

Review Article

Mechanisms of Glycolysis and Fermentation: A Non-Equilibrium Thermodynamics Perspective

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Every single chemical formula of modern models of glycolysis violates the scientific rules formulated by Newton (Physics) and Lavoisier (Chemistry). Yet, the formulae of the pioneers who investigated metabolism did not violate the laws of nature. Recently, the well-established models of metabolism have collapsed by re-introducing hydrogen (Chemistry, Physics) as the energy entity driving the Krebs cycle (Biochemistry). This review builds on a scientific concept of metabolism by introducing that glycolytically generated energy is either transformed into ATP or drives a biological process. The dynamic production and utilization of lactate (lactate flow non-equilibrium) is introduced as a central ATP-driven process and the first step of biosynthesis. A metabolism model based on non-equilibrium thermodynamics replaces the current understanding that one end product of glycolysis is consumed by mitochondria with two intermediates of the two-cell model of metabolism that are consumed by mitochondria. The pyruvate dehydrogenase complex, consuming pyruvic acid, saves one redox unit (2H) for storage as lipid or glycogen, whereas mitochondrial consumption of lactic acid enhances ATP recovery. An uncounted number of signalling pathways temporarily regulate the distribution of this single redox unit. Glycogenolysis massively impacts the flow non-equilibrium, an event permanently memorized by cells. The two-cell model of metabolism starts to functionally unite fields such as memory formation, obesity, exercise, schizophrenia, cancer, and inflammation by the common denominator: metabolism.

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Introduction (Physics)

Equilibrium thermodynamics, the 2nd law of thermodynamics, provides the understanding that an apple falls on Earth and Earth falls on an apple^[1]. Non-equilibrium thermodynamics, the 4th law of thermodynamics, has to provide an understanding of how the apple has reached flow non-equilibrium with Earth in order to fall down. Non-equilibrium thermodynamics is the Physics of Biology.

Maxwell's observation of the dissipation of energy in nature guided Thomson to the formulation of the thermodynamic demon hypothesis in 1879^[2]. The hypothesis implies that thermodynamic demons carry apples on trees to let them drop on physicists' heat. Physicists have hunted the demons for more than a century, in vain. Recently, the demon hypothesis was updated to the zombie-hypothesis: Maxwell's zombies mulling and mauling the 2nd law of thermodynamics^[3]. The undead are open systems; they eat and drink. Open systems do not fall under the jurisdiction of the 2nd law of thermodynamics. The undead metabolize and temporarily retain the gained energy by transferring the energy to a more stable energy particle. A living organism is an open system in steady state flow non-equilibrium with the environment and falls under the jurisdiction of the 4th law of thermodynamics.

Physicists have developed a comprehensive working plan to understand life. The aim of biochemistry is to provide physicists with the name of the energy particle (demon, zombie).

Today, in Biology, it is known that apple trees transform sunshine into the fuel glucose, and glucose is burned to water, carbon dioxide, and apple. The chemistry of the process burning

glucose to water, carbon dioxide, and $\sim -\Delta G$ 2800 kJ/mol is known. Apple trees do not emit the energy of $\sim -\Delta G$ 2800 kJ/mol. The biochemistry of the metabolic burning of glucose to water, carbon dioxide, and apple is unknown. The historical name of the thermodynamic demon is *Phlogiston*, which is emitted when substances are burned^[4]. The *Phlogiston* cult was replaced by the formulation of the law of the conservation of mass. Thus, if the demon has a molecular mass, then *Phlogiston* is hidden in the chemical formulae of glycolysis and fermentation. In open systems, *Phlogiston* turns into a zombie. In vivo, *Phlogiston* is generated during the metabolic burning or oxidation ($\text{NAD}^+ \rightarrow \text{NADH-H}^+$ -system) of glucose and moves to a defined coupled biological process. The coupled biologic process retains energy from emitting as heat by transforming energy to a temporarily more stable energy particle. The thermodynamic zombie vanishes from chemical formulae during transformation.

The original chemical formulae of the metabolic burning of glucose show carboxylic acids (R-COOH), even if it is known that acids promptly dissociate in water to anion (R-COO^-), proton (H^+), and heat^[5]. Now consider the kinetics of a proton-linked Monocarboxylate Transporter (proton-linked MCT): 1st, a proton (H^+) binds; 2nd, an anion (R-COO^-) binds; and the charge-neutral acid (R-COOH) moves through the membrane^[6]. The kinetics is in line with the 3rd law of thermodynamics, the law of motion. First energy, second reaction. Thus, H^+ is the energy particle freed during burning (NADH-H^+), the energy particle that freely dissociates (emits) in water, and the energy particle that initiates and guides unidirectional movement in biological processes.

Enzymes catalysing the burning of glucose are organized in complexes or metabolons^{[7][8][9]}. Enzyme complexes, such as the pyruvic acid dehydrogenase complex (PADHC), lactic acid-consuming Citric Acid Cycle complex, and the mitochondrial ATP-synthase complex, have in common that the intermediates are acidic (figure 1).

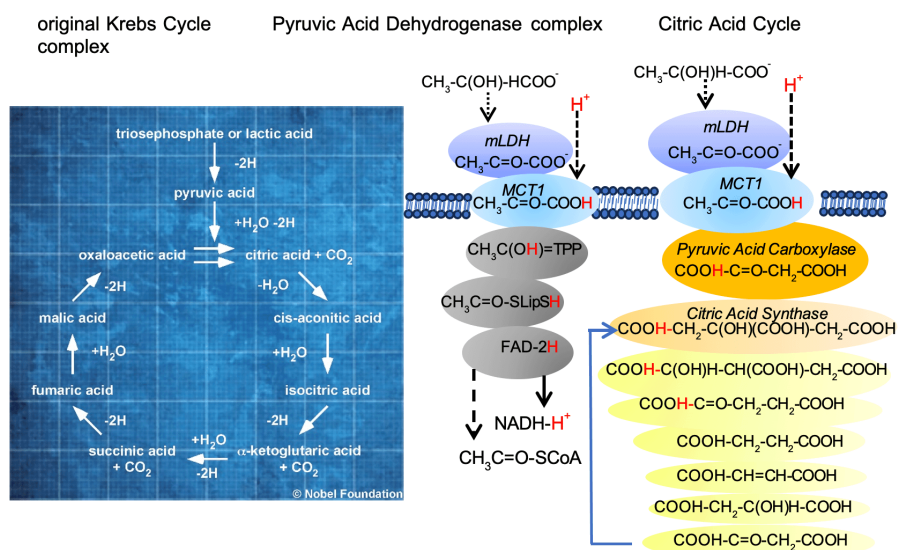


Figure 1. Mitochondrial consumption of lactic acid. The blueprint of the original concept of the Citric Acid Cycle encrypts the biochemical path of metabolic burning of lactic acid (Pyruvic Acid Dehydrogenase complex) and biosynthesis of di- and tri-carboxylic acids (Citric Acid Cycle complex)^[10]. Mitochondrial lactate dehydrogenase metabolically burns lactate ($\text{CH}_3\text{-C(OH)H-COO}^-$) to pyruvate ($\text{CH}_3\text{-C=O-COO}^-$)^[11]. Proton-linked Monocarboxylate Transporter 1 (MCT1) transfers pyruvic acid ($\text{CH}_3\text{-C=O-COOH}$) to the Pyruvic Acid Dehydrogenase complex or Citric Acid Cycle complex. The Pyruvic Acid Dehydrogenase complex metabolically burns pyruvic acid to acetyl-SCoA and NADH-H^+ ^{[12][13]}. Pyruvic Acid Carboxylase refills the Citric Acid Cycle complex with oxaloacetic acid ($\text{COOH-C=O-CH}_2\text{-COOH}$). Pyruvic Acid Carboxylase catalyses ATP-driven synthesis of oxaloacetic acid. Citric Acid Synthase catalyses the biosynthesis of citric acid^{[14][15][16][17]}.

This review introduces that the first intermediates of the glycolytic metabolon are acids. The Proton Transport Chain hypothesis states that intermediates are directly or water-free transferred within complexes to save dissociation heat for the coupled biological process^[17]. First, the proton is transferred to the coupled enzymatic process; 2nd, the anion follows. In 1976, the flow of energy (particle H^+) through mitochondrial ATP synthase was named proticity^[18]. The biochemistry of enzyme complexes indicates that enzyme complexes are bio-wires (figure 2).

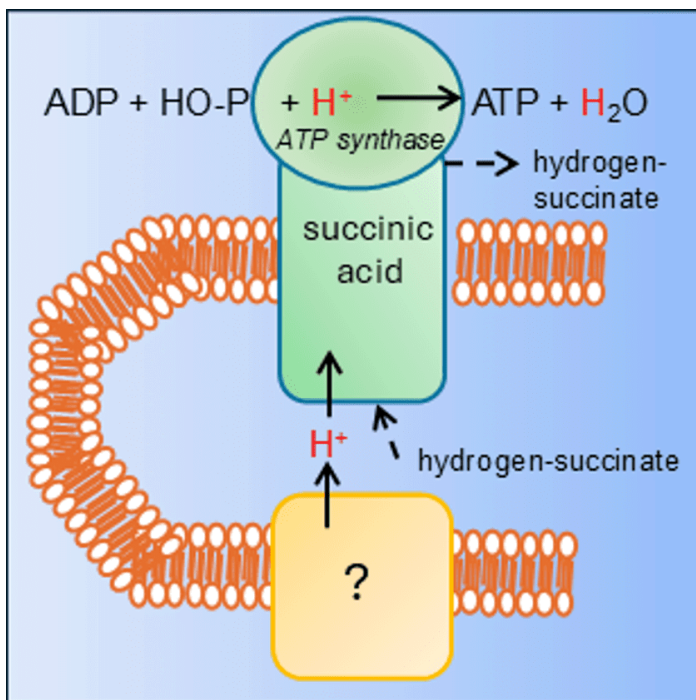


Figure 2. Mitochondrial ATP synthase. Proton (H^+) and hydrogen-succinate form charge-neutral succinic acid. Succinic acid is wired through the ATP-synthase complex. The complex catalyses the transfer of energy (particle H^+) from unstable succinic acid to the temporarily more stable tri-phosphoric acid anhydride group of ATP. The proton (H^+) is discharged to water^{[19][20]}.

Enzymes stabilize their substrate water-free within the active site. The substrate is solved by the enzyme complex, not by water. The physical quantity [mol/L] is applied to calculate an enzymatically catalysed reaction of a thermodynamic equilibrium. Direct or water-free transfer of an intermediate within an enzyme complex is mathematically one molecule in zero Litres of water. 1 molecule of acid divided by 0 L of water is an infinite molarity. An infinite molarity of one product changes the understanding (Physics and Mathematics) of a reversible chemical process into the understanding of the biochemical process of an irreversible metabolic flow.

One enzyme catalyses 1 molecule at a time. Molecule/time is the physical quantity of a flow. Acid/time is the physical quantity of the flow of energy (particle H^+) and material (anion) through enzyme complexes.

In ecology, the tentative 4th law of thermodynamics states: a flow of energy and material is sufficient to form ordered structures^[21]. In this case, we suggest calling the thermodynamic zombie, proton (H^+).

The biochemistry of the proton-linked MCT₁-Carbonic Anhydrase II complex, located at the cell membrane, is best visualized as an 'ordered structure' generating a flow non-equilibrium. Carbonic Anhydrase II activity is located outside the cell (Figure 3). At time point 0, the environment and the cell are in lactate equilibrium. Carbonic Anhydrases contain zinc (Zn^{2+}),

which is a Lewis acid in their active sites. Zn^{2+} reacts with environmental water to initiate the Proton Transport Chain: $\text{Zn}^{2+} + \text{H}_2\text{O} \rightarrow \text{Zn}^{2+}[\text{OH}^-] + \text{H}^+$ [22].

Thus, firstly, the energy particle H^+ taken from water is water-free transferred to the proton-linked MCT_1 ; secondly, lactate binds, and lactic acid moves into the cell.

The cell continuously emits carbon dioxide (the acid anhydride of carbonic acid). This emission restores the activity of Carbonic Anhydrase II, following the reaction: $\text{Zn}^{2+}[\text{OH}^-] + \text{CO}_2 \rightarrow \text{Zn}^{2+} + \text{HCO}_3^-$. The flow of CO_2 permanently restores Carbonic Anhydrase II activity. This process generates a proticity that pumps lactic acid from the environment into the cell, causing the cell to enter a state of lactate flow non-equilibrium with its surroundings.

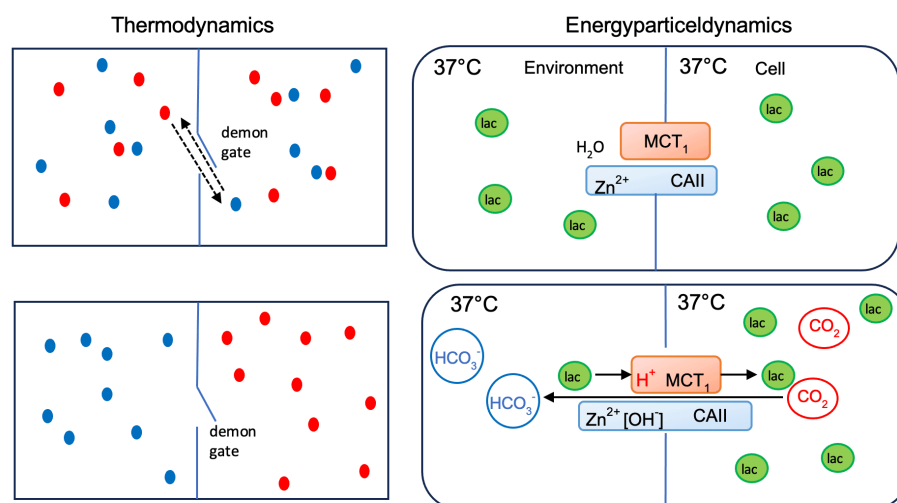


Figure 3. Thermodynamic demons and zombies. (left) A thermodynamic demon guards the demon gate and sorts blue and red circles. (right) Cells and environment are in lactate (lac) equilibrium. Carbonic anhydrase II (CAII) carries the acid zinc²⁺[22]. Zn^{2+} reacts with water. Freed H^+ is transferred to proton-linked MCT_1 (MCT_1). Emitting CO_2 (red) recovers the activity of the CAII-proton-linked MCT_1 complex. A lactate flow non-equilibrium is generated. The end product of the reaction is bicarbonate (HCO_3^- , blue).

The path the authors have taken to this review started with the formulation of the Proton Transport Chain hypothesis and is documented in five reviews^{[24][21][25][26][17]}. This review discusses the chemical formulae of glycolysis and fermentation published by Meyerhof^[5]. Meyerhof's review is the most recently published concept of glycolysis and fermentation not violating the law of the conservation of mass that we have found.

The understanding of Chemistry and the 2nd law of thermodynamics guided scientists to define lactic acid as the exported end product of fermentation and lactate as the end product of glycolysis^{[27][28]}. This review updates the chemistry of glycolysis and fermentation to the biochemistry of glycolysis and fermentation. In line with the 4th law of thermodynamics, the path of energy (particle H^+) within the glycolytic metabolon was followed.

We assume this review presents the first scientific (in line with the laws of nature and published pioneer work) integration of the $\text{NAD}^+/\text{NADH}+\text{H}^+$ system and ATP in glycolysis and fermentation^[2]. Understanding the tri-phosphoric acid anhydride group of ATP as the storage of the energy of a proton allowed us to introduce lactate as the first ATP-driven biosynthesis product. The glycolytic flow of energy (particle H^+) and material generates the cytosolic lactate flow non-equilibrium. The generation of this cytosolic lactate concentration gradient enables the mitochondrial lactate dehydrogenase-proton-linked MCT_1 complex (figure 1) to catalyse the

metabolic burning of lactate to pyruvate and NADH-H^+ and the membrane transfer of charge-free pyruvic acid to the membrane-associated PADHc and Citric Acid Cycle complex.

Finally, the first part of the apple is on the tree. Energy (particle H^+) freed during the hexokinase-catalysed hydrolysis of ATP ($\text{ATP} + \text{H}_2\text{O} \rightarrow \text{ADP-H}^+ + \text{HO-P}$) does not chaotically emit, heat up, and acidify the cell but is wired to Pyruvate Hydrogenase (PH) to generate the cytosolic lactate flow non-equilibrium. Lactate, understood as the first biosynthesis product, falls into mitochondrial degradation to be metabolically burned to pyruvic acid and NADH-H^+ . Thus, 85 years after the formulation of the lactic acid-consuming Citric Acid Cycle, Krebs' work is rationally linked with glycolysis^{[14][15][29]}. A century after the discovery of the $\text{NAD}^+/\text{NADH-H}^+$ system and ATP, the co-enzymes are finally scientifically (in line with the laws of nature) integrated into glycolysis and fermentation.

Introduction II (Biochemistry)

F. Cori: "Some 50 years ago - in 1929, to be exact - we proposed a cycle of the glucose molecule which could be in turn liver glycogen, blood glucose, muscle glycogen. The conversion of lactic acid to liver glycogen would complete the cycle"^{[30][31]}. The Cori cycle was formulated in the year K. Lohmann discovered ATP^[32]. At this time, the citric acid cycles and thereby mitochondrial participation in biosynthesis and ATP recovery were undiscovered. F. and G. Cori determined that insulin accelerates the Cori cycle from liver to muscle glycogen, while epinephrine accelerates the cycle in the opposite direction^[30]. Today, it is known that other organs such as blood (thrombocytes, macrophages, T-cells) or brain (astrocytes) also contain glycogen. The Cori cycle developed into the two-cell model of metabolism^[25].

Cells memorize the event of glycogenolysis^{[33][34][35]}. Astrocytic glycogenolysis, or more specifically the astrocyte neuron lactate shuttle hypothesis (ANLS), was introduced as energy on demand. Briefly, memory formation is triggered by a boost of fuels directed from astrocytes to neurons. More recently, the ANLS hypothesis was interpreted as astrocyte neuron communication (ANC)^[24]. ANC shifted the biological function of glycogenolysis from fuel to a signalling pathway and added pyruvic acid as a metabolic messenger to the two-cell model of metabolism. Glycogen was introduced as storage of the metabolic signalling molecules glucose, lactic acid, and pyruvic acid. Transmitters, such as insulin and epinephrine, temporarily adapt the steady-state flow non-equilibrium between two cells according to environmental changes^{[30][31]}. The steady-state lactate/pyruvate flow ratio in the liver is approximately 7/1, whereas the steady-state lactate/pyruvate flow ratio in resting muscle is approximately 12/1 and 159/1 in working muscles^{[36][37]}. Transmitters triggering glycogenolysis massively impact the cytosolic lactate/pyruvate flow ratio. The temporary imbalance is memorized by changes in expression levels of enzyme isoforms and enzyme subtypes in a way leading to a permanent acceleration of the flow of metabolic messengers^[38].

The two-cell model of metabolism also comprises cell adhesion proteins physically stabilizing cell-cell communication^[25]. The two-cell model of metabolism has given D.O. Hebb's 'reverberatory activity' the molecular background of glucose metabolism^[39]. Our approach has opened avenues by linking the scientific fields of memory formation and schizophrenia via a functional concept of metabolism. Genetic and environmental factors statistically associated with schizophrenia are candidates to be sorted into the two-cell model of metabolism. Predisposition to schizophrenia was functionally linked to instability of the physical interaction between astrocytes and neurons. Pronounced leakage of fuels into the interstitial fluid during stress-triggered astrocytic glycogenolysis was linked to acute schizophrenia^[40]. Leaked glucose is a signalling molecule of inflammation, an activator of microglia cells, and triggers the release of inflammatory messengers^[25]. Thus, glucose metabolism is the common denominator of all biological processes.

Unfortunately, all of the above cannot be rationally followed on a molecular level. As already stated at the beginning of the review, the synthesis, export, import, and mitochondrial consumption of lactic acid have been under investigation for more than a century. Today, however, the well-established chemical formulae of glucose metabolism violate the law of the conservation of mass, as well as published pioneer work. The chemical formulae no longer provide the experimentally determined glycolytic intermediate proton (H^+). The proton (H^+) is obligatory to export lactic acid (lactate-H^+) or to be stored as ATP ($\text{ADP} + \text{P-OH} + \text{H}^+ \rightarrow \text{ATP} + \text{H}_2\text{O}$).

Whereas the synthesis, export, and import of lactic acid have been under investigation for more than a century^{[41][42]}, the molecular mechanisms behind “energy for storage as glycogen” and “energy on demand” lie far outside common understanding. *In vivo*, a single cell simultaneously synthesizes lactate from pyruvate and metabolically burns lactate to pyruvate. Mitochondrial consumption of lactate entails that the mitochondrial LDH-proton-linked MCT₁ complex unidirectionally catalyses the reaction aiming to equalize a cytosolic lactate flow non-equilibrium^[43]. Whereas Pyruvate Hydrogenase (LDH isoform) anchored with the glycolytic metabolon must unidirectionally generate a cytosolic lactate flow non-equilibrium (biosynthesis) to enable mitochondrial consumption of lactate.

Two LDH isoforms simultaneously acting in opposite directions are introduced in the context of memory formation and formulated in the ANLS hypothesis. Nevertheless, the understanding behind the molecular mechanisms of memory formation/in vivo metabolism lies far outside common understanding because it is well-established that the isoforms of LDH catalyse the identical thermodynamic equilibrium and that formation of a memory is a violation of the 2nd law of thermodynamics^{[44][45]}. Therefore, it is well-established that we do not understand biosynthesis or the generation of a cytosolic lactate flow non-equilibrium on the basis of equilibrium thermodynamics (enzyme kinetics)^[45]. Nevertheless, *in vivo*, lactate is simultaneously synthesized, exported, imported, and degraded^{[42][28]}. Biology is obsessed with thermodynamic demons. These demons are known to have the ability to stop, strike, push, or pull any lactate molecule at will, thereby altering its natural course of motion^[2].

This review continues to challenge an understanding based on equilibrium thermodynamics and enzyme kinetics by developing a flow non-equilibrium thermodynamics-based concept of metabolism. “Equilibrium is an inert state of death; here the flow of matter and energy through a biological system stops, and the system reaches a lifeless state of thermodynamic equilibrium”^[46]. In other words, equilibrium thermodynamics and enzyme kinetics are the path to understanding lifeless material. The mathematics of equilibration thermodynamics is well-established from basic enzyme kinetics; isolated enzymes in homogeneous and closed systems catalysing the identical lifeless state of a thermodynamic equilibrium. Isolated enzymes are dead material acting in line with the 2nd law of thermodynamics.

Our approach to beginning to understand the dissipation of the energy (particle H⁺) in nature^[2] is to open the reference of glycolysis/fermentation, published by O.F. Meyerhof in 1951^[5].

Methods

Meyerhof’s Scheme 1 is the peak of knowledge and understanding of the chemistry of glycolysis and fermentation and the scientific reference for glycolysis and fermentation^[5]. Meyerhof understood that the 2nd law of thermodynamics excludes the storage of energy (particle H⁺) as ATP; he also understood that the law of the conservation of mass excludes that 1 H⁺ is both used to generate ATP ((ADP + P-OH)⁻ + H⁺ → ATP + H₂O) and exported as lactic acid (lactate-H⁺). Meyerhof’s understanding of science prevented him from integrating ATP (discovered 1929) and NAD⁺ (discovered 1906) in Scheme 1^{[47][32][19]}. Neither ATP nor the NAD⁺/NADH-H⁺ system can be scientifically integrated into a concept based on equilibrium thermodynamics and randomly distributed enzymes. Consequently, all models of glycolysis showing ATP are alchemistic inventions not supported by the law of the conservation of mass, the 2nd law of thermodynamics, and published pioneer work on glycolysis and fermentation.

Set theory: the one and only common intersection of the chemical formulae presented by Meyerhof and actual glycolysis inventions is glucose. Subduction of the atoms of glycolytic intermediates from the scientific reference indicates the atoms and molecules not currently accounted for or recognized in the current models of glycolysis and fermentation. In modern models of glycolysis, lactate and all protons (H⁺) have been deleted from chemical formulae. Lactate and proton are experimentally determined glycolytic intermediates.

Changes to the original concept, such as replacing phosphoric acid^[5] with ATP, entail that the glycolytically generated H⁺ to be stored as ATP must be removed from the chemical formula of the metabolic pathway. Meyerhof understood that the integration of ATP necessitates changing the end product from lactic acid to lactate. Stoichiometry dictates that 1 H⁺ is either exported as lactic

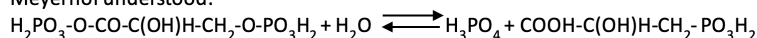
acid or stored as ATP. Therefore, pioneers were aware that the chemical mechanisms of fermentation are nearly completely understood and just the fate of some protons remained unexplainable^[48].

Notably, pioneers knew that acids rapidly dissociate in water, but Meyerhof's Scheme 1 shows a Proton Transport Chain starting with undissociated 3-(dihydrogen)-phosphoglyceric acid and ending with the export of lactic acid^[5]. The law of the conservation of mass dictates that the hydrolysis of the acid anhydride 1,3-diphosphoglyceric acid provides the acid 3-(dihydrogen)-phosphoglyceric acid. Pioneers experimentally demonstrated that the glycolytic metabolon protects the acidic intermediates from dissociation. Kennedy and Lehninger:

"Fluoride was added to inhibit enolase and the endpoint measured manometrically indicated the formation of 3-phosphoglyceric acid, which causes CO₂ liberation from a bicarbonate buffer."^[49].

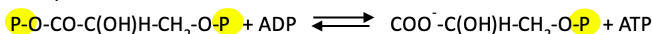
Let us consider the state of knowledge and understanding in 1951^[5] and the common state of knowledge and understanding^[50].

Meyerhof understood:



1,3-Diphosphoglyceric acid and Water are in equilibrium with Phosphoric acid and 3-Phosphoglyceric acid.

Today:



1,3-bisPhosphoglycerate and ADP are in equilibrium with 3-Phosphoglycerate and ATP.

Our approach glycolysis:



1-(monohydrogen), 3-(dihydrogen)-Diphosphoglyceric acid and ADP are unidirectionally metabolized to ATP and 3-(monohydrogen)-Phosphoglyceric acid

Our approach biosynthesis:



1,3-(monohydrogen)-Diphosphoglyceric and ADP are unidirectionally synthesised from ATP and 3-(monohydrogen)-Phosphoglycerate.

Depending on the individual point of view, the modern formula is either illustrated as an equilibrium or a unidirectional process. The phospho-groups are more commonly illustrated as two-times negatively charged (all protons are deleted) than as yellow dots. A common feature of modern models is the change in nomenclature of 1,3-Diphosphoglyceric acid to 1,3-bisPhosphoglycerate.

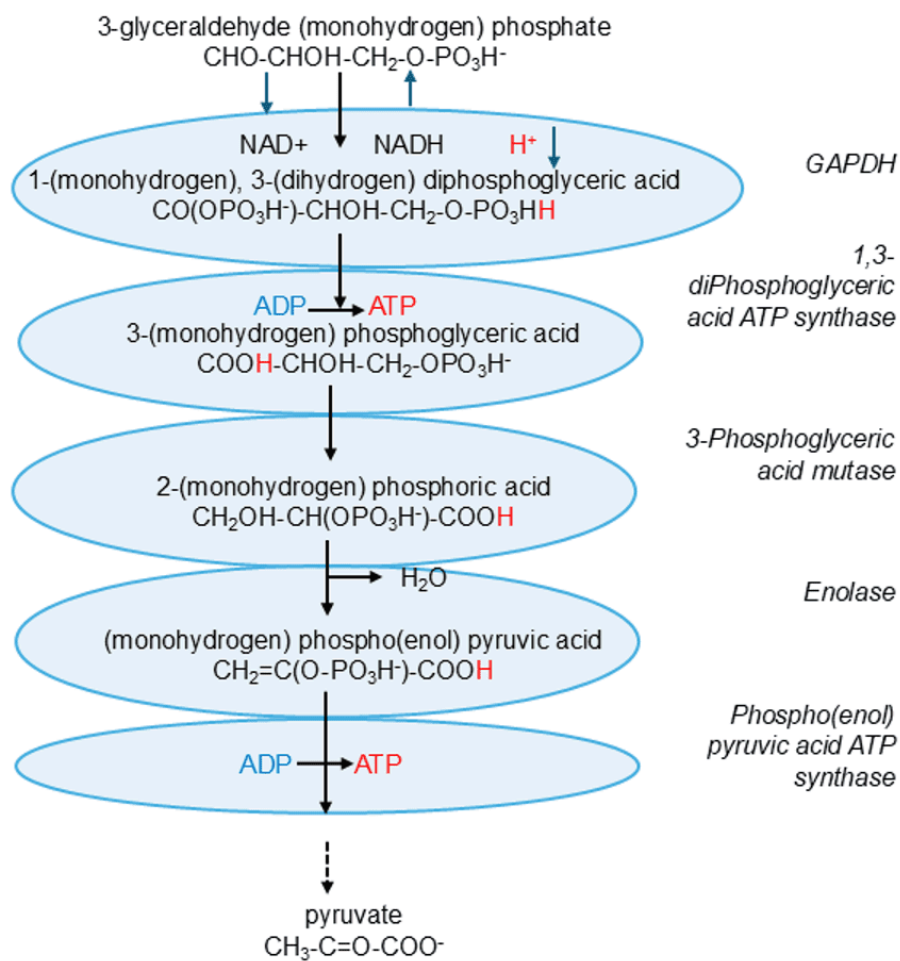
Our approach physically separates the pathways of glycolysis and biosynthesis and chemically differentiates between the acidic glycolytic intermediates and the pH-neutral cytosolic pool of biosynthesis intermediates.

The aims of this review are to (i) determine the glycolytic reactions generating energy (particle H⁺), (ii) trace the proticity driving the reduction of pyruvate to generate a lactate flow non-equilibrium, (iii) trace the flow of energy (particle H⁺) initiating monocarboxylic acid export (fermentation), and (iv) trace the flow of energy and material storing the energy (particle H⁺) as a temporarily stable glycolytic steady state ATP flow non-equilibrium (glycolysis).

The amalgamation of the law of the conservation of mass and the 4th law of thermodynamics constitutes the principles guiding the scientific domain of biochemistry. The authors' interpretation and nature's application of natural laws may vary. Hence, we recommend seizing the opportunity to align the laws of nature with the individual well-established model of metabolism. Do all chemical formulas of well-established glycolysis genuinely fail to adhere to stoichiometry? Does glycolysis generate 2 protons twice, aiming to store the energy as a steady state ATP flow non-equilibrium?

Integration of ATP generation into the original Embden-Meyerhof-Parnas pathway

Glycolysis produces 2 H^+ twice for storage as two sets of 2 ATP. Meyerhof depicted 3-glyceraldehyde-(dihydrogen)-phosphate as the substrate of oxidative phosphorylation. We suggest cytosolic 3-glyceraldehyde-(monohydrogen)-phosphate as the primary substrate of the oxidative branch of the glycolytic metabolon and to replace $-2H$ by integrating $NADH-H^+$ into glycolysis (pathway 1). Glyceraldehyde-3-phosphate dehydrogenase (GAPDH) catalyses the oxidative phosphorylation (metabolic burning) of 3-glyceraldehyde-(monohydrogen)-phosphate to 1-(monohydrogen), 3-(dihydrogen)-diphosphoglyceric acid. The glycolytically formed energy (particle H^+) is suggested to be attached at the 3-(monohydrogen)-phosphate group. 1-(monohydrogen), 3-(dihydrogen)-diphosphoglyceric acid moves to the coupled enzymatic process. 1,3-Diphosphoglyceric acid ATP synthase (PGK isoform) catalyses energy transfer from H^+ (dihydrogen-phosphate group) to the more stable adenosine *tri*-phosphoric acid anhydride (ATP) and the hydrolysis of 1,3-diphosphoglyceric acid to 3-(monohydrogen)-phosphoglyceric acid. The second energy particle is hidden in the acid anhydride group of 1,3-diphosphoglyceric acid.



Pathway 1. Oxidative branch of the glycolytic flow. Meyerhof's Scheme 1 presents the chemistry of glycolysis and fermentation^[2]. We divided the glycolytic metabolon into oxidative and reductive branches. Here, all reversible reactions have been changed into a unidirectional flow. The proton (H^+) of the acidic intermediates protected from dissociation by the metabolon (blue) is illustrated in red. Phosphoric acid is represented by ATP, and the $\pm 2\text{H}$ transfer is depicted as the $\text{NAD}^+/\text{NADH-H}^+$ system. The metabolic burning of 3-glyceraldehyde (monohydrogen)-phosphate generates two H^+ to be stored as two ATP. The first H^+ (NADH-H^+) attaches at the (monohydrogen)-phosphate group to form the acidic (dihydrogen)-phosphate group and initiate the Proton Transport Chain. The next catalytic reaction stores one glycolytically generated H^+ as ATP and hydrolyses the acid anhydride group of 1,3-diphosphoglyceric acid. Hydrolysis frees the second H^+ as 3-phosphoglyceric acid. Phospho(enol)pyruvic acid ATP synthase transfers the energy of the carboxylic acid to ATP, and pyruvate is given into the cytosol of carboxylates.

The glycolytic flow of energy and material is unidirectional. Therefore, all reaction arrows Meyerhof indicated as reversible processes (Chemistry) are changed into a unidirectional or irreversible flow (Biochemistry). Flow-generated energy is stored as cytosolic ATP flow non-equilibrium. *In vivo*, energy does not chaotically emit; energy flow has formed the glycolytic metabolon. The glycolytic flow of energy (particle H^+) ends with the storage of energy as temporarily more stable cytosolic ATP flow non-equilibrium catalysed by Phospho(enol)pyruvic acid ATP synthase (or a specific isoform of pyruvate kinase)^[51]. Pyruvate, the end product of the oxidative branch, is given into the cytosolic pool of carboxylates.

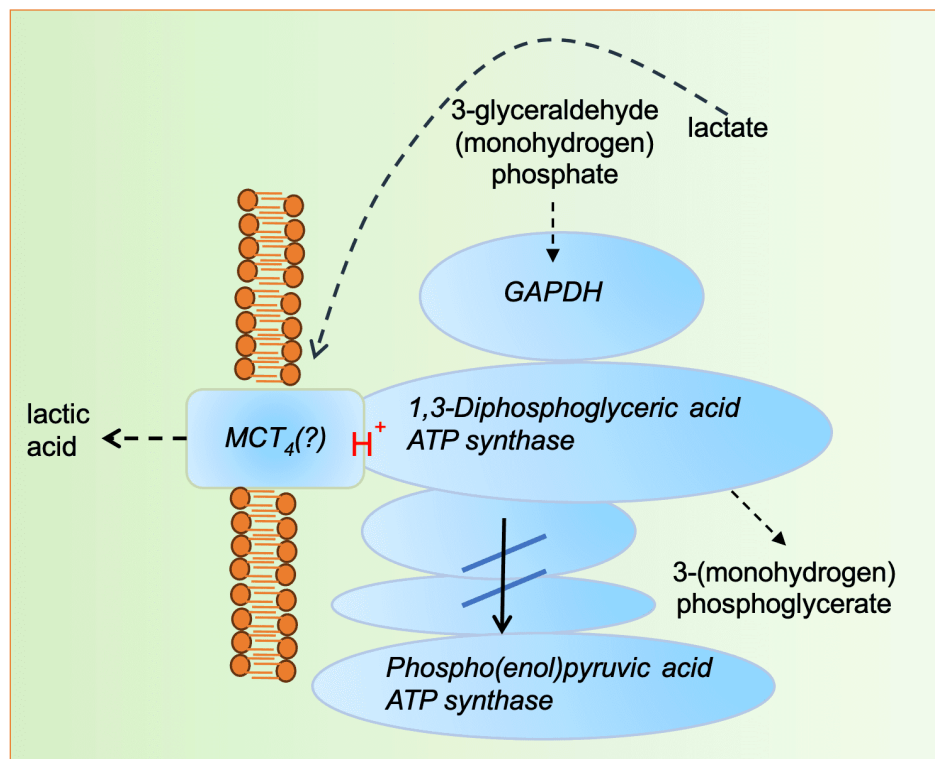
The integration of ATP and NADH-H^+ into the chemistry of glycolysis and fermentation challenges stoichiometry by opening two questions: firstly, fermentation: how can lactic acid (lactate-H^+) be exported when the proton of lactic acid already vanished from the chemical formula during

discharging to water and storage of the energy as ATP? Secondly, regarding the recovery of the co-enzyme NADH or the cytosolic lactate flow concentration gradient; what is the origin of the obligatory proton (H^+) driving the biosynthesis of lactate when glycolytically generated energy ($NADH-H^+$) is stored as cytosolic ATP flow non-equilibrium?

The fermentation pathway

Experimental work suggests that the association of GAPDH and 1,3-Diphosphoglyceric acid ATP synthase (isoform of PGK) at the plasma membrane is a regulated process^[52]. An acid, such as 3-(monohydrogen)-phosphoglyceric acid, released in the immediate proximity of proton-linked MCT, initiates the export of monocarboxylic acid^[6]. Close proximity can be interpreted as creating an acidic micro-environment, encouraging the transporter to equalize the generated membrane pH gradient. In this scenario, 3-(monohydrogen)-phosphoglyceric acid reacts with the bicarbonate-buffered micro-environment of the glycolytic metabolon, and hydration energy is emitted as heat and carbon dioxide. Nevertheless, an acidic micro-environment is sufficient to explain the characterization of proton-linked MCT_4 as an exporting carboxylic acid transporter^[53] ^[54].

Evolution surely optimizes the efficiency of proton (H^+) flow by transitioning from the creation of an acidic flow micro-environment (pH) to the direct transfer of energy (particle H^+) from unstable 3-(monohydrogen)-phosphoglyceric acid to proton-linked MCT. It is rational to postulate that the glycolytic metabolon releases 3-(monohydrogen)-phosphoglyceric acid if the flow of energy and material is blocked^[49]. The flow can be blocked by low cytosolic ADP. Thus, fermentation is introduced as regulated incomplete glycolysis (Pathway 2).



Pathway 2. Fermentation. The oxidative branch of the glycolytic metabolon (blue) is attached at the plasma membrane. Quenched glycolytic flow triggers the release of 3-phosphoglyceric acid^[49]. 3-phosphoglyceric acid transfers the proton (H^+) to proton-linked MCT, initiates the binding of lactate, and the export of lactic acid. The end product 3-phosphoglycerate is given into the cytosolic pool of carboxylates.

Proton-linked MCT-glycolytic metabolon unidirectionally catalyses:

3-(monohydrogen)-phosphoglyceric acid (glycolytic metabolon) + lactate (cytosolic pool) → lactic acid (environment) + 3-(monohydrogen)-phosphoglycerate (cytosol)

Altering the historical perspective from the gradual degradation of the carbon backbone to the flow of the energy particle can be a somewhat perplexing concept. Notably, the exported monocarboxylate is not synthesized during fermentation but rather sourced from the cytosolic pool. The ATP balance of fermentation is +/- 0. These findings prompt the question: What purpose does fermentation serve?

Fermentation is the biosynthesis of 3-(monohydrogen)-phosphoglycerate (3-PG). For the price of one monocarboxylate taken from the cytosolic pool and one H⁺ not given into storage as ATP, a cytosolic 3-PG non-equilibrium is generated. Similar to the recently reintroduced term 'incomplete burning', fermentation can be understood as incomplete glycolysis^[55].

Chemistry guided the understanding of synthesis as the reverse reaction of degradation. Non-equilibrium thermodynamics allows us to understand synthesis as blocked degradation. The mechanism of incomplete metabolic burning was introduced in the context of the regulation of the Citric Acid Cycle complexes: an imbalance of the fuel/oxygen ratio in favour of the fuel quenches the recovery of the co-enzymes. Quenched recovery of the activity of mitochondrial malic acid dehydrogenase (NAD⁺) blocks metabolic burning, and malic acid is NAD⁺-regulated pushed into the pool of carboxylates. Generation of a cytosolic (monohydrogen)-malate flow non-equilibrium provides the substrate of biosynthesis pathways consuming (monohydrogen)-malate.

Fermentation (generating a 3-PG flow non-equilibrium) is the provision of the substrate of serine biosynthesis. Proliferation of some cancer cells depends on serine biosynthesis; 3-phosphoglycerate dehydrogenase and proton-linked MCT₄ are targets for anti-cancer treatment. 3-PG levels seem to be directly involved in p53 activation to control cell fate^{[56][57][58][59]}. Thus, we suggest that obligate aerobic cells can regulate the switch to aerobic fermentation at a high ATP/ADP ratio to open biosynthesis pathways consuming 3-PG.

We generalized the observation that enzyme complexes protect acidic intermediates from dissociation and thereby integrated original pioneers' work into the concept of glycolysis ^{[49][60]}. Applying stoichiometry replaces the current understanding considering fermentation as anaerobic recovery of two ATP and questions the current understanding of mitochondrial recovery of ATP^[61]. In 1976, the flow of energy (particle H⁺) through enzyme complexes was named proticity^[19]. Walker and coworkers determined that 3-5 protons are necessary to mechanically drive the recovery of 1 ATP^[62]. Current knowledge sets that 32 ATP are recovered during the metabolic burning of two pyruvate, suggesting that the metabolic burning of two pyruvate generates ~ 128 H⁺ (32 x 4)^{[60][62]}.

The biochemistry of the glycolytic metabolon questions the stoichiometry of mitochondrial ATP recovery by introducing that one H⁺ equals the recovery of one ATP. Water-free or direct transfer of nascent proton skips the exothermic reaction of the acid with water; thereby, the pH gradient turns into a non-factor. Nath further developed Mitchell's and Walker's theory by firstly postulating and later experimentally demonstrating that charge-free succinic acid is wired through mitochondrial ATP synthase^{[63][64]}.

The cytosolic lactate flow non-equilibrium

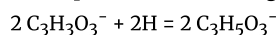
Glycolysis generates 2 H⁺ twice to be stored as two sets of 2 ATP. The net chemical formula of the glycolytic flow of protons (H⁺) leads to pyruvate:

Glucose → 4 ATP + 2 pyruvate⁻ + 2 NADH

$C_6H_{12}O_6 = 4 H^+ + 2 C_3H_3O_3^- + 2 H^-$

The glycolytic flow has the physical quantity (acid/time). An enzyme complex catalyses one metabolite at a time, not two metabolites as the net chemical formula suggests. Therefore, a flow is accomplished when the reduced co-enzyme (NADH) is recovered twice to NAD⁺ and the process can start again. The GAPDH-PH complex arrests the co-enzyme (NAD⁺/NADH) and thereby determines that the redox unit (2H) is transferred to pyruvate^[65]. The biochemical formula of Pyruvate Hydrogenase (PH)-catalysed recovery of NAD⁺ is:

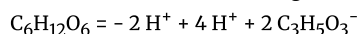
Pyruvate (cytosolic pool) + NADH (glycolytic metabolon) + H⁺ (proticity) → (cytosolic) lactate (flow non-equilibrium) + NAD⁺ (glycolytic metabolon)



Pyruvate is the end product of the oxidative branch of glycolysis, an intermediate of glycolysis, and a substrate of the reductive branch (biosynthesis). The hydrolysis of the acid anhydride ATP is a well-established source of energy for driving biosynthesis.

Providing an acid to the GAPDH-PH complex provides a mathematically infinite concentration of H⁺, driving a lactate flow non-equilibrium. Lactate stores energy by discharging the energy entity and the covalent binding of H. Therefore, the net chemical formula of glycolytic flow ends in the first ATP-driven biosynthesis product:

Glucose → - 2 ATP (consuming) + 4 ATP (non-equilibrium) + 2 lactate (non-equilibrium)



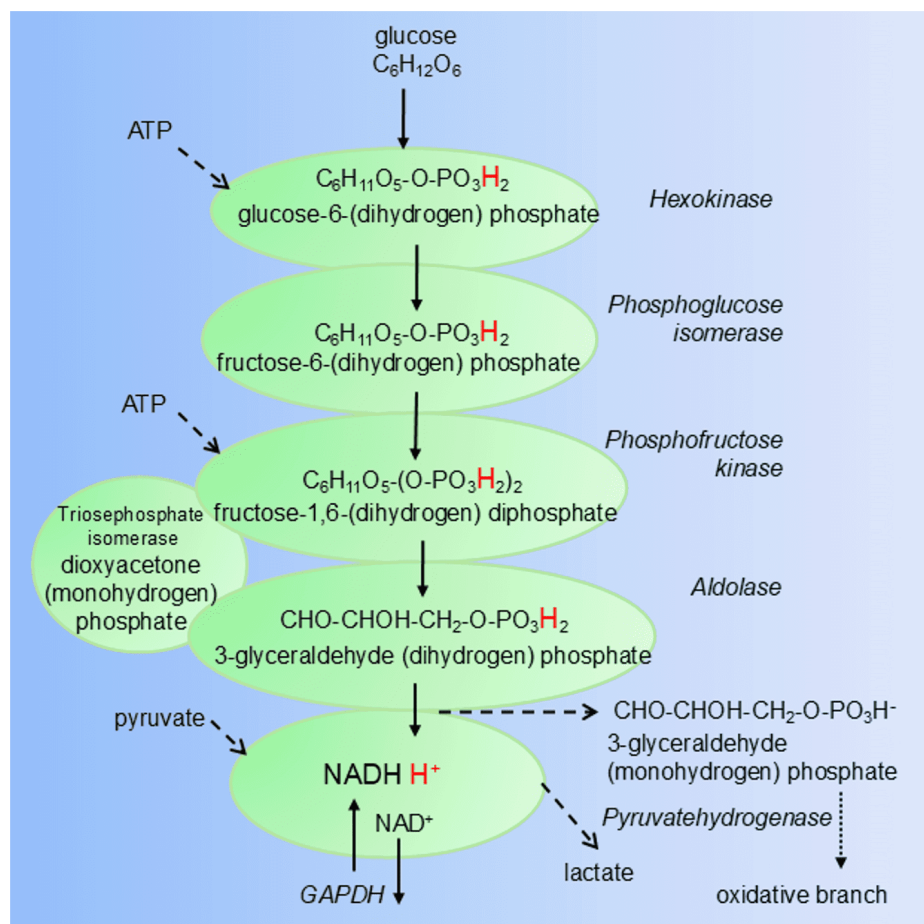
The original EMP pathway was presented in line with the law of the conservation of mass and equilibrium thermodynamics. One molecule of glucose has to be understood to be in equilibrium with two molecules of lactic acid (2 H⁺ + 2 lactate)^[5]. Chemistry is based on a homogeneous pool of all components. In a pool, -2 H⁺ and + 4 H⁺ represent identical mathematical parameters to be equalized by the reactions (basic enzyme kinetics) to +2 H⁺. To navigate the review more effectively, the established notion of a pool (closed system) needs to be broadened to include an understanding of a flow (open system). A flow of protons operates along a timeline. E. Rutherford might elucidate the physics of a flow in this manner: "No man ever drinks the same draft (beer) twice, for it is not the same beer and not the same man." Similarly, successful bartenders comprehend that two empty glasses and four beers differ from two beers.

By incorporating the cytosolic lactate flow non-equilibrium as the initial ATP-driven synthesis product, we are on the verge of taking the first step towards bringing Newton's apple back up to the tree. What is lacking, however, is the beginning of the knot: the reductive branch of the glycolytic metabolon.

The first glycolytic metabolon

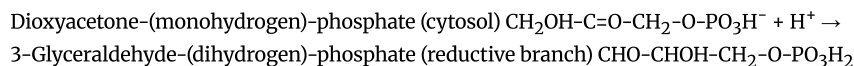
Although Kurganov and co-workers did not consider hexokinase (HK) as part of the first glycolytic multienzyme complex^[8], we have integrated HK into the first glycolytic Proton Transport Chain as follows (Pathway 3):

HK catalysed the phosphorylation of glucose and freed 1 proton (H⁺). This proton can either associate with glucose-6-(monohydrogen)-phosphate or chaotically emit into the environment. Attaching energy (particle H⁺) to glucose-6-(monohydrogen)-phosphate forms the acid glucose-6-(dihydrogen)-phosphate and initiates a Proton Transport Chain. This flow can be directed, with HK forming complexes with either phospho-glucose isomerase (glycolysis) or glucose-6-phosphate dehydrogenase (Pentose Phosphate Pathway). The pH sensitivity of the phospho-glucose isomerase-catalysed reaction supports that the proton (H⁺) stays attached as glucose-6-(dihydrogen)-phosphate^[66]. The coenzyme of glucose-6-phosphate dehydrogenase (NADP⁺) is an ideal target to double the energy (particle H⁺) (NADPH-2H⁺) for forcing anabolic processes^[67]. Therefore, we count HK as the first enzyme of a Proton Transport Chain ending at the GAPDH-PH complex, generating the cytosolic lactate flow non-equilibrium (Pathway 3).



Pathway 3. Reductive branch of the glycolytic flow of protons. We divided the glycolytic metabolon into oxidative and reductive branches. We have replaced reversible reactions with a unidirectional flow, with phosphoric acid substituted by the acid anhydride ATP, and $+/-2H$ replaced by the $NAD^+/NADH-H^+$ system. The adjustments have been made in line with the law of the conservation of mass. Phosphorylation of glucose forms the acidic intermediate glucose-6-(dihydrogen)-phosphate and initiates the Proton Transport Chain. The acidic intermediate 3-glyceraldehyde (dihydrogen)-phosphate provides the proton to drive the Pyruvate Hydrogenase-catalysed reduction of pyruvate to lactate. Recovered NAD^+ returns to GAPDH (oxidative branch). The introduced substrate of the oxidative branch, 3-phosphoglycerate (monohydrogen)-phosphate, is given into the cytosolic pool of carboxylates.

This review started with the sentence “Every single chemical formula of modern models of glycolysis violates two laws of nature.”. The reaction catalysed by phosphofructokinase (PFK) liberates the second energy entity (H^+), theorized to combine with pyruvate for lactate biosynthesis. We propose triosephosphate isomerase as the partnering molecule of PFK. The transfer of one H^+ to triosephosphate isomerase is expected to drive the isomerase-catalysed reaction in a unidirectional manner to form 3-glyceraldehyde-(dihydrogen)-phosphate:



The transferred energy (particle H^+) is further given to the GAPDH-(NADH)-PH complex to generate the cytosolic lactate flow non-equilibrium (biosynthesis). 3-Glyceraldehyde-(monohydroxgen)-phosphate ($CHO-CHOH-CH_2-O-PO_3H^-$) is the end product of the reductive branch and is given into the cytosolic pool of carboxylates. The cytosolic pool of 3-glyceraldehyde-(monohydrogen)-phosphate was introduced as the primary substrate of the oxidative branch of the glycolytic metabolon.

Therefore, we suggest a PFK-triosephosphate isomerase complex pumps cytosolic dioxycetone-(monohydrogen)-phosphate into the reductive branch of the glycolytic metabolon. The energy particle finally fuses with pyruvate to produce the cytosolic lactate flow non-equilibrium.

This represents a provisional framework, quite possibly the initial instance where the distinct pathways of fermentation and glycolysis, cytosolic lactate flow non-equilibrium, and a scientific incorporation of ATP and NADH-H^+ in metabolism are presented. Should we reconsider our assertion that lactate is the primary step in returning Newton's apple to the tree? It is plausible that the initial step in creating an "apple" non-equilibrium with Earth is, perhaps, the introduction of metabolic pumping cytosolic dioxycetone-(monohydrogen)-phosphate into biosynthesis.

Discussion

The chemistry (Lavoisier) of glycolysis and the Citric Acid Cycle was ascertained before World War II started: Glucose is glycolytically split into lactic acid^[60]. The Citric Acid Cycle metabolically burns lactic acid to water and carbon dioxide^{[68][69][70]}. Physics (Newton) does not allow us to understand the state of knowledge pioneers reached. In order that the Citric Acid Cycle metabolically burns lactic acid to pyruvic acid (figure 1), glucose must be split to pyruvate, and pyruvate must be the substrate of glycolytic biosynthesis of lactate. Physics did not open the door to understand biosynthesis^[71]. Physics defined lactate/lactic acid as the end product of glycolysis/fermentation (waste) and not the first product of biosynthesis.

Since WW II, at least two fatal mistakes have guided the development of understanding metabolism. Firstly, Lardy and Ziegler investigated the ex vivo synthesis of phosphopyruvate from pyruvate^[72]. The authors presented biosynthesis as the reverse reaction of degradation by sorting the finding of phosphopyruvate synthesis into the scientific concept of glycolysis. Doing so, phospho(enol)pyruvic acid (glycolysis) was replaced by phosphopyruvate (synthesis). Deleting one H^+ from Scheme 1 violates the rule A. and M. Lavoisier formulated and provoked Meyerhof to comment on the article^[73]. The law of the conservation of mass, as well as published pioneer work, dictate opening a second, an independent pathway for carboxylates. H^+ is an experimentally determined glycolytic intermediate. We separated carboxylic acids (glycolysis, figure 5) from carboxylates (synthesis).

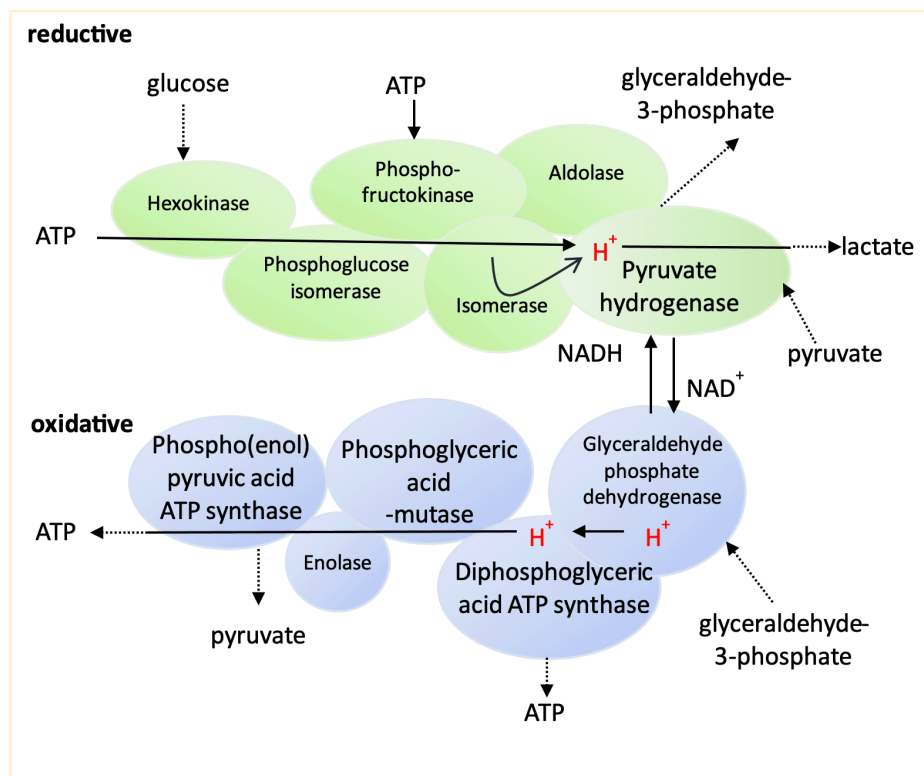


Figure 5. Glycolytic metabolon. The reductive branch (green) of the glycolytic metabolon wires the proton to lactate synthesis. The hexokinase-catalysed reaction starts the proton transport chain by transferring the proton of (ADP-H⁺) to form glucose-6-(dihydrogen)-phosphate. Energy (particle H⁺) guides the intermediates through the reductive branch. Pyruvate hydrogenase (PH) catalyses the ATP-driven biosynthesis of lactate from the glycolytic intermediate pyruvate. The oxidative branch (blue) starts with glyceraldehyde phosphate dehydrogenase (GAPDH)-catalysed metabolically burning of glyceraldehyde-3-(monohydrogen) phosphate to 1-(monohydrogen), 3-(dihydrogen)-phosphoglyceric acid. Burning of the aldehyde to a carboxylic acid group provides 2 H⁺. Firstly, the carboxylic acid group of 3-(monohydrogen)-phosphoglyceric acid; secondly, NADH-H⁺. The H⁺ of NADH-H⁺ is transferred to 3-(monohydrogen)-phosphoglyceric acid to form 3-(dihydrogen)-phosphoglyceric acid. The carboxylic acid group is capped by forming the acid anhydride 1-(monohydrogen), 3-(dihydrogen)-diphosphoglyceric acid. Diphosphoglyceric acid ATP synthase is suggested to catalyse two steps: firstly, the storage of the free H⁺ (dihydrogen-phospho-group) and secondly, the hydrolysis of the cap. Energy (particle H⁺) of the carboxylic acid group of 3-(monohydrogen)-phosphoglyceric acid guides the intermediates through the complex. Phospho(enol)pyruvic acid ATP synthase catalyses the final reaction of the oxidative branch: the transfer of wired H⁺ to temporarily more stable ATP.

Not opening an independent pathway for carboxylates (synthesis), the scientific concept of glycolysis (Scheme 1) transformed into an alchemistic illustration^[73].

Secondly, metabolism – a topic under investigation – has developed into a topic at school. Pupils learn the incorrect names of intermediates (carboxylates) and the incorrect names of enzymes like the alphabet, by rota. Consequently, glycolysis and the Krebs cycle have developed into the Tower of Babel. For example, the scientific concept of the Citric Acid Cycle presenting unidirectional burning of lactic acid to water and carbon dioxide (Figure 1) has mutated into an uncounted number of alchemistic illustrations. Carboxylates, such as citrate, oxaloacetate, and succinate, are illustrated under the title Citric Acid Cycle or TCA cycle. Wikipedia (German) propagates that H.A. Krebs discovered the citrate cycle and has understood synthesis as the reverse reaction of burning. The nomenclature of basic molecules has turned into a matter of discussion.

Our way out of chaos is referring to, citing, and working with the original publications. We learned this way during the preparation of Re-thinking the Citric Acid Cycle. G.A. Brooks' laboratory characterized the functional mitochondrial LDH-proton-linked MCT₁ complex. The complex

catalyses the metabolic burning of lactic acid to pyruvic acid. The reaction provides 2H^+ [11]. The complex catalyses the first chemical formula of the original Citric Acid Cycle[68][69][70]. Karpusas and co-workers analysed the structure of citrate synthase and found “The structure resembles a proposed transition state of the condensation reaction and suggests that the condensation reaction proceeds through a neutral enol rather than an enolate intermediate”[74]. We understood the sentence: oxaloacetic acid is a substrate of Citric Acid Synthase. The enzyme re-filling a citrate cycle was Pyruvate Carboxylase (PC). PC catalyses: $\text{pyruvate}^- + \text{HCO}_3^- + \text{ATP} \rightarrow \text{oxaloacetate}^{2-} + \text{ADP} + \text{P}$ (alchemy). We introduced the enzyme Pyruvic Acid Carboxylase (PAC). PAC catalyses: $\text{pyruvic acid} + \text{HCO}_3^- + \text{H}^+ (\text{ATP}) \rightarrow \text{oxaloacetic acid} + \text{ADP} + \text{P}$ (chemistry). Re-thinking the Citric Acid Cycle revealed the information that the hydrolysis of ATP provides one H^+ . Chemistry dictates that 1H^+ has to be given into the recovery of ATP: $\text{ADP} + \text{H}^+ + \text{HO-P} \rightarrow \text{ATP} + \text{H}_2\text{O}$.

This manuscript reviewed glycolysis and fermentation. We found that glycolytic enzymes are organized in the glycolytic metabolon. Enzyme complexes, such as the Citric Acid Cycle complexes and the Pyruvic Acid Dehydrogenase complex, unidirectionally catalyse acidic intermediates. Reviewing pioneer work revealed the names and chemical formulas of the acidic intermediates of the glycolytic metabolon. Following the flow of H^+ through the glycolytic metabolon allowed us to integrate ATP (H^+) and NADH- H^+ in the pathway of glycolysis and the pathway of fermentation. By following the flow of energy (particle H^+), we found the tentative 4th law of thermodynamics[75]. Transferring the tentative 4th law of thermodynamics to the subatomic level (H^+) of metabolism, this review changed the tentative law into the 4th law of thermodynamics.

Our reviews are the references for glycolysis, fermentation, and Citric Acid Cycles. Thus, scientists understanding carboxylates, such as 3-PG, 2-PG, or phospho(enol)pyruvate, as glycolytic intermediates violating the laws of nature and the original literature, will violate the most recently published literature and above all the ethics of science. The publishing policy of journals does not tolerate scientific misconduct, i.e., the violation of ethical principles in performing and publishing scientific research. Kennedy and Lehninger have experimentally determined glycolytically formed 3-phosphoglyceric acid. The Principle of Biochemistry (first edition) introduced 3-PG to explain glycolysis. The publishing policy of scientific journals cannot tolerate illustrations, discussions, and evaluations based on these falsifications and fabrications anymore, or?

Statements and Declarations

Author Contributions

Conceptualization, D.R. and G.S.C.; Writing—Original Draft Preparation, D.R.; Writing—Review and Editing, D.R. and G.S.C.

References

1. ^ΔFara P (1999). "Catch a Falling Apple: Isaac Newton and Myths of Genius." *Endeavour*. 23(4):167–70. doi:[10.1016/S0160-9327\(99\)80040-4](https://doi.org/10.1016/S0160-9327(99)80040-4).
2. ^Δ, ^Δ, ^ΔThomson W (1879). "The Sorting Demon of Maxwell 1." *Nature*. 20(501):126–126. doi:[10.1038/020126a0](https://doi.org/10.1038/020126a0).
3. ^ΔSheehan D (2020). "Maxwell Zombies: Mulling and Mauling the Second Law of Thermodynamics." *Journal of Scientific Exploration*. 34(3):513–36. doi:[10.31275/20201645](https://doi.org/10.31275/20201645).
4. ^ΔDavis TL (1924). "Neglected Evidence in the History of Phlogiston, Together with Observations on the Doctrine of Forms and the History of Alchemy." *Annals of Medical History*. 6(3):280–87.
5. ^Δ, ^Δ, ^Δ, ^Δ, ^Δ, ^Δ, ^Δ, ^Δ, ^Δ, ^Δ Meyerhof O (1951). "Mechanisms of Glycolysis and Fermentation." *Canadian Journal of Medical Sciences*. 29(2):63–77. doi:[10.1139/cjms51-008](https://doi.org/10.1139/cjms51-008).
6. ^Δ, ^ΔBruijne AW de, Vreeburg H, van Steveninck J (1985). "Alternative-Substrate Inhibition of L-Lactate Transport via the Monocarboxylate-Specific Carrier System in Human Erythrocytes." *Biochimica Et Biophysica Acta*. 812(3):841–44. doi:[10.1016/0005-2736\(85\)90280-9](https://doi.org/10.1016/0005-2736(85)90280-9).
7. ^Δ, ^ΔRoosterman D, Cottrell GS (2021a). "Rethinking the Citric Acid Cycle: Connecting Pyruvate Carboxylase and Citrate Synthase to the Flow of Energy and Material." *International Journal of Molecular Sciences*. 22(2):604. doi:[10.3390/ijms22020604](https://doi.org/10.3390/ijms22020604).

8. ^{a, b}Kurganov BI, Sugrobova NP, Mil'man LS (1985). "Supramolecular Organization of Glycolytic Enzymes." *Journal of Theoretical Biology*. **116**(4):509–26. doi:[10.1016/S0022-5193\(85\)80086-2](https://doi.org/10.1016/S0022-5193(85)80086-2).
9. ^ΔWu F, Minter S (2015). "Krebs Cycle Metabolon: Structural Evidence of Substrate Channeling Revealed by Cross-Linking and Mass Spectrometry." *Angew. Chem. Int. Ed.* **54**(6):1851–1854. doi:[10.1002/ange.201409336](https://doi.org/10.1002/ange.201409336).
10. ^ΔKrebs HA (1953). "The Nobel Prize in Physiology or Medicine 1953." NobelPrize.org. <https://www.nobelprize.org/prizes/medicine/1953/summary/>.
11. ^{a, b}Hashimoto T, Hussien R, Brooks GA (2006). "Colocalization of MCT1, CD147, and LDH in mitochondrial inner membrane of L6 muscle cells: evidence of a mitochondrial lactate oxidation complex." *Am. J. Physiol. Endocrinol. Metab.* **290**(6):E1237–1244. doi:[10.1152/ajpendo.00594.2005](https://doi.org/10.1152/ajpendo.00594.2005).
12. ^ΔDas ML, Koike M, Reed LJ (1961). "ON THE ROLE OF THIAMINE PYROPHOSPHATE IN OXIDATIVE DECARBOXYLATION OF α -KETO ACIDS*." *Proceedings of the National Academy of Sciences*. **47**(6):753–59. doi:[10.1073/pnas.47.6.753](https://doi.org/10.1073/pnas.47.6.753).
13. ^ΔReed LJ, Hackert ML (1990). "Structure-Function Relationships in Dihydrolipoamide Acyltransferases." *The Journal of Biological Chemistry*. **265**(16):8971–74.
14. ^{a, b}Krebs HA, Johnson WA (1937). "Metabolism of Ketonic Acids in Animal Tissues." *The Biochemical Journal*. **31**(4):645–60. doi:[10.1042/bj0310645](https://doi.org/10.1042/bj0310645).
15. ^{a, b}Krebs HA, Salvin E, Johnson WA (1938). "The Formation of Citric and Alpha-Ketoglutaric Acids in the Mammalian Body." *The Biochemical Journal*. **32**(1):113–17. doi:[10.1042/bj0320113](https://doi.org/10.1042/bj0320113).
16. ^ΔKarpusas M, Branchaud B, Remington SJ (1990). "Proposed Mechanism for the Condensation Reaction of Citrate Synthase: 1.9-A Structure of the Ternary Complex with Oxaloacetate and Carboxymethyl Coenzyme A." *Biochemistry*. **29**(9):2213–19.
17. ^{a, b}Roosterman D, Meyerhof W, Cottrell GS (2018). "Proton Transport Chains in Glucose Metabolism: Mind the Proton." *Frontiers in Neuroscience*. **12**:404. doi:[10.3389/fnins.2018.00404](https://doi.org/10.3389/fnins.2018.00404).
18. ^ΔMitchell P (1976). "Vectorial Chemistry and the Molecular Mechanics of Chemiosmotic Coupling: Power Transmission by Protonic Activity." *Biochem. Soc. Trans.* **4**(3):399–430. doi:[10.1042/bst0040399](https://doi.org/10.1042/bst0040399).
19. ^{a, b, c}Mitchell P (2011). "Chemiosmotic Coupling in Oxidative and Photosynthetic Phosphorylation. 1966." *Biochimica Et Biophysica Acta*. **1807**(12):1507–38. doi:[10.1016/j.bbabi.2011.09.018](https://doi.org/10.1016/j.bbabi.2011.09.018).
20. ^ΔNath S, Villadsen J (2015). "Oxidative phosphorylation revisited." *Biotechnol. Bioeng.* **112**(3):Art. no. 3. doi:[10.1002/bit.25492](https://doi.org/10.1002/bit.25492).
21. ^ΔJorgenson S (1999). "Tentative Fourth Law of Thermodynamics, Applied to Description of Ecosystem Development." *Annals of the New York Academy of Sciences*. <https://nyaspubs.onlinelibrary.wiley.com/doi/abs/10.1111/j.1749-6632.1999.tb10438.x>.
22. ^ΔLindskog S (1997). "Structure and Mechanism of Carbonic Anhydrase." *Pharmacology & Therapeutics*. **74**(1):1–20. doi:[10.1016/S0163-7258\(96\)00198-2](https://doi.org/10.1016/S0163-7258(96)00198-2).
23. ^ΔBecker HM, Klier M, Schüler C, McKenna R, Deitmer JW (2011). "Intramolecular Proton Shuttle Supports Not Only Catalytic but Also Noncatalytic Function of Carbonic Anhydrase II." *Proceedings of the National Academy of Sciences*. **108**(7):3071–76. doi:[10.1073/pnas.1014293108](https://doi.org/10.1073/pnas.1014293108).
24. ^{a, b}Roosterman D, Cottrell GS (2020). "Astrocytes and Neurons Communicate via a Monocarboxylic Acid Shuttle." *AIMS Neuroscience*. **7**(2):94–106. doi:[10.3934/Neuroscience.2020007](https://doi.org/10.3934/Neuroscience.2020007).
25. ^{a, b, c, d}Roosterman D, Cottrell GS (2021b). "The Two-Cell Model of Glucose Metabolism: A Hypothesis of Schizophrenia." *Molecular Psychiatry*. **26**(6):1738–47. doi:[10.1038/s41380-020-00980-4](https://doi.org/10.1038/s41380-020-00980-4).
26. ^ΔRoosterman D, Cottrell GS (2023). "Discovery of a Second Citric Acid Cycle Complex." *Heliyon*. **9**(5):e15968. doi:[10.1016/j.heliyon.2023.e15968](https://doi.org/10.1016/j.heliyon.2023.e15968).
27. ^ΔRogatzki MJ, Ferguson BS, Goodwin ML, Gladden LB (2015). "Lactate Is Always the End Product of Glycolysis." *Frontiers in Neuroscience*. **9**(February). doi:[10.3389/fnins.2015.00022](https://doi.org/10.3389/fnins.2015.00022).
28. ^{a, b}Green DE (1949). "ENZYMES IN TEAMS." *Scientific American*. **181**(3):48–51.
29. ^ΔKrebs HA (1953). "The Nobel Prize in Physiology or Medicine 1953." NobelPrize.Org. <https://www.nobelprize.org/prizes/medicine/1953/summary/>.
30. ^{a, b, c}Cori CF (1981). "The Glucose–Lactic Acid Cycle and Gluconeogenesis." In *Current Topics in Cellular Regulation*, 18:377–87. Elsevier. doi:[10.1016/B978-0-12-152818-8.50028-1](https://doi.org/10.1016/B978-0-12-152818-8.50028-1).
31. ^{a, b}Cori CF, Cori GT (1929). "GLYCOGEN FORMATION IN THE LIVER FROM D- AND L-LACTIC ACID." *Journal of Biological Chemistry*. **81**(2):389–403. doi:[10.1016/S0021-9258\(18\)83822-4](https://doi.org/10.1016/S0021-9258(18)83822-4).
32. ^{a, b}Langen P, Hucho F (2008). "Karl Lohmann and the Discovery of ATP." *Angewandte Chemie (International Ed. in English)*. **47**(10):1824–27. doi:[10.1002/ange.200702929](https://doi.org/10.1002/ange.200702929).

33. [△]Sun RC, Dukhande VV, Zhou Z, Young LEA, Emanuelle S, Brainson CF, Gentry MS (2019). "Nuclear Glycogenolysis Modulates Histone Acetylation in Human Non-Small Cell Lung Cancers." *Cell Metabolism*. 30(5):903–916.e7. doi:[10.1016/j.cmet.2019.08.014](https://doi.org/10.1016/j.cmet.2019.08.014).
34. [△]Gibbs ME (2016). "Role of Glycogenolysis in Memory and Learning: Regulation by Noradrenaline, Serotonin and ATP." *Frontiers in Integrative Neuroscience*. 9(January):70. doi:[10.3389/fnint.2015.00070](https://doi.org/10.3389/fnint.2015.00070).
35. [△]Zhang H, Liu J, Yang Z, Zeng L, Wei K, Zhu L, Tang L, et al. (2022). "TCR Activation Directly Stimulates PYGB-Dependent Glycogenolysis to Fuel the Early Recall Response in CD8+ Memory T Cells." *Molecular Cell*. 82(16):3077–3088.e6. doi:[10.1016/j.molcel.2022.06.002](https://doi.org/10.1016/j.molcel.2022.06.002).
36. [△]Sahlin K, Harris RC, Nylinde B, Hultman E (1976). "Lactate Content and pH in Muscle Samples Obtained after Dynamic Exercise." *Pflügers Archiv*. 367(2):143–49. doi:[10.1007/BF00585150](https://doi.org/10.1007/BF00585150).
37. [△]Liaw KY, Wei TC, Hsu SC, Lin JK (1985). "Effect of Severe Injury and Critical Illness on High-Energy Phosphates in Human Liver and Muscle." *The Journal of Trauma*. 25(7):628–33. doi:[10.1097/00005373-198507000-00009](https://doi.org/10.1097/00005373-198507000-00009).
38. [△]Tadi M, Allaman I, Lengacher S, Grenningloh G, Magistretti PJ (2015). "Learning-Induced Gene Expression in the Hippocampus Reveals a Role of Neuron–Astrocyte Metabolic Coupling in Long Term Memory." *PloS One*. 10(10):e0141568. doi:[10.1371/journal.pone.0141568](https://doi.org/10.1371/journal.pone.0141568).
39. [△]Hebb D (1949). *The Organization of Behavior; a Neuropsychological Theory*. <https://awspntest.apa.org/record/1950-02200-000>.
40. [△]Rowland LM, Pradhan S, Korenic S, Wijtenburg SA, Hong LE, Edden RA, Barker PB (2016). "Elevated Brain Lactate in Schizophrenia: A 7 T Magnetic Resonance Spectroscopy Study." *Translational Psychiatry*. 6(11):e967. doi:[10.1038/tp.2016.239](https://doi.org/10.1038/tp.2016.239).
41. [△]Meyerhof O (1922). "The Nobel Prize in Physiology or Medicine 1922." NobelPrize.Org. <https://www.nobelprize.org/prizes/medicine/1922/meyerhof/biographical/>.
42. [△][‡]Hui S, Ghergurovich JM, Morscher RJ, Jang C, Teng X, Lu W, Esparza LA, et al. (2017). "Glucose Feeds the TCA Cycle via Circulating Lactate." *Nature*. 551(7678):115–18. doi:[10.1038/nature24057](https://doi.org/10.1038/nature24057).
43. [△]Hashimoto T, Hussien R, Brooks GA (2006). "Colocalization of MCT1, CD147, and LDH in Mitochondrial Inner Membrane of L6 Muscle Cells: Evidence of a Mitochondrial Lactate Oxidation Complex." *American Journal of Physiology. Endocrinology and Metabolism*. 290(6):E1237–1244. doi:[10.1152/ajpendo.00594.2005](https://doi.org/10.1152/ajpendo.00594.2005).
44. [△]Bak LK, Schousboe A (2017). "Misconceptions Regarding Basic Thermodynamics and Enzyme Kinetics Have Led to Erroneous Conclusions Regarding the Metabolic Importance of Lactate Dehydrogenase Isoenzyme Expression." *Journal of Neuroscience Research*. 95(11):2098–2102. doi:[10.1002/jnr.23994](https://doi.org/10.1002/jnr.23994).
45. [△][‡]Szilard L (1929). "On the Decrease of Entropy in a Thermodynamic System by the Intervention of Intelligent Beings." *Z. Für Phys*. 53(11):840–856. doi:[10.1007/BF01341281](https://doi.org/10.1007/BF01341281).
46. [△]Schrödinger E (1944). *What Is Life?: With Mind and Matter and Autobiographical Sketches*. Canto Classics. Cambridge: Cambridge University Press. doi:[10.1017/CBO9781107295629](https://doi.org/10.1017/CBO9781107295629).
47. [△]Harden A, Young WJ, Martin CJ (1906). "The Alcoholic Ferment of Yeast-Juice." *Proceedings of the Royal Society of London. Series B, Containing Papers of a Biological Character*. 77(519):405–20. doi:[10.1098/rspb.1906.0029](https://doi.org/10.1098/rspb.1906.0029).
48. [△]Warburg O (1956). "On the Origin of Cancer Cells." *Science (New York, N.Y.)*. 123(3191):309–14. doi:[10.1126/science.123.3191.309](https://doi.org/10.1126/science.123.3191.309).
49. [△][‡][△]Kennedy EP, Lehninger AL (1949). "Oxidation of Fatty Acids and Tricarboxylic Acid Cycle Intermediates by Isolated Rat Liver Mitochondria." *The Journal of Biological Chemistry*. 179(2):957–72.
50. [△]Nelson DL, Cox MM (2012). *Lehninger Principles of Biochemistry*. 6th Edition. Macmillian Learning.
51. [△]Menard L, Maughan D, Vigoreaux J (2014). "The Structural and Functional Coordination of Glycolytic Enzymes in Muscle: Evidence of a Metabolon?" *Biology*. 3(3):623–44. doi:[10.3390/biology3030623](https://doi.org/10.3390/biology3030623).
52. [△]De BK, Kirtley ME (1977). "Interaction of Phosphoglycerate Kinase with Human Erythrocyte Membranes." *The Journal of Biological Chemistry*. 252(19):6715–20.
53. [△]Contreras-Baeza Y, Sandoval PY, Alarcón R, Galaz A, Cortés-Molina F, Alegría K, Baeza-Lehnert F, et al. (2019). "Monocarboxylate Transporter 4 (MCT4) Is a High Affinity Transporter Capable of Exporting Lactate in High-Lactate Microenvironments." *The Journal of Biological Chemistry*. 294(52):20135–47. doi:[10.1074/jbc.RA119.009093](https://doi.org/10.1074/jbc.RA119.009093).
54. [△]Dimmer KS, Friedrich B, Lang F, Deitmer JW, Bröer S (2000). "The Low-Affinity Monocarboxylate Transporter MCT4 Is Adapted to the Export of Lactate in Highly Glycolytic Cells." *The Biochemical Journal*.

55. ^ΔMeyerhof O (1927). "RECENT INVESTIGATIONS ON THE AEROBIC AND AN-AEROBIC METABOLISM OF CARBOHYDRATES." *The Journal of General Physiology*. 8(6):531–42. doi:[10.1085/jgp.8.6.531](https://doi.org/10.1085/jgp.8.6.531).
56. ^ΔGoldberg FW, Kettle JG, Lamont GM, Buttar D, Ting AKT, McGuire TM, Cook CR, et al. (2023). "Discovery of Clinical Candidate AZD0095, a Selective Inhibitor of Monocarboxylate Transporter 4 (MCT4) for Oncology." *Journal of Medicinal Chemistry*. 66(1):384–97. doi:[10.1021/acs.jmedchem.2c01342](https://doi.org/10.1021/acs.jmedchem.2c01342).
57. ^ΔLi M, Wu C, Yang Y, Zheng M, Yu S, Wang J, Chen L, Li H (2021). "3-Phosphoglycerate Dehydrogenase: A Potential Target for Cancer Treatment." *Cellular Oncology (Dordrecht, Netherlands)*. 44(3):541–56. doi:[10.1007/s13402-021-00599-9](https://doi.org/10.1007/s13402-021-00599-9).
58. ^ΔSadiqa A, Rasul A, Hassan M, Sultana S, Jabeen F (2022). "Identification of Novel Natural Inhibitors to Human 3-Phosphoglycerate Dehydrogenase (PHGDH) for Cancer Treatment." *Molecules (Basel, Switzerland)*. 27(18):6108. doi:[10.3390/molecules27186108](https://doi.org/10.3390/molecules27186108).
59. ^ΔWu YQ, Zhang CS, Xiong J, Cai DQ, Wang CZ, Wang Y, Liu YH, et al. (2023). "Low Glucose Metabolite 3-Phosphoglycerate Switches PHGDH from Serine Synthesis to P53 Activation to Control Cell Fate." *Cell Research*. 33(11):835–50. doi:[10.1038/s41422-023-00874-4](https://doi.org/10.1038/s41422-023-00874-4).
60. ^Δ^b ^ΔMeyerhof O (1951). "Mechanisms of glycolysis and fermentation." *Can. J. Med. Sci.* 29(2):Art. no. 2. doi:[10.1139/cjms51-008](https://doi.org/10.1139/cjms51-008).
61. ^ΔChaudhry R, Varacallo MA (2025). "Biochemistry, Glycolysis." In StatPearls. Treasure Island (FL): StatPearls Publishing. <http://www.ncbi.nlm.nih.gov/books/NBK482303/>.
62. ^Δ^bWatt IN, Montgomery MG, Runswick MJ, Leslie AGW, Walker JE (2010). "Bioenergetic Cost of Making an Adenosine Triphosphate Molecule in Animal Mitochondria." *Proceedings of the National Academy of Sciences of the United States of America*. 107(39):16823–27. doi:[10.1073/pnas.1011099107](https://doi.org/10.1073/pnas.1011099107).
63. ^ΔNath S (2016). "The Thermodynamic Efficiency of ATP Synthesis in Oxidative Phosphorylation." *Biophysical Chemistry*. 219(December):69–74. doi:[10.1016/j.bpc.2016.10.002](https://doi.org/10.1016/j.bpc.2016.10.002).
64. ^ΔNath S, Villadsen J (2015). "Oxidative Phosphorylation Revisited." *Biotechnology and Bioengineering*. 112(3):429–37. doi:[10.1002/bit.25492](https://doi.org/10.1002/bit.25492).
65. ^ΔSvedruzić ZM, Spivey HO (2006). "Interaction between Mammalian Glyceraldehyde-3-Phosphate Dehydrogenase and L-Lactate Dehydrogenase from Heart and Muscle." *Proteins*. 63(3):501–11. doi:[10.1002/prot.20862](https://doi.org/10.1002/prot.20862).
66. ^ΔDyson JE, Noltmann EA (1968). "The Effect of pH and Temperature on the Kinetic Parameters of Phosphoglucose Isomerase. Participation of Histidine and Lysine in a Proposed Dual Function Mechanism." PMID [5647261](https://pubmed.ncbi.nlm.nih.gov/5647261/).
67. ^ΔMullen AR, Wheaton WW, Jin ES, Chen PH, Sullivan LB, Cheng T, Yang Y, Linehan WM, Chandel NS, DeBerardinis RJ (2011). "Reductive Carboxylation Supports Growth in Tumour Cells with Defective Mitochondria." *Nature*. 481(7381):385–88. doi:[10.1038/nature10642](https://doi.org/10.1038/nature10642).
68. ^Δ^bKrebs HA, Johnson WA (1937). "Metabolism of ketonic acids in animal tissues." *Biochem. J.* 31(4):645–660. doi:[10.1042/bj0310645](https://doi.org/10.1042/bj0310645).
69. ^Δ^bKrebs HA, Salvin E, Johnson WA (1938). "The formation of citric and alpha-ketoglutaric acids in the mammalian body." *Biochem. J.* 32(1):113–117. doi:[10.1042/bj0320113](https://doi.org/10.1042/bj0320113).
70. ^Δ^bGreen DE (1949). "ENZYMES IN TEAMS." *Sci. Am.* 181(3):Art. no. 3.
71. ^ΔBak LK, Schousboe A (2017). "Misconceptions regarding basic thermodynamics and enzyme kinetics have led to erroneous conclusions regarding the metabolic importance of lactate dehydrogenase isoenzyme expression." *J. Neurosci. Res.* 95(11):Art. no. 11. doi:[10.1002/jnr.23994](https://doi.org/10.1002/jnr.23994).
72. ^ΔLardy HA, Ziegler JA (1945). "THE ENZYMATIC SYNTHESIS OF PHOSPHOPYRUVATE FROM PYRUVATE." *J. Biol. Chem.* 159(2):343–351. doi:[10.1016/S0021-9258\(19\)52795-8](https://doi.org/10.1016/S0021-9258(19)52795-8).
73. ^Δ^bMeyerhof O, Oesper P (1949). "The enzymatic equilibria of phospho(enol)pyruvate." *J. Biol. Chem.* 179(3):1371–1385.
74. ^ΔKarpusas M, Branchaud B, Remington SJ (1990). "Proposed mechanism for the condensation reaction of citrate synthase: 1.9-A structure of the ternary complex with oxaloacetate and carboxymethyl coenzyme A." *Biochemistry*. 29(9):2213–2219.
75. ^ΔJorgenson S (1999). "Tentative Fourth Law of Thermodynamics, Applied to Description of Ecosystem Development." *Annals of the New York Academy of Sciences*. <https://nyaspubs.onlinelibrary.wiley.com/doi/abs/10.1111/j.1749-6632.1999.tb10438.x>.

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