HIF-1α–dependent fun ction." Proceedings of the National Academy of Sciences. 109(43):17478–17483. doi:10.1073/pnas.1209281109.
- 118. <sup>△</sup>Srivastava S, Zahra FT, Gupta N, Tullar PE, Srivastava SK, Mikelis CM (2020). "Low Dose of Penf luridol Inhibits VEGF-Induced Angiogenesis." International journal of molecular sciences. 21(3):7 55. doi:10.3390/ijms21030755.
- 119. <sup>△</sup>Deng ZH, Zhong J, Jiang HL, Jeong HW, Chen JW, Shu YH, Tan M, Adams RH, Xie KP, Chen Q, Liu Y (2022). "Antipsychotic drugs induce vascular defects in hematopoietic organs." The FASEB Jour nal. 36(10):e22538. doi:10.1096/fj.202200862R.
- 120. <sup>△</sup>Kanmodi R, Bankole H, Oddiri R, Arowosegbe M, Alabi R, Rahmon S, Ahmodu O, AbdulRasheed B, Muritala R (2023). "Repurposing Antipsychotic Agents Against Targets of Angiogenesis Pathw ays for Cancer Therapy: An in-silico Approach." Current Drug Discovery Technologies. 20(6):38–46. doi:10.2174/1570163820666230606113158.
- 121. ≜Kato Y, Umetsu R, Abe J, Ueda N, Nakayama Y, Kinosada Y, Nakamura M (2015). "Hyperglycemic adverse events following antipsychotic drug administration in spontaneous adverse event report s." Journal of Pharmaceutical Health Care and Sciences. 1:15. doi:10.1186/s40780-015-0015-6.
- 122. <sup>△</sup>Chen J, Huang XF, Shao R, Chen C, Deng C (2017). "Molecular Mechanisms of Antipsychotic Dru g-Induced Diabetes." Frontiers in Neuroscience. 11:643. doi:10.3389/fnins.2017.00643.

- 123. △Antuna-Puente B, Disse E, Lavoie ME, Rabasa-Lhoret R, Laville M, Bastard JP (2009). "Evaluati on of the SIisOGTT index in a healthy population with normal glucose tolerance." Diabetes & Me tabolism. **35**(3):236–237. doi:10.1016/j.diabet.2009.02.004.
- 124. <sup>△</sup>D'Alessio DA, Kieffer TJ, Taborsky GJ Jr, Havel PJ (2001). "Activation of the Parasympathetic Nerv ous System Is Necessary for Normal Meal-Induced Insulin Secretion in Rhesus Macaques1." The J ournal of Clinical Endocrinology & Metabolism. 86(3):1253–1259. doi:10.1210/jcem.86.3.7367.
- 125. <sup>a, b</sup>Kiba T (2004). "Relationships Between the Autonomic Nervous System and the Pancreas Including Regulation of Regeneration and Apoptosis." Pancreas. **29**(2):e51–e58. doi:<u>10.1097/0000667</u>6-200408000-00019.
- 126. Anelles E, Butzler C, Jung D, Temme A, Gabriel HD, Dahl U, Traub O, Stumpel F, Jungermann K, Zie lasek J, Toyka KV, Dermietzel R, Willecke K (1996). Defective propagation of signals generated by sympathetic nerve stimulation in the liver of connexin32-deficient mice. Proceedings of the National Academy of Sciences. 93(18):9565–9570. doi:10.1073/pnas.93.18.9565.
- 127. <sup>△</sup>Nonogaki K (2000). "New insights into sympathetic regulation of glucose and fat metabolism." Diabetologia. **43**(5):533–549. doi:<u>10.1007/s001250051341</u>.
- 128.  $\triangle$ Gilon P, Henquin JC (2001). "Mechanisms and Physiological Significance of the Cholinergic Cont rol of Pancreatic  $\beta$ -Cell Function." Endocrine Reviews. 22(5):565–604. doi:10.1210/edrv.22.5.044  $\underline{0}$ .
- 129. <sup>△</sup>Miller RE, Waid TH, Joyce MP (1976). "Direct neural inhibition of insulin secretion in response to systemic hypoglycemia." American Journal of Physiology–Legacy Content. 230(4):1090–1094. do i:10.1152/ajplegacy.1976.230.4.1090.
- 130. △van Veen KEB, Brouwer MC, van der Ende A, van de Beek D (2016). "Bacterial meningitis in diab etes patients: a population-based prospective study." Scientific Reports. 6(1):36996. doi:10.1038/srep36996.
- 131. △Chawla S, Pund A, B V, Kulkarni S, Diwekar-Joshi M, Watve M (2018). "Inferring causal pathway s among three or more variables from steady-state correlations in a homeostatic system." PLOS ONE. 13(10):e0204755. doi:10.1371/journal.pone.0204755.
- 132. <sup>△</sup>Ono T, Steffens A, Sasaki K (1983). "Influence of peripheral and intracerebroventricular glucose a nd insulin infusions on peripheral and cerebrospinal fluid glucose and insulin levels." Physiology & Behavior. 30(2):301–306. doi:10.1016/0031-9384(83)90023-9.
- 133. ^Lipska KJ, Montori VM (2013). "Glucose Control in Older Adults With Diabetes Mellitus—More H arm Than Good?" JAMA Internal Medicine. 173(14):1306. doi:10.1001/jamainternmed.2013.6189.
- 134. <sup>△</sup>Meneilly GS, Tessier DM (2016). "Diabetes, Dementia and Hypoglycemia." Canadian Journal of Diabetes. **40**(1):73–76. doi:10.1016/j.jcjd.2015.09.006.
- 135. <sup>△</sup>Rhee SY (2017). "Hypoglycemia and Dementia." Endocrinology and Metabolism. **32**(2):195. doi:<u>1</u> 0.3803/EnM.2017.32.2.195.
- 136. ^Yaffe K (2013). "Association Between Hypoglycemia and Dementia in a Biracial Cohort of Older Adults With Diabetes Mellitus." JAMA Internal Medicine. 173(14):1300. doi:10.1001/jamainternmed.2013.6176.
- 137. △Brown JB, Nichols GA, Perry A (2004). "The Burden of Treatment Failure in Type 2 Diabetes." Di abetes Care. 27(7):1535–1540. doi:10.2337/diacare.277.1535.
- 138. <sup>△</sup>DeFronzo RA (2010). "Overview of Newer Agents: Where Treatment Is Going." The American Jo urnal of Medicine. **123**(3):S38–S48. doi:<u>10.1016/j.amjmed.2009.12.008</u>.
- 139. <sup>△</sup>Rosengren A, Jing X, Eliasson L, Renström E (2008). "Why Treatment Fails in Type 2 Diabetes." PLoS Medicine. 5(10):e215. doi:10.1371/journal.pmed.0050215.
- 140. ^García-Hermoso A, Cavero-Redondo I, Ramírez-Vélez R, Ruiz JR, Ortega FB, Lee DC, Martínez-Vi zcaíno V (2018). "Muscular Strength as a Predictor of All-Cause Mortality in an Apparently Healt hy Population: A Systematic Review and Meta-Analysis of Data From Approximately 2 Million M en and Women." Archives of Physical Medicine and Rehabilitation. 99(10):2100–2113.e5. doi:10.10 16/j.apmr.2018.01.008.
- 141. △Stensvold D, Viken H, Steinshamn SL, Dalen H, Støylen A, Loennechen JP, et al. (2020). "Effect of exercise training for five years on all cause mortality in older adults-the Generation 100 study: ra ndomised controlled trial." BMJ (Clinical research ed.). 371:m3485. doi:10.1136/bmj.m3485.
- 142. <sup>△</sup>Patil P, Lalwani P, Vidwans H, Kulkarni S, Bais D, Diwekar-Joshi M, Rasal M, Bhasme N, Naik M, Batwal S, Watve M (2021). "A multidimensional functional fitness score has a stronger associatio

- n with type 2 diabetes than obesity parameters in cross sectional data." PloS one. **16**(2):e024509 3. doi:<u>10.1371/journal.pone.0245093</u>.
- 143. Amomma H, Kawakami R, Honda T, Sawada SS (2022). "Muscle-strengthening activities are ass ociated with lower risk and mortality in major non-communicable diseases: a systematic review and meta-analysis of cohort studies." British Journal of Sports Medicine. **56**(13):755–763. doi:10.11 36/bjsports-2021-105061.
- 144. ^Ojha A, Vidwans H, Watve M (2023). "Does sugar control arrest complications in type 2 diabete s? Examining the rigor in statistical methods and causal inference in clinical trials." Qeios. IH7KE P.2.
- 145. △Rojas Vega S, Knicker A, Hollmann W, Bloch W, Strüder HK (2010). "Effect of resistance exercise o n serum levels of growth factors in humans." Hormone and metabolic research = Hormon- und S toffwechselforschung = Hormones et metabolisme. 42(13):982–986. doi:10.1055/s-0030-1267950.
- 146. △Jiang Q, Lou K, Hou L, Lu Y, Sun L, Tan SC, Low TY, Kord-Varkaneh H, Pang S (2020). "The effect of resistance training on serum insulin-like growth factor 1(IGF-1): A systematic review and meta –analysis." Complementary Therapies in Medicine. 50:102360. doi:10.1016/j.ctim.2020.102360.
- 147. ≜Rashid AL (2012). Effects of aggressive and non-aggressive exercises on insulin sensitivity in yo ung men; across sectional pilot study. MRes thesis, University of Glasgow.

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