

Review of: "*RUNX1* isoform disequilibrium in the development of trisomy 21 associated myeloid leukemia"

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This study by Gialesaki, Brauer-Hartmann, et al. addresses important topics in the molecular biology of the *RUNX1* gene and its encoded transcription factor and in the pathogenesis of Down syndrome-associated myeloid leukemias (ML-DS). The topic entails a unique situation in which *GATA1* mutations drive leukemogenesis, but only in the trisomy 21 (T21) genetic background. While the chromosome 21-linked *RUNX1* gene has been suspected (largely a priori) to be involved in this situation, direct positive supporting data have been slow to emerge. The current preprint describes convincing data supporting a key role for *RUNX1*, but interestingly not via its generic over-expression but rather through a shift in relative amounts of specific differentially spliced isoforms, i.e., the *RUNX1A*:*RUNX1C* ratio.

The evidence presented is of several types, both correlative and functional, and the data figures are quite clear and convincing. The first functional experiment was a CRISPR-Cas9 screen, which revealed *RUNX1* as being the top “hit” (among chromosome 21 genes) required for survival of a ML-DS cell line. The authors then examined *RUNX1* isoforms and discovered an increased A:C ratio in ML-DS cases, compared to normal hematopoietic stem/progenitor cells. So, they pursued this hypothesis further in experiments using lentiviral expression vectors to modulate the A:C ratio in HSPCs, and then in mouse fetal liver cells that they had engineered to contain *Gata1*s mutations. Readouts of these manipulations, including assays of cell proliferation and differentiation, plus transcriptomics, all converged on the conclusion that *RUNX1* “isoform dysequilibrium” is indeed sufficient to drive ML-DS. Moreover, biochemical analyses using pull down assays and mass spectrometry were also done - which revealed protein interactions - including relevant *RUNX1A*-specific ones such as an association with *MYC*:*MAX*, and differential interactions of specific *RUNX1* isoforms with *GATA1*. Lastly, the authors pursued the idea that *MYC*:*MAX* would be a potential therapeutic target specifically in ML-DS, with evidence for a therapeutic window of sensitivity, compared to normal HSPCs. For this reviewer, all of the data figures are convincing, thus supporting the authors' conclusion that - “through detailed functional validation, we discovered that, rather than *RUNX1* gene dosage, a disequilibrium of *RUNX1* isoforms and *RUNX1A* bias is key to T21-associated leukemogenesis.” After this important work, the main as yet unanswered question seems to be - how does T21 lead to the specific shift in *RUNX1* isoforms? The effects of T21 on alternative promoter epigenetic states, and alternative splicing of mRNAs, has become an active general research area (e.g., Wang, Y., et al., *Front. Cell Dev. Biol.*, 30 September 2021; Palmer et al. *PNAS* 2021 Vol. 118 No. 47 e2114326118; Muskens et al., *Nature Comm.* (2021) 12:821), so answers to this question should soon emerge.